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STUDY THE PATHO-PHYSIOLOGICAL CHANGES AFTER INDUCE DIABETES IN MOUSE

Muna Sachit Hashim¹ AL-Aamery¹, Layla Hashim Alol² & Eman Hashim Yousif Ataee³ ^{1,3}Department of Pathology, College of Veterinary Medicine/University of Baghdad ²Department of Physiology, College of Veterinary Medicine/University of Baghdad

ABSTRACT

Diabetes mellitus induce by using alloxan which related to hyperglycemias' chemicals due to increase hepatic glucose. Alloxan consider as eta -cytotoxic that produced diabetes mellitus through targeting these cells of the pancreas by accumulates as glucose analogues, so causes low insulin release (Bhaskar, 2014). Diabetes is a chronic metabolite disorder conducts a major health concern today whose prevalence has continuously increased over worldwide in the past few decades. Diabetes mellitus consider as incurable metabolic disease that affects around 2.82% of the world people. Ankur (2012), there was poorly focusing in diabetes mellitus effects in different organs in mice such as skin, thymus, salivary gland and other organs, in spite of many studies were established in this concern. This study was designed to point the patho with physiological changes in different organs after induction diabetes in mice. The main histo-patho-physiological findings were elevated glucose in blood, fatty change, which appear as clear vacuoles of lipid pushed the nucleus a side fatty changes indicates the precipitation of lipid within tissues, evidence of necrosis and inflammatory infiltration in different organs, evidence of brain tissues damage, hyalinization of heart muscle, hyperkeratosis in esophagus and skin lesions as hair lost. Conclusion diabetes mellitus caused different pathophysiological changes in mice mainly increase glucose level in blood; appearance of fatty changes resulted from failure of liver to lipid metabolism. In addition, caused necrosis in pancreatic cell which responsible for insulin production and controlling the glucose and lipid in blood. Further to damage vital organs (brain and heart).

KEY WORDS: D.M. mouse; different organs.

INTRODUCTION

Alloxan is considered as toxic glucose analogue target to insulin-producing cells (beta cells) when injected to rodents. Insulin-dependent diabetes mellitus or D.M induced by alloxan is same to type I diabetes in humans^[1]. And work as specific toxicity for cells by accumulating within cytoplasm by increase uptakes GLUT² glucose transporter. That results in over producing of reactive oxygen species ROS associated to reduction of dialuric acid. These intracellular imbalances are initiates the toxicity to cells^[2]. Several experimental studies reveal that alloxan stimuli a sudden shock in insulin level in present or absent of glucose, which were occur yet alloxan injection^[3,4]. with full suppress to pancreatic cells response to glucose at low or high concentrates ^[5] Besides alloxan action on pancreas' beta cells which propose as the important mark to determine alloxan diabetogenic similar status occurs through different reducing agents as glutathione; cysteine; ascorbic acid; sulfhydryl (-SH) groups^[6,7]. Alloxan acts with two-SH groups in the sugarbinding site of glucokinase results on form hard bond and in active form of enzyme then reduction in dialuric acid with highly value of ROS. superoxide radicals^[8,9]. These superoxide's radicals liberates ferrous ions from ferritin^[10]. Superoxide's radicals undergo modification results in release hydrogen peroxide H₂O₂. These ROS causes effects on the Deoxyribo Nucleic Acid of pancreases' islets. DNA damage stimulate poly ADP-ribosylation in which DNA repair. Anti-oxidizing agents must be used ^[11] disturbances in intracellular calcium's ion homeostasis

were reported to be indicates induces' diabetes. Experimenters show that Calcium concentration become higher within beta cells of pancreases' islets^[12]. Calcium influxes are results from uncontrolled opens pores of calcium channels that's promoting calcium entrance in pancreatic cells. These high concentrations add to mentioned above were furthers contributes to over physiological insulin releasing cause obvious damage within pancreas' islets^[13]. This study was aimed to:

- 1- Pointed the physiological effects of D.M.
- 2- Reported the histopathological changes of different organs in mouse after induced diabetes mellitus.

MATERIALS & METHODS

Current experiment was conducted in 30-mixed white mice of uniform age (4week) at (18-24) gram body weight. They divided into sex groups of 5 animals to each (three groups were experimental and three groups were control); Group I was injected with alloxan for one week; Group II was treated for two weeks; Group III was for three weeks. Others three groups were controls besides each experiment groups and given normal saline or distilled water. Animals were housed in animals' house in Veterinary Medicine College of Baghdad University and given commercial-pellet feed, and tap water. Alloxan monohydrate (150 mg/kg) has been prepared in 10% saline solution and injected in mice intraperitoneal at single dose of (0.1ml/10 mg B.W) weekly for three weeks, Sugar level of these mice was test at 1-2-3 weeks for diabetes detection on blood from each mouse blood by glucometer (ACCU-CHEK, Germany). Animals have more than 200 mg/dL were diabetic ^[1,16]. Tissue samples from different organs were fixed with formalin (10%) processing and prepared slides done according to ^[17]. **Statistical analysis**

Test data depend on means \pm S.D. and analyzed according

RESULTS & DISCUSSION Estimation of Glucose level

Each mouse of each two groups submitted to test for glucose using glucometer on samples of blood (ACCU-CHEK, Germany). As shown in table (1).

to ANOVA and t test. P < 0.05 considered significant (18).

TABLE 1: showed results of mice blood glucose level, the estimation performed after alloxan injection as com	nparison to
control mice	

Animals groups	Time of treatment with Alloxan	Glucose level mg/dL(rang)
Group I / n=5	One week	80 -120 mg/DL***
Group II / n=5	Two week	125 -175 mg/dL**
Group III / n=5	Three week	200 -400 mg/dL*
Control / n=15	Isotonic solution only	70 -100 mg/dL

Significantly different from corresponding control at variance: *p < 0.05;** p < 0.01;*** p < 0.001

There was high glucose level after two week of alloxan injection with comparison to healthy mouse and there were significantly differences at p< 0.05 and these results are agree with ^[1,19]. Increase glucose level occurs due to failure of body function that's' associated to glucoses' and lipids' metabolism ^[20].

Histopathological Investigations Pancreas

Histopathological investigations on pancreatic section showed; inflammatory infiltration and necrosis in interlobular septa destructive change on epithelial lining cell with hemorrhage and congested blood vessel. As showed in figure (1-A; B).





FIGURE B: Pancreatic section of mouse showing necrosis within lobules (black arrow) vessels dilation ' with thrombi (red arrow) with hemorrhagic changes (blue arrow).

Pathological lesions in pancreases were severe necrotic changes after two weeks; increased eosinophilia (nuclear fragmentation), cellular infiltration in interlobular septa, hemorrhage, fibrous' proliferating in pancreatic lobules and pancreatic' acini disarrangement's', nuclear changes: karyohexis and fragments' then absent of nucleus and rarefies in nuclei on cells ^[19]. And degeneration islets of Langerhans are induced by alloxan were agreed with other authors findings in animals ^[14, 21, 22].

Liver

Liver tissues showed degenerative hepatocytes with hepatosis with fatty droplet as clear vacuoles; infiltration of kuffer cell; cloudy swelling and absence of sinusoids; hemorrhagic change with degenerative changes within hepatocytes including absence of nuclei, Figure (2-A; B). Histopathological findings in liver were agreed with (23). These liver changes are due to alloxan induced D.M ^[21; 24] in spite liver, insulin-dependence organ and play a vital role in Sugar and fat homoeostatic', is seriously affects' during diabetes^[25]. Causes' prominent alteration in the lipids' concentrating and compositing^[26]. Disturbance in sugar metabolizing so there was decreased glycolysis, stimuli glycogenesis and increased glucose-genesis in diabetics' liver^[27]. Other authors said that untreated diabetics' liver cause hyperglycemia which is known to activate isoforms of proteins' kinases C (PKC) in other tissue^[28] besides hepatocytes, PKC is major step for insulin transductions' pathway which stimulates PKC ^[29]. Leads to decreased apoptosis and hyperplasia that's termed diabetics' hepatomegaly ^[24].

multiple area of vacuolation of neuron due to

demyelination (fatty degeneration), Figure (3).

Brain

Brain and cerebellum showed widely speared degenerative changes and these appear as necrotic foci and there are

FIGURE 2A: Liver section of mouse showing hepatosis (black arrow)and fatty change(red arrow)and cloudy swelling(blue arrow) (H & E X 40).

B: Liver section of mouse showing hemorrhage (red arrow) with degenerative hepatocytes with fragmented nuclei (black arrow) with infiltration of kuffer cell (blue arrow). (H & E X40).



FIGURE 3A: Brain tissue of mouse showed: necrosis (black arrow) with neuronal vacuolation (red arrow) (H&E).X100. B: Brain tissue of mouse showed: necrotic foci arrow X 400.



FIGURE 4: Lung sections of mouse: A showing congestion (black arrow) and desquamation epithelium of bronchioles (red arrow) with emphysema (destruction of alveoli wall) (blue arrow) (H&E) X 400.
B: showing Interstitial Pneumonia as whole slide (H&E) 100X.

These changes in nervous system after alloxan injection, termed diabetics' neuropathy and affects: brain; spinals' cord and peripherals nerves ^[23, 30]. Diabetes initiates brain damage in experimental and clinical subjects, promotes neuronal damage; increases infracted postischaemic seizures^[31]. Diabetic neuropathy associated with excessive

production of sorbitol through reduction of maintaining hyperglycemia and related processes ^[32; 33].

Lung

Our investigations on lung showed evidence of Pneumonia represented by inflammatory infiltration type with bronchial wall damage; alveolus wall damage with heavy cellular debris and exudation, Figure (4). Changes in lung sections agreed with Author ^[21] who observed edema, collapse of alveoli, congestion and hemorrhage in lung sections of alloxan-induced diabetic dogs and agreed with author ^[34] in rabbit.

Gastrointestinal tract

Histopathological investigation on GIT and its glands showed fatty droplet with necrosis (Figure 5) and inflammation of Esophagus (Figure 6).



FIGURE 5A: G.I.T. section of mouse showing: marked fatty deposits (black arrow) with hydropic degeneration(red arrow)and necrosis (blue arrow) and mucinous degeneration(white arrow). (H & E) X100. **B:** Retropharyngeal salivary gland with thymus gland section of mouse showing: congestion with necrosis in salivary gland (black arrow). AS well as in thymus gland (red arrow). (H & E X 400).

Esophagus

Histopathological investigation on esophagus showed different pathological changes characterized by hyperkeratosis with calcification in the wall of esophagus accompanied with inflammatory responses mainly monocytes with fibrous connective tissues proliferation (Figure 6).



FIGURE 6A: Esophagus section of mouse showing: hyper keratinization (black arrow) with inflammation and thickening and increase proliferation of fibrous' connective tissue F.C.T (red arrow). (H&E) X100. **B:** Esophageal gland of mouse showed metaplasia with calcification and keratin debris (blue arrow)(H&E) X400.

These changes in G.I.T. resulted from disturbances in cellular homeostasis and vascular system in diabetes mellitus (35) no controlling stimulus of hypertrophy's' and hyperplasia's' and dysplasia and metaplasia of intestines' epithelium in diabetic animals are due to chronicity period accompanies with growth abnormalities ^[36]. Previous studies in experimental animals caused intestines' degeneration and necrosis on intestinal villi ^[21] and gained weight with high tissue waters' content^[37]

increases growth of yeasts on the GIT mucosa also caused permanents hyperglycemia in diabetic rabbits. Hyperglycemia in D.M responsible for hurt tissues injuries [38].

Heart and Spleen

Pathologic investigations' on heart and spleen showed inflammatory reaction and fat deposition with hemorrhage Figure (7).



FIGURE 7A: Heart section of mouse showing congestion (black arrow); myofibrils hyalinization (red arrow) ; inflammatory cells (blue arrow) (H & E) X 400.

B: Spleen section of mouse showing distractive change and hemorrhage (black arrow) with evidence of fat deposition (red arrow) (H & E) X 400.

Changes in heart are agreed with author (34) who found edema, histiocyte proliferation and myocarditis in heart sections in diabetic rabbits.

Kidney

Sections of kidneys showed, Nephritis with hyaline and hemorrhagic cast with atrophied glomeruli and necrosis of renal tubules (Figure 8).



FIGURE 8: Kidney section showing: A: Nephritis with hyaline and hemorrhagic cast (black arrow)and glomeruli necrosis (red arrow).X400

B: Atrophic glomeruli (black arrow) with necrotic proximal tubules (red arrow). (H&E) X400.



FIGURE 9: Testis section showing: A: empty seminiferous tubules from loss of spermatogenesis (black arrow) with congestion (red arrow) and fatty droplet within cell of epithelial lining.(blue arrow)(H & E X400).B: empty seminiferous tubules from loss of spermatogenesis (arrow))(H & E X100).

Our changes in kidneys were agreed with author^[23] and they are attributed to defecated metabolism^[39] in hyperglycemic or diabetic animals^[22, 40] other authors showed proliferations' cell changes and nephrons hypertrophy in diabetics' rabbits. In diabetics' dogs there are degenerating glomeruli with necrotic tubules' epithelium existence of hyaline casts^[21] several researchers showed glomerular nephropathy and sever interstitial changes ^[39,41] and others showed that controlling metabolism is essential for reduction of nephropathy in diabetes, and when there is prolonged diabetes' duration and no normoglycaemia reinstitutions' so nephropathy is difficult reverse to normal status ^[42, 43].

Testes

Tissues' of testes showed failure of spermatogenesis with inflammation represented by infiltration of MNCs enter and outer seminiferous' tubules with proliferation and thickening of fibrous connective tissue septa with evidences of congested bloods' vessels; necrosis within lumen of tubules.(Figure 9). These results on testicular tissue of mice after alloxan uses are agree with authors ^[44, 45].

Skin

Skin tissues after alloxan injected mouse showed loss of hair follicles with hyperplasia of collagen fibers and fibrous connective tissue F.C.T. (Figure 10).



FIGURE 1: Skin section showing A: aborted hair follicles (black arrow) and inflammatory response (red arrow) increase F.C.T thickness. (H & E X100).

B: atrophied sebaceous and sweet gland (blue arrow); increase F.C.T thickness (red arrow) (H &E ×400).

REFERENCES

- Bhaskar Sharma, Mohd. Sufiyan iddiqui, Gurudayal Ram, Ranjeet Kumar Yadav, Arti Kumari, Gaurav Sharma, and Nakuleshwar Dut Jasuja⁽²⁰¹⁴⁾. Rejuvenating of Kidney Tissues on Alloxan Induced Diabetic. Advances in Pharmaceutics; Volume (2014), Article ID 439158,9 pages. http:// dx. doi.org /10. 11 55 /2014/439158.
- [2]. Ankur Rohilla, Shahjad Ali (2012) Alloxan Induced Diabetes: Mechanisms and Effects Department of Pharmaceutical Sciences, Shri Gopi Chand Group of Institutions, Baghpat, Uttar Pradesh, India. Vol. 3 (2)Apr–June 2012 International Journal of Research in Pharmaceutical and Biomedical Sciences. ISSN: 2229-3701. www.ijrpbsonline.com 819.
- [3]. Szkudelski, T., Kandulska, K., Okulicz, M. (1998) Alloxan *in vivo* does not only exert deleterious effects on pancreatic B cells. Physiol. Res.;47:343– 346.[PubMed]
- [4]. Lachin, T., Reza, H. (2012) Anti diabetic effect of cherries in alloxan induced diabetic rats.Recent Pat Endocr Metab Immune Drug Discov; 6:67-72.
- [5]. Kliber, A., Szkudelski, T., Chichlowska, J. (1996) Alloxan stimulation and subsequent inhibition of insulin release from in situ perfused rat pancreas. J Physiol Pharmacol; 47:321-8.
- [6]. Lenzen, S. (2008) The mechanisms of alloxan and streptozotocin-induced diabetes. Diabetologia; 51:216-26.

- [7]. Zhang, X., Liang, W., Mao, Y., Li, H., Yang, Y., Tan, H. (1992) Hepatic glucokinase activity is the primary defect in alloxan-induced diabetes of mice. Biomed Pharmacother
- [8]. Munday, R. Dialuric acid autoxidation (1988) Effects of transition metals on the reaction rate and on the generation of reactive oxygen species. Biochem Pharmacol; 37:409-13.
- [9]. Das, J., Vasan, V., Sil, P.C. (2012) Taurine exerts hypoglycemic effect in alloxan-induced diabetic rats, improves insulin-mediated glucose transport signaling pathway in heart and ameliorates cardiac oxidative stress and apoptosis. Toxicol. Appl. Pharmacol; 258:296-308.
- [10]. Sakurai, K., Ogiso, T. (1995) Effect of ferritin on DNA strand breaks in the reaction system of alloxan plus NADPH cytochrome P450 reductase: ferritin's role in diabetogenic action of alloxan. Biol Pharm Bull; 18:262-6.
- [11]. Ebelt, H., Peschke, D., Bromme, H.J., Morke, W., Blume, R., Peschke, E. (2000) Influence of melatonin on free radical-induced changes in rat pancreatic beta-cells in vitro. J Pineal Res; 28: 65-72.
- [12]. Park, B.H., Rho, H.W., Park, J.W., Cho, C.G., Kim, J.S., Chung, H.T., Kim, H.R. (1995) Protective mechanism of glucose against alloxan induced pancreatic beta-cell damage. Biochem Biophys Res Commun., 210:1-6.

- [13]. Etuk, E.U. (2010) Animals models for studying diabetes mellitus. Agric Biol J N Am; 1:130-4.
- [14]. Szkudelski, T. (2001)The Mechanism of Alloxan and Streptozotocin Action in B Cells of the Rat Pancreas Physiol Res., 50:536-46.
- [15]. Lenzen, S. (2008) The Mechanisms of Alloxan- and Streptozotocin-induced Diabetes". Diabetologia 51
 (2): 216–226. doi:10.1007/s00125-007-0886-7. PMID 18087688.
- [16]. Zuo-fa Zhang , Guo-ying, Lv, Lian-gen Shi, Huijuan Pan, yong-zhi Wu &Lei-fa Fan(2009) Oxyresveratrol from *Ramulus mori* attenuatesalloxan-induced liver damage in mice.Institute of Horticulture, Zhejiang Academy of Agricultural Science, Hangzhou 310021, China College of Animal Sciences, Zhejiang University, Hangzhou 310029, China 978-1-4244-2902-8/09/\$ 25.00 ©2009 IEEE.
- [17]. Kiernan, J.A. (2012) Histological and Histochemical methods. Theory and practice 4th ed. Pp: 334-337 Boxham, UK: scion.
- [18]. Mélisande Albert, Yann Bouret, Magalie Fromont and Patricia Reynaud-Bouret (2015) Bootstrap and permutation tests of independence for point processes. arXiv: [math.ST] 13 Jan. 1406.1643v3.
- [19]. Rerup, C.C. (1970) Drugs producing diabetes through damage of insulin secreting cells. Pharmacol. Rev.; 22:485–518.[PubMed].
- [20]. T. Szkudlski, K. Kandulska, M. Okulicz, Dep. of Agriculture, Poznan, Poland. (1998). Alloxan in vivo Dose Not only Exert Deleterious Effects on Pancreatic B cell.Physiol.Res.47:343-346.ISSN 0862-8408.
- [21]. Sandu, K., Randhawa, S.S., Brar R.S. (2000) Clinical and pathological Changes in alloxan induced diabetes mellitus alone and in combination with ethylene glycol induced nephropathy in dogs. Indian J. Vet. Pathol. 24, 12–15.
- [22]. Mir, S.H., Baqui, A., Darzi, M.M., Mir, M.S. (2006) Pathoanatomy of experimental diabetes in rabbits (*Oryctolagus cuniculus*) Oriental Sci.; 11:69–72.
- [23]. Masood Saleem Mir, Mohammad Maqbool Darzi, Hilal Musadiq Khan, Shayaib Ahmad Kamil, Asif Hassan Sofi C, Sarfraz Ahmad Wani (2013) Pathomorphological effects of Alloxan induced acute hypoglycaemia in rabbits. a Division of Veterinary Pathology, F.V.Sc. & A.H., Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Shuhama, Alusteng, 190 006, Kashmir (J&K), India.
- [24]. Herrman, C.E., Sanders, R.A., Klaunig, J.E., Shwarz, L.R., Watkins, J.B. (1999) Decreased apoptosis as a mechanism for hepatomegaly in streptozotocin-induced diabetic rats. Toxicol. Sci.;50:146–151.[PubMed]
- [25]. Seifter, S., England, S. (1982) Energy metabolism. In: Arias I, Popper H, Schacter D, et al., editors. The Liver: Biology and Pathobiology. New York: Rauen Press; pp. 219–249.
- [26]. Sochor, M., Baquer, N.Z., McLean, P. (1985) Glucose over and under utilization in diabetes: comparative studies on the change in activities of

enzymes of glucose metabolism in rat kidney and liver. Mol. Physiol.; 7:51-68.

- [27]. Baquer, N.Z. Glucose over utilization and under utilization in diabetes and effects of antidiabetic compounds. 1998 An. Real. Acad. Farm.; 64:147– 180.
- [28]. Porte, D., Schwartz, M.W. (1996) Diabetes complications: why is glucose potentially toxic? Science.; 272:699–700.[PubMed].
- [29]. Adachi, T., Nakashima, S., Saji, S., Nakamura, T., Nozawa, Y. (1996) Mitogen-activated protein kinase activations in hepatocyte growth factor stimulated rat hepatocytes: Involvement of protein tyrosine kinase and protein kinase C. Hepatology.; 23:1244–1253.[PubMed].
- [30]. Gallego, M., Setien, R., Izquierdo, M.J., Casis, O.C.E. (2003) Diabetes-induced biochemical changes in central and peripheral catecholaminergic systems. Physiol. Res. 52, 735–741.
- [31]. Muranyi, M., Fujioka, M., HE, Q., Han, A., Yong, G., Csiszar, K., Li Ping, A.N. (2003) Diabetes activates cell death pathway after transient focal cerebral ischemia. Diabetes 52, 481–486.
- [32]. Greene, D., Lattimer, S., Sina, A. (1987) Sorbitol, phosphoinositidesand sodium-potassium-ATPase in the pathogenesis ofdiabetic complications. N. Eng. J. Med. 316, 599–606.
- [33]. Tomlinson, K.C., Gardiner, S.M., Hebden, A., Bennett, T. (1992) Functional consequences of streptozotocin-induced diabetesmellitus, with particular reference to the cardiovascular system. Pharmacol. Rev. 44, 103–150.
- [34]. Sajad Hussain Mir and Mohd Maqbool Darzi (2009) Histopathological abnormalities of prolonged alloxan-induceddiabetes mellitus in rabbits Int. J. Exp. Path. (2009), 90, 66–73.
- [35]. Brownlee, M. (2001) Biochemistry and molecular cell biology of diabetic complications. Nature 414, 813–82.
- [36]. Younoszai M.K., Parekh V.V., Hoffman J.L. (1993) Polyaminesand intestinal epithelial hyperplasia in streptozotocin-diabeticrats. Proc. Soc. Exp. Biol. Med. 202, 206–211.
- [37]. Schedl, H.P. & Wilson H.D. (2005) Effect of diabetes on intestinal growth in the rat. J. Exp. Zool. 176, 487–495.
- [38]. McMohan M.M. & Bistrain B.R. (1995) Host defenses and susceptibility to infection in patients with diabetes mellitus. Infet. Am. 9, 1–9.
- [39]. Rasch, R. (1979) Prevention of diabetic glomerulopathy in streptozotocin diabetic rats by insulin treatment Glomerular basement membrane thickness. Diabetologia 16, 319–324.
- [40]. Bulut, H.E., Onarlioglu, B., Kaloglu, C., Ozdemir, O., Ayan, S. (2001) Effects of experimental diabetes an insulin treatment on rabbit renal morphology: a quantitative and qualitative study. Turk. J. Med. Sci. 31 209–216.
- [41]. Hirose, K., Qsterby, R., Nozawa, M. (1982) Development of glomerular lesions in experimental long-term diabetes in the rat. Kidney Int. 21, 689– 695.

- [42]. Renu, A., Saiyada, N.A., Odenbach, S. (2004) Effect of reinstitution of good metabolic control on oxidative stress in kidney of diabetic rats. J. Diabetes Complicat. 5, 282–288.
- [43]. Floretto, P., Steffes, M.W., Sutherland, E.R.D., Goetz, C.F., Mauer, M. (1998) Reversal of lesions of diabetic nephropathy after pancreas transplantation. N. Eng. J. Med. 339, 69–75.
- [44]. Ramin Jahangirfard and Sajjad Hejazi (2015). Antidiabetic effect of Coenzyme Q 10 on testis tissue in Alloxan induced Rats .Adv. Biores., Vol 6[1] January 2015: 54-61. DOI: 10.15515/abr.0976-4585.6.1.5461. *Corresponding author's Email: sajjad .hejazi @iaut.ac.ir
- [45]. Baris Atalay Uslu, Fatma IIlan; Fetih Gulyuzf,Mehmet Karaca and Ahmet Cihat Oner

(2009) Assessment of the Histopathological Changes Occurribg in the Testis of the Mice Suffering from Experimental Diabetes Induced Using Alloxan.Journal of Animal and Veterinary Advances 8(10).1929-1935,2009

- [46]. Flávio Pola dos Reis; Angelo Sementilli; Antonio Ricardo de Toledo Gagliardi.(2013). Experimental diabetes exacerbates skin transplant rejection in rats-Acta Cir. Bras. vol.28 no.5 São Paulo May 2013.
- [47]. Siamak Kazemi-Darabadi; Farshid Sarrafzadeh-Rezaei; Amir-Abbas Farshid, and Reza Baradar-Jalili.(2013) Healing of excisional wound in alloxan induced diabetic sheep: A planimetric and histopathologic study. Vet Res Forum. 2013 summer; 4(3): 149–155.