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Review Article

# ENTOMOPATHOGENIC POTENTIAL OF THE FUNGI *METARHIZIUM* (HYPOCREALES, CLAVICIPITACEAE) IN THE MANAGEMENT OF INSECT PESTS

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#### **ABSTRACT**

Metarhizium, a genus of entomopathogenic fungi belongs to Class Sordarimomycetes, is a soil-borne fungus and found in almost all habitats. Its action is epizootic in nature and does not hold any unwanted threat for humans or animals, as it a natural biocontrol agent and attacks only insects. Conservation biological control can be defined as manipulation of the environment to favor natural enemies, either by removing or mitigating adverse factors or by providing lacking requisites. Conservation of natural enemies can also be understood as one of the oldest form of biological control. Natural enemies, if established successfully, provide eco-friendly management of insect pests, which in turn reduces human and environmental burdens. Taking in to consideration the effective utilization of entompathogenic fungi in particular the Metarhizium sp. as an effective biocontrol agent in control of number of insect pests of agricultural and horticultural importance, the documentation of this fungal pathogen is important. In this review paper more than 150 insect species belonging to 11 different orders getting infected by Metarhizium sp. are detailed along with their classification.

KEY WORDS: Metarhizium, Hypocreales, Entomopathogenic Fungi, Biocontrol.

#### INTRODUCTION

Insects are the major culprits for considerable losses of wood in forest, crops, stored grains, textile industries, furnished wood etc. They affect the wood quality and thereby affect the revenue that also leads to accidents when it comes to building, bridges, rail tracks, where sleepers are made up of wood. Though synthetic insecticides/pesticides that are effective for small period of time are readily available commercially, their applications have serious consequences. Due to their broad spectrum nature, pesticides not only affect the targeted pest but also affect the beneficial organisms. Moreover, excessive and indiscriminate use of pesticides contaminates the soil, water and land. Pesticides have terrible adverse impact on humans, animals and the environment due to their toxic effects in food chain, development of resistance in target pest and destruction of natural enemies (Paray and Rajabalee, 1997; Joshi et al., 2000). Increased environmental awareness and concerns about the adverse effect of pesticides has made it imperative to identify safer and effective alternatives to the agrochemicals (Logan et al., 1990). Due to the ill effects of pesticides, alternative eco-friendly strategies are being increasingly appreciated and practiced for pest management (Padmaja et al., 2005). During the last decade, there has been increased and continued interest in the employment of natural biological enemies for the management of forest pests (Sandhu et al., 2012). Ecologically compatible strategies were developed to manage insect pest populations by utilizing entomopathogenic virus, fungi, nematodes, protozoa and bacteria as biological control agents (Castillo

et al., 2000; Lacey et al 2001). Once established, natural enemies has the capacity of self-replicating and surviving, which is one of the advantages of using biological control over synthetic pest control. Moreover, it reduces the excessive use of pesticides, hence reducing the risk to environment. Interestingly, biological control can even work with other pest management practices in an integrated pest management programme (IPM). Fungal entomopathogens with several life history specializations makes them a distinctive group from other types of pathogens. One of the important characteristics of entomopathogens is that they infect through the cuticle of their host (Hajek and Leger, 1994), usually not having to be ingested to cause infection. Deuteromycetes fungi that have been shown good potential as pest control agents for commercial production includes B. bassiana, Metarhizium anisoplaie, Verticillium lecanii and Trichiderma viridae. Metarhizium is known for its epizootic action and it also has been the subject of several molecular studies on fundamental topics such as virulence and host specificity. They can be grown on different artificial and natural media for field tests and application. High virulence, broad host range, quick mode of action, stability in culture, safety to workers, easy production and storage method are the essential characteristics required for a fungus to be used as an effective microbial insecticide (Mc Coy, 1990). Metarhizium anisopliae is an important entomopathogenic fungus, found in soils throughout the world. Fungi under the genus Metarhizium have been isolated from infected insects and soil of all continents, and there have been isolations

from near the Antarctic Circle also (Roddam and Rath, 1997). Although some isolates of these fungi have rather restricted host range, the group is better known for its ability to kill a wide spectrum of insects of at least seven orders (Veen, 1968) Four groups of insect pests (termites, locusts, spittlebugs and beetles) were previously targeted using M. anisopliae (Zimmermann, 1993). The common name for Metarhizium-induced disease is green muscardine, based on the encrustation of insect cadavers with green conidia. Metarhizium anisopliae was the first fungus worldwide to be mass produced and utilized for insect-pest control (Krassilstschik, 1888; Steinhaus, 1975). In the last few decades, many attempts with varying levels of success were made by utilizing *Metarhizium* sp. for pest control. In this paper. We attempt to review the effects of Metarhzium sp. on some major forest pests in Indian and international contexts. Further, considering the magnitude of risk factors associated with the prolonged use of commercially available insecticides, integrated pest management (IPM) methods have been suggested which have minimum side effects.

#### **ISOLATION**

Metarhizium is a soil-borne fungus found in almost all the habitats. The fungus can be isolated from soil samples, collected with the help of soil core borer or core sampler to the depth of 20 cm. The highly preferred method is insect bait method (Zimmermann, 1993). In this method the soil samples collected in clean polythene bags separately to be stored in refrigerated condition at 5 C. The samples should be subjected to insect bait method with in five to seven days by using the wax moth larvae Galleria mellonella as insect bait. The soil samples collected and kept in a polythene covers is to be moistened and filled in small 100 ml plastic containers and to release 10-15 grown up third instar larvae of G. mellonella in to the soil filled plastic containers for incubation for 14 days. Every day the soil has to be agitated to ensure that the larvae remain exposed to the soil. Diseased and mummified larvae should be collected from the container for isolation of the entomopathogenic fungi. If no external fungal sporulation is seen, the dead larvae will be surface sterilized by submersion in 1% sodium hypochlorite, followed by rinsing with sterilized water and allowed for sporulation in a moistened Petri plates. The fungi isolated from the cadavers are to be subcultured in Sabouraud dextrose agar medium (SDA). The fungi isolated from the artificial media will be again subcultured until the pure cultre is obtained (Raja Rishi et al. 2013).

In another method *Metarhizium* can also be isolated from roots. In this the roots were washed with sterile distilled water to remove excess soil. Soil that adhered closely to the root was kept as representative of the rhizosphere. The roots were cut into 0.5 cm pieces, placed in 5 ml distilled water and homogenized using a rotary homogenizer (Greiner Scientific). Samples (100 ml) of homogenate were spread, in duplicate, onto selective media, containing 39 g potatodextrose agar (PDA) 121 (Difco), 0.5 g cycloheximide 121, 0.2 g chloramphenicol 121, 0.5 g 65% dodine 121 and 0.01 g crystal violet 121. The plates were incubated at 27 °C for 20 days. Metarhizium isolates were identified by colony

morphology, namely white mycelia with green conidia, as well as microscopic identification of conidial morphology. Morphologically differing colonies were individually isolated from the selective plate of each plant root sample and grown on PDA plates at 27 °C for 10 days. Morphologically similar colonies were also isolated multiple times from the same plant root sample (Michael Wyrebek *et al.* 2011). Metarhizium fungal pathogens were also isolated directely from the naturally infected cadavers by incubating or streaking a sub-sample on surface of selective media (potato dextrose agar, PDA, Criterion, USA with 0.1% chloramphenicol and 0.05% cyclohexamind) (Tangthira sunun *et al.* 2010).

In another process of isolation, the soil samples were processed by diluting 10 g in 100 ml of sterile water that was added 0.01% Tween-80. One milliliter of the soil dilution was spread over those selective media. Then, single colony of *Metarhizium* spp. was transferred to PDA for purification. All were propagated and maintained with a 5 mm diameter mycelial plug taken from the growing edge of a 7 days old culture grown on PDA plates for 28 days at room temperature. Pure culture can be obtained and the identification has to be done by observation of conidia, colony and mycelia (Tangthirasunun *et al.* 2010).

#### SUCCESS STORIES

Metarhizium was tested against almost 150 species of insect pests in about more than 200 experiments on insects of different orders namely Coleoptera, Lepidoptera, Blattoidea, Isoptera, Hemiptera, Orthoptera, Diptera, Hymenoptera, Thysanoptera, Neoptera and Trombidiformes. Review of the literature reveals that Metarhizium has been most effectively and extensively used for the order Coleoptera. More than 50 species under the order Coleopters were studied and most of them were found to be susceptible to attack by Metarhizium. The second most studied order was Lepidoptera. More than 25 different species belonging to order Lepidoptera were recorded to be attacked by Metarhizium, in more than 30 separate studies. Hemiptera was the third extensively studied order where more than 20 experiments have been conducted. Isoptera was the fourth extensively studied order which included 16 species with more than 30 cases. Ten species of Blattoidea were recorded in more than 40 studies, both in lab as well as in field conditions. In addition to these orders, Orthoptera, Diptera, Hymenoptera, Thysanoptera, Neoptera and Trombidiformes were also observed to be attacked by Metarhzium. Under the order Coleoptera Metarizium was found to be most effective against members of the family Curculionidae. Sixteen species of this family were infected by Metarizium, followed by Cerambycidae, Scarabaeidae and Chrysomelidae. Interestingly, among different stages, larval stages were most prone to get attacked. Results varied from species to species where mortality percentage varied from a minimum of 15% to a maximum of 100%. The underlying reasons for this level of variation could be attributed to the climatic factors that influence the virulence of the fungus, and the differences in the susceptibility of the insects and there stages (Table-1).

TABLE 1. Pathogenicity of Metarhizium against forest and crop pests

S.No	Target organism	Order	Family	Stage	Effect	Reference
2.	Adoryphorus couloni Aeolesthes sarta	Coleoptera Coleoptera	Scarabaeidae Cerambycidae	Grub	90-100% mortality 50.99% mortality	Brownbridge <i>et al.</i> , 2009 Farashiani <i>et al.</i> , 2008
					47.50% mortality	Mohi-Uddin et al., 2009
ώ	Agrilus auriventris	Coleoptera	Buprestidae	Grub	72.7-94.8% mortality 59.7% mortality	Fan <i>et al.</i> , 1990 Fan <i>et al.</i> , 1988
4.	Amrasca devastans	Hemiptera	Cicadellidae	Larva	$9x10^9$ cfu/g @ 2500 g/ha produced 12.3 % Mortality in field	Neelima <i>et al.</i> , 2011
5.	Anomala cuprea	Coleoptera	Scarabaeidae	Grub	Pathogenic	Shimazu et al., 1993
6.	Anopheles gambiae	Diptera	Culicidae	Adult	Susceptiblity to the funguswas more with mineral oil formulation applied to polyester net than pyrethroid	Howard, 2010
7.	Anoplophora glabripennis	Coleoptera	Cerambycidae	Larva Adult	Pathogenic Shortened longevity and decreased oviposition. Percentages of eggs that did not hatch were greater in fungal-treated females and 60% of unhatched eggs contained signs of fungal infection	Wang <i>et al.</i> , 2010 Hajek <i>et al.</i> , 2008
				Adult	Beetles in two density treatments died in fewer days than beetles exposed to environments without conidia.	Shanley and Hajek, 2008
				Adult	Numbers of adult beetles from plots of each fungal species died in <10 days were greater than controls	Hajek <i>et al.</i> , 2006
					was 5 days.	
				Grub and adult	Decreased ALB longevity and fitness in two application methods. Longer activity of fungus in cages treated with fungal bands compared with sprays.	Hajek <i>et al.</i> , 2007
∞ ∞	Aphis craccivora	Hemiptera	Aphididae	Adult	Pathogenic with a varying degree of infectivity.	Pegu <i>et al.</i> , 2013
9. 10.	Aphis gossypii Apriona germari	Hemiptera Colelptera	Aphididae Cerambycidae	Adult Grub	Pathogenic with a varying degree of infectivity. Cumulative corrected mortality reached to 89.47% and 88.92% after 14 d ay treatment.	Pegu et al., 2013 Chen DeLan, 2013
11.	Apriona rugicollis	Coleoptera	Cerambycidae	Grub	Produced 81.2% mortallity	Fan <i>et al</i> .,1988
12. 13.	Atta bisphaerica Atta sexdens piriventris	Hymenoptera Hymenoptera	Formicidae Formicidae	Adult Adult	More than 80% mortality recorded Among the strains E <sub>6</sub> , Al Rj and E <sub>sa</sub> , E <sub>sa</sub> were most	Castilho <i>et al.</i> , 2010 Silvaand Diehl,1988
14.	Atta sexdens rubropilosa	Hymenoptera	Formicidae	Adult	More than 80% of mortality recorded	Castilho et al., 2010
15. 16.	Atta sexdens sexdens Batocera horsfieldi	Hymenoptera Coleoptera	Formicidae <i>Cerambycidae</i>	Adult Grub	Above 80% mortality obtained 78.3 % mortality recorded	Loureiro <i>et al.</i> , 2005 Fan <i>et al.</i> , 1988
17.	Bemisia tabaci	Hemiptera	Aleyrodidae	Maggot Maggot	Effective against nymphs  Pathogenic with varying degree of infectivity	Azevedo et al., 2005

Delate et al., 1995 Wells et al., 1995 Wells et al., 2007 Maketon et al., 2009 oil Ashraf et al., 2009	LT <sub>50</sub> was comparatively more (93-175 hrs) in soil	Adult				
Delate et al., 1995 Wells et al., 1995 Maketon et al. 20	LT <sub>50</sub> were comparatively more (65-106 h) in soil than on filter paper (50-83 h).	Adult	Rhinotermitidae	Isoptera	Coptotermes heimi	36.
Delate <i>et al.</i> 199	100% fermite mortality after 1 month	Adult	Rhinotermitidae	Riattodea	Contotormos nostroi	3
Chai 1005	100% mortility recorded  Rapid termite mortality	Adult				
Hussain <i>et al.</i> , 2010	M.~anisopliae was found to be highly virulent (LT <sub>50</sub> - $3.10$ days)					
Wright and Cornelius, 2012	100% mortality recorded					
d Hussain and Yi, 2013	Higher percentage of exposed individuals could transfer inoculum resulting in high mortality					
Sun et al., 2008	99% mortality recorded					
Jones <i>et al.</i> , 1996	Termite workers dusted with dry conidia were capable of transmitting the pathogen to other					
Sun <i>et al.</i> , 2003	Infection eached Epizootic level	Adult	Nilliotetiiliudae	Diamodea	Copiotermes Jormosunus	. <del>1</del>
Sajap and Kaur, 1990	100% mortality recorded	A 4-1.		Distriction	Control of the contro	2
Milner et al., 1997 Hoe et al., 2009	No consistent effect on pathogenicity 100% mortality recorded	Adult	Rhinotermitidae	Blattodea	Coptotermes curvignathus	33.
	26 were identified as promising					
say, Milner <i>et al.</i> , 1998	26 were identified as promising Of the 93 isolates screened in the grooming assay,	Adult	Rhinotermitidae	Blattodea	Coptotermes acinaciformis	32.
say, Milner <i>et al.</i> , 1998	Of the 93 isolates screened in the grooming assay,	Adult	Rhinotermitidae	Blattodea	Coptotermes frenchi	31.
Brito et al., 2008	57.3 and 88.6% mortality recorded	Adult	Curculionidae	Coleoptera	Conotrachelus psidii	30.
Mendes et al., 2001	42.7% mortality observed	Grub	Curculionidae	Coleoptera	Conotrachelus humeropictus	29.
Silveiraet al., 2002	Affected pupae	Pupa	Chrysomelidae	Coleoptera	Coelomera lanio	28.
Haque and Ghosh, 2007	37.35-43.13% mortality recorded	Adult	Pseudococcidae	Hemiptera	Coccidohystrix insolita	27.
Pan <i>et al</i> 2012	commercially available insecucides. $Ma_{789}$ was the most virulent strain	Adult	Notodontidae	Lepidoptera	Clostera anachoreta	26.
Nitharwal et al., 2013	The treatment was least toxic compared to	Adult	Chrysopidae	Neuroptera	Chrysoperla carnea	25.
Brousseau <i>et al.</i> , 1996 arva.	Destruxins were toxic to 3rd, 4th and 5 <sup>th</sup> instar larvae with LD <sub>50</sub> of 0.116, 0.35 and 1.52 μ g/larva.	Larva				
	destruxins.	1 11	rotutotano	Topico postu	Circles retreated family come	ţ
Bronssean <i>et al</i> 1998	DAT.  The I December 72 h was 1 Au g/larva for	I arva	Tortricidae	Lenidonters	Charistanoura fumiforana	24
nd 6 Barman and Nath, 2002	Mortality of 6.66, 13.33 and 73.33% at 1, 3 and 6	Larva	Pieridae	Lepidoptera	Catopsilia crocale	23
Fan <i>et al.</i> , 1990	84.5-95.3% mortality recorded	Larva	Carposinidae	Lepidoptera	Carposina nipponensis	22.
Nedveckyte et al., 2011	100% mortality was seen in 18 days	Larva	Geometridae	Lepidoptera	Bupalus piniaria	21
Sabbour <i>et al.</i> , 2007	Gave good protection to broad beans.	Adult	Chrysomelidae	Coleoptera	Bruchus rufimanus	20.
ngal Retinam and Singh, 2007	Spraying of improved strains of the entomofungal	Grub	Chrysomelidae	Coleoptera	Brontispa longissima	19.

	52.	<u>1</u>	50.	49. 49.	47. 48		46.	45.	<del>1</del>	43.	42.	41.	40			39.	38.									37.
	Galleria mellonella	Franklinialla cooidantalia	Eutectona machaeralis	Esjanataria obesa Funroctis nseudoconsnersa	Epilachna beetle		Dysdercus cingulatus	Dociostaurus maroccanus	Denarocionus frontatis	Cydia funebrana	Cyclocephala comata	Curculio sikkimensis	Curculio sayi			Curculio nucum L	Corythucha ciliata									Coptotermes lacteus
	Lepidoptera	Thronostoro	Lepidoptera	Lepidontera	Coleoptera		Hemiptera	Orthoptera	Coreoptera	Lepidoptera	Coleoptera	Coleoptera	Coleoptera			Coleoptera	Hemiptera									Blattodea
	Pyralidae	Th	Pyralidae	A <i>criaidae</i> I ymantriidae	Coccinellidae		Pyrrhocoridae	Acrididae	Curcunonidae	Tortricidae	Dynastidae	Curculionidae	Curculionidae			Curculionidae	Tingidae									Rhinotermitidae
Adult	Larva	A daile	Larva	Larva	Grub	and Adult Adult	Nymph	Larva	Orub	Adult	Grub	Grub	Grub			nympn Grub	Adult and	Larva	Larva		Adult		Larva	Adult		Adult
Thoride, decreased toxicity.  The strong effect of destruxins (DE) was characterized by changes in the morphology of the cells.	Inhibition of proteinase by phenyl-methylsulfonyl	treatment causing 50.77 to 68.07% mortality.	M. anisopliae with 3 g/L found to be the best	98 - 100% mortanty in larval stages. 77 4% mortality recorded	90.42% mortality on 2nd and 7th day.	nymphs and adults of red cotton bugs. The LC <sub>50</sub> values ranged from $2.25 \times 10^5$ to $3.66 \times 10^8$ spores/ml.	and adult stages.  Shown highly effective against both 6±1 day old	anisopliae.  Strong mortality in the insects treated at the larval	conidia were $7.2 \times 10^3$ and $1.1 \times 10^5$ for $M$ .	Caused the highest mortality to moth species.	Susceptible to <i>M. anisopliae</i> , with LD <sub>50</sub> of 2.1x10 <sup>11</sup> conidia per larva.	At 25°C pathogenecity of <i>Metarhizium</i> was Maximum	56% mortality recorded	The total number of species and individuals were slightly lower in the sprayed areas than in the unsuraved areas	was 6.38x10 <sup>8</sup> spores litre <sup>-1</sup> . Median lethal time was 8.60-15.19 days for 1.0x10 <sup>11</sup> -1.0x10 <sup>7</sup> spores litre <sup>-1</sup> .	80% mortality recorded	Pathogenic	75% mortality achieved	M. anisopliae colonies were restricted to larval	significant infection.	M. anisopliae may have potential to cause	and the tiles were dug up 20 days later and examined for living and dead larvae	20 weevil larvae were applied to the soil surface	Isolates such as FI-1037, FI-1099 and FI-1186 were markedly less renellent	termites	Repellency of Metarhiziumanisopliae conidia to
Vey and Quiot, 1989	Kucera, 1980	A 2007: at al. 2007	Chavan <i>et al.</i> , 2012	Bagneri <i>et al.</i> , 2007 Fan <i>et al</i> 1988	Haque <i>et al.</i> , 2007	Sahayaraj and Borgio, 2009	Rai <i>et al.</i> , 2013	Outtar <i>et al.</i> , 2011	1980	Stephan and Herker, 2011	Ponce et al., 2004	Ihara <i>et al.</i> , 2009	Ihara <i>et al.</i> , 2003	Jiang <i>et al.</i> , 2005	Cheng, 2000	Carrera et al., 2013	Sevim <i>et al.</i> , 2013	1964 Ilan <i>et al.</i> , 2009	Gottwald and Tedders,		Ilan et al., 2009		Tedders et al., 1973	Staples and Milner, 2000		Milner, 2000

Balachander et al., 2012	Highly lethal with significantly lower LD50 value					
Remadevi <i>et al.</i> , 2010	The LC <sub>50</sub> of the isolates ranged from $0.01x10^5$ to $759.21x10^5$	Larva	Pyralidae	Lepidoptera,	Hypsipyla robusta	70.
Salvatierra and Berrios, 1972	Larvae are most susceptible to <i>Metarhiziumanisopliae</i> during the fifth instar.	Larva	Pyralidae	Lepidoptera	Hypsipyla grandella	69.
Burjanadze <i>et al.</i> , 2013 Montermini <i>et al.</i> , 1985	Mortality ranged from 52% to 68% 83%-100 mortality recorded					
Tserodze and Murvandze,	Mortality up to 86.8%	Larva	Arctiidae	Lepidoptera	Hyphantria cunea	68.
Schabel, 1976	759.21x10 <sup>3</sup> for the different pests.  High mortality. When larger doses were employed, the insects succumbed faster	Adult	Curculionidae	Coleoptera	Hylobius pales	67.
Remadevi et al., 2010	The LC <sub>50</sub> of the isolates ranged from $0.01 \times 10^5$ to	Adult				
Sakchoowong, 2002	(32.3 aphids/floret). 37.69 % mortality. Pathogenic to large	Larva	Hyblaeidae	Lepidoptera	Hyblaea puera	66.
Ramanujam <i>et al.</i> , 2013	pathogenic tungi.  Reduced maximum population after first spray	Adult	Aphididae	Hemiptera	Hyadaphis coriandri	65.
Sharma <i>et al.</i> , 1999	Penicillium was the most antagonistic towards the	Grub	Scarabaeidae	Coleoptera	Holotrichia consanguinea.	64.
Chouvenc et al., 2009 Chouvenc et al., 2009	Large diversity in susceptibility to disease.  Large diversity in susceptibility to disease.	Adult Adult	Hodotermitidae Termopsidae	Isoptera Isoptera	Hodotermes mossambicus Hodotermopsis sjoestedti	62. 63.
Pegu <i>et al.</i> , 2013	Four species of entomogenous fungi were isolated from 8 insect species .Confirmed their pathogenic relationship with varying degree of infectivity.	Adult	Acrididae	Orthoptera	Hieroglyphus banian	61.
Castiglioni <i>et al.</i> , 2005 Moino Junior <i>et al.</i> , 2002	Pathogenic. The penetration, colonization and conidiogenesis phase is faster for <i>M. anisopliae</i> which results in a faster rate of insect mortality.					
visnwakarina et al., 2011 Sterling et al., 2011 Castiglioni et al., 2003	16.15 % mortully 100% of insects eliminated in six days.  Nimkol-L was compatible with <i>M. anisopliae</i> strain up to 1% a.i.	Adult	Rhinotermitidae	Isoptera	Heterotermes tenuis	60.
Swaminathn and Hussain, 2010	Decrease in feeding	Grub	Coccinellidae	Coleoptera	Henosepilachna vigintioctopunctata	59.
Patil and Naik, 2004	32.5% mortility.	Adult	Miridae	Hemiptera	Helopeltis antonii	58.
Rao et al., 2011	73% mortaliy and 77% weight reduction over	Larva				
Haque <i>et al.</i> , 2007	29.12-58.25% mortality recorded	Larva	Noctuidae	Lepidoptera	Helicoverpa armigera	57.
Leite <i>et al.</i> , 2011	10 <sup>7</sup> conidia mL <sup>-1</sup> caused mortality in insect.	Grub	Cerambycidae	Coleoptera	Hedypathes betulinus	56.
2010 Pena <i>et al.</i> , 2011	75% mortality occurred	Adult	$Phlae othripidae \$	Thysanoptera	gyllenhal Gynaikothrips uzeli	55.
Molina and Carbone,	lerp psyllid Weak toxicity noticed	Grub	Curculionidae	Coleoptera	Gonipterusscutellatus	54.
Pogetto et al., 2011	Tested products were pathogenic to the red gum	Nymph	Psyllidae	Hemiptera	Glycaspis brimblecombei	53.

92.	91.	90.	89.	87.	86.	85.	84. 83.	<u>81.</u>	80 80	78.	7 <u>5.</u> 76. 77.	<u>74.</u>	73.	71. 72.
Monochamus alternatus	Microcerotermes diversus Silvestri	Micadina yingdeensis	Melanotus cribricollis	Maruca vitrata	cinerascens Lymantria dispar	Locusta migratoria	Lipaphis erysimi Liriomyza trifolii Locusta migratoria	Leucopholis lepidophora	Leptopharsa heveae Leucopholis irrorata	Leptoglossus zonatus	Leptispa pygmaea Leptocorisa oratorius Leptoglossus occidentalis	Lepidiota stigma	Kalotermes flavicollis	lcerya purchasi Ips typographus
Coleoptera	Isoptera	Phasmida	Coleoptera	Lepidoptera	Lepidoptera	Orthoptera	Hemiptera Diptera Orthoptera	Coleoptera	Hemiptera Coleontera	Hemiptera	Coleoptera Hemiptera Hemiptera	Coleoptera	Dictyoptera	Hemiptera Coleoptera
Cerembycidae	Termitidae	Heteronemiidae	Elateridae	Crambidae	Erebidae	Acrididae	Aphididae Agromyzidae Acrididae	Scarabaeidae	Tingidae Melolonthidae	Coreidae	Chrysomelidae Alydidae Coreidae	Scarabaeidae	Kalotermitidae	Monophlebidae Curculionidae
Adult		Adult	Grub	Larva	Larva Larva	Nymph Larva	Adult Larva Adult	Grub	Larva Gruh	Adult	Adult Adult Adult	Grub	Adult	Adult Adult
respectively 90% mortality recorded	Treated-sawdust bait was applied by two methods, LC <sub>50</sub> and LC <sub>90</sub> were $8.4 \times 10^6$ and $3.9 \times 10^7$ spore/ml,	Dosage of 1.5x10 <sup>13</sup> spores/ha resulted in 70%	Mortality increased with increase of temperature and reached 06 7%, at 30°C	Metarhizium anisopliae was moderately effective	larval and adult stages 100% mortality within 120h of treatment The insecticide activity was studied in relation to the fungus spore concentration, the incubation time and the larval instar, in order to establish the different lethal doses.	Walking activity was reduced to 24% High mortality in the insects treated at both the	The applications of <i>Metarhiziumanisopliae</i> 2 x 10 <sup>8</sup> at 20 g/palm caused 60.94 and 50.97% mortality Recorded 64% mortality in 3.8 days Efficient compound <i>Metarhizium</i> can suppress small local populations of <i>L. migratoria</i>	With 4x10 <sup>8</sup> conidia 33.33% mortality was achieved 60.94 % mortility was recorded	Large variation of virulence observed	94% Mortality recorded.	42 % effective mortality Effective against gundhi bug, damaged grain Sensitive to infection, killed the treated individuals within 10 to 30 days.	Lower number of grubs observed in fields applied with $1 \times 10^{13}$ /ha (0.33/m row)	Metarhizium anisopliae was found for the first time to attack I. typographus  Large diversity in disease susceptibility	of 2.6% 20.3% Mortality recorded Infection was highest when beetles had contact with freshly suspension treated bark for five minutes 97% mortality recorded
He et al., 2009	Habibpour et al., 2011	ShiLan et al., 1994	Wang <i>et al.</i> , 2010	Sunithaet al., 2008	Wasti and Hartmann, 1982 Ouakid <i>et al.</i> , 2005	Ranaivo <i>et al.</i> , 1996 Outtar <i>et al.</i> , 2011	Araujo Junior et al., 2010 Araujo Junior et al., 2009 El-Salam et al., 2013 Hunter et al., 1999	Rakesha et al., 2012 Channakeshavamurthyet	Silva <i>et al.</i> , 2012 Braza, 1990	Grimm and Guharay, 1998	Japur <i>et al.</i> , 2012 Kalita <i>et al.</i> , 2009 Rumine and Barzanti, 2009	Rachappa et al., 2004	2013 Keller <i>et al.</i> , 2004 Chouvenc <i>et al.</i> , 2009	Fan et al., 1988 Herrmann and Wegensteine, 2011 Mudroncekova et al.,

104.	102. 103.	95. 96. 97. 98. 99. 100.	94.
Otiorhynchus ovatus	Odontotermeswallonensis Oryctes rhinoceros	Nasutitermes sp Nasutitermes voeltzkowi Neocerambyx mandarinus Ocinara varians Odontotermes sp. Odontotermes formosanus Odontotermes obesus	Myzus persicae Nasutitermes exitiosus
Coleoptera	Isoptera Coleoptera	Isoptera Isoptera Coleoptera Lepidoptera Isoptera Isoptera	Hemiptera Isoptera
Curculionidae	Termitidae <i>Scarabaeidae</i>	Termitidae Termitidae Cerembycidae Bombycidae Termitidae Termitidae	Aphididae Termitidae
Adult Adult Grub Adult Grub	Adult Adult Adult Adult and grub Grub Grub	Adult Adult Grub Larva Adult Adult	Adult Adult and grub Grub Adult Adult
Wet inoculum was effective for the control of <i>Oryctes rhinoceros</i> on oil palm 83% infection in 4 months after treatment Dipping 3rd-instar larvae briefly in conidial suspensions at 10 million spores/ml led to the development of brown lesions on the body of the larvae  The pathogen can reduce <i>O. rhinoceros</i> populations  The fungus was highly virulent even at the low	Transmission of <i>Metarhizium anisopliae</i> was 50 to 98% and 16 to 78% for the isolates tested At 5 ml/litre showed lower efficacy 100% of mortality of termites recorded Killing 91.7% of the larvae and between 63% and 69% adults  Recorded 90% mortality after treatment The fungus was highly virulent against all stages of the pest except egg.	Decreasing RH reduced germination but no consistent effect on pathogenicity Infected termites died 1 to 3 days post-inoculation Large diversity in susceptibility to disease Recorded 92.7% mortalitity Fungal spores retarded larval growth by 35% -76% LC <sub>50</sub> of the isolates ranged from 0.01x10 <sup>5</sup> to 759.21x10 <sup>5</sup> . Effective 100% mortality 3 days post-inoculation Pathogenic relationship with varying degree of	Results showed that <i>Metarhiziumanisopliaers</i> 9 (Ma <sub>789</sub> ) was the most virulent one.  85% mortality recorded  The mortality was 80.0% and 76.0% for larvae and 83.4% and 88.9% for adults  With the cadaver rates being as high as 76.9-93.1% (1.0x10 <sup>7</sup> conidia/ml) and 57.9-75.0% (6.5x10 <sup>5</sup> -3.4x10 <sup>6</sup> conidia per individual) for the larvae at 15 days after inoculation and adults at 20 days after inoculation, respectively  Pathogenic relationship with varying degree of infectivity  Isolate FI-610 was found to be one of the most effective  Almost half the treated colonies became moribund. Samples from treated colonies showed high levels
Moslim et al., 1999 Tey and Ho,1995 Sivapragasam and Tey, 1994 Bedford, 2013 Fisher and Bruck, 2008	Balachander et al., 2013 Premalatha et al., 2008 Nagaraju et al., 2013 Moslim et al., 2011 Moslim et al., 2009 Kalidas and Prasad, 2001	Milner et al., 1997 Gutierrez et al., 2004 Chouvenc et al., 2009 Fan et al., 1988 Hussain et al., 2009 Remadevi et al., 2010 ChangJin et al., 2019 Pegu et al., 2013	YongSheng et al., 2010 He et al., 2008 He et al., 2007 He et al., 2005  Pegu et al., 2013  Milner et al., 1998  Hanel, 1983 Hanel and Watson, 1983

115.	114	112. 113.	109 110 111.	108	107.	105. 106.	
. Popillia japonica	. Polyphagotarsonemus latus	. Pleonomus canaliculatus . Plocaederus ferrugineus	. Pityogenes chalcographus . Plagiodera versicolora . Platypus cylindrus	. Phyllophaga sp.	. Pantana phyllostachysae	. Pachycoris klugii Paliga machoeralis	
Coleoptera	Trombidiformes	Coleoptera Coleoptera	Coleoptera Coleoptera Coleoptera	Coleoptera	Lepidoptera	Hemiptera Lepidoptera	
Scarabaeidae	Tarsonemidae	Elateridae Cerambycidae	Curculionidae Chrysomelidae Curculionidae	Scarabaeidae	Lymantriidae	Scutelleridae Crambidae	
Adult	Grub Grub Grub Larva and	Grub Grub	Adult Grub Adult	Grub	Larva	Adult Larva	Adult Grub Grub
Infected 1.2% of the population	Pouring conidial suspension achieved 33.3-36.4% recovery of infested trees followed by swabbing conidial slurry with 23-25% and soil application with 15.4-16.6%. 91.66% mortality recorded Fungus caused a green muscardine disease in larvae 100% mortality recorded Successful in controlling both larvae and adult	Pathogenic.  All the treatments were effective and superior over	treated plots.  Capable of causing high mortality.  Pathogenic.  Infected adults were able to contaminate larvae by	instar larvae. 47.16% decrease in plant mortality over control in	759.21x10 <sup>5</sup> Among the 25 isolates tested, MIS2, MIS7, MIS1 and MIS3 were found to be more effective All strains showed pathogenicity to 2nd- and 3rd-	Recorded 65% mortality The LC <sub>50</sub> of the isolates ranged from $0.01 \times 10^5$ to	dose of 1x10 <sup>4</sup> Over 98% control achieved Temperatures below 20°C (68F) significantly slowed fungal growth and the speed at which <i>M. anisopliae</i> infected BVW larvae. Reduction in larval numbers ranging from 0 to 96% and 0 to 90% at East Mailing and Little hampton respectively Three of the 5 isolates of <i>Metarhiziumanisopliae</i> controlled larvae Pathogenic at 1 x 10 <sup>6</sup> spores/cm³.Ultimately gave complete kill even at 1 x 10 <sup>5</sup> conidia ml <sup>-1</sup> 74 and 81% control of the pest achieved Significant reduction on root herbivory Pathogenic at 1 x 10 <sup>6</sup> spores/cm³. Ultimately gave complete kill even at 1 x 10 <sup>5</sup> conidia ml <sup>-1</sup> Higher efficacy when conidia mixed with neem seed cake (5 g/l of peat) The fungus was highly virulent even at the low dose of 1x10 <sup>4</sup> spores per g dry soil.
Hanula anf Andreadis,	Ambethgar, 2010  Ambethgar, 2001 Choudhuri, 1973 Ambethgar et al., 1999 Maketon et al., 2008	HuaChao <i>et al.</i> , 2002 Sahu and Sharma, 2008	Pehl and Kehr, 1994 Demir <i>et al.</i> , 2013 Glare <i>et al.</i> , 2002	Bhagat, 2005	Sapna Bai <i>et al</i> ., 2013 Zhang <i>et al</i> ., 2002	Grimm and Guharay, 1998 Remadevi <i>et al.</i> , 2010	Shah et al., 2007  Bruck, 2007  Moorhouse et al., 1993  Tol and Van, 1993  Prado, 1980  Stenzel et al., 1992 Oddsdottir et al., 2010 Prado, 1980  Shah et al., 2008  Fisher and Bruck, 2008

130.	129.	128.	127.	126.	125.		123. 124.											122.				121.	120.	119.	118.	,	116. 117.
Schistocerca gregaria	Saperda populnea	Saccharosydne saccharivora	Leucopholis lepidophora	Rhyacionia frustrana	Rhagoletis cerasi		Reticulitermes grasset Reticulitermes sp.											Reticulitermes flavipes			speratus	Reticutermes	Raphidopalpa foveicollis	canarum Pyrausta coclesalis	Pseudophacopteron	Personal Personal	Prorhinotermes canalifrons Pseudacysta nerseae
Orthoptera	Coleoptera	Hemiptera	Coleoptera	Lepidoptera	Diptera		Blattodea											Isoptera				Blattodea	Coleoptera	Lepidoptera	Hemiptera		Isoptera Hemintera
Acrididae	Cerambycidae	Delphacidae	Scarabaeidae	Tortricidae	Tephritidae		Kninotermitidae Rhinotermitidae											Rhinotermitidae				Rhinotermitidae	Chrysomelidae	Crambidae	Phacopteronidae	Ö	Rhinotermitidae Tinoidae
Larva and adult			Grub	YMAK	Larva and Adult	Adult	Adult		Adult	Adult	Adult Adult		Adult	Adult	Adult	Adult	, xci ci t	Adult				Adult		Larva		Nymphs	Adult and
Recorded 100% mortality	Recorded 94.1% mortality	Reduced population of the pest from 61.25 to 20.14%	2 x 108 at 20 g/palm caused 60.94 and 50.97%	Population of <i>R. frustrana</i> decreased to very low levels on the treated plots	termites were pathogenic to healthy termites Fungal strains were able to cause mycosis	Fungi reisolated from the hind gut of diseased	Recorded 93.44% mortality Healthy termites concentrate in grooming activity on diseased individuals and thereby became infected	Good controlling agent	Recorded 92% mortality	Recorded 100% mortality	Recorded 116% mortality	of the cadaver	increased 24 h after fungal exposure  Hyphal growth was generalized in the body cavity	The relative number of hemocytes per termite	germmated 90% mortality recorded	None of the conidia found in the alimentary tracts	anisopliae varied greatly	Large diversity observed in susceptibility to disease  Physiological cost to successfully encapsulate M	conidia were detected in the foreguts of the termites reared individually.	contained the conidia in their foreguts, but no	Within 3 h. almost all of the termites held in groups	The conidia affiliated with the termites reared	Reduction in damage 64.7% at 3.0 g/lit.	Recorded 36 % mortality	Recorded8.7-53% mortality	exceeding 90%.	Large diversity in susceptibility to infection  At 30 day, treatments showed a biological efficacy
Miscellaneous. 1992	Fan <i>et al.</i> , 1988	Vanegas et al., 2006	Murthy et al., 2010	Duarte <i>et al.</i> , 1992	Daniel <i>et al.</i> , 2008	Kramm and West, 1982	Alvarez <i>et al.</i> , 2005 Kramm <i>et al.</i> , 1982	Milner <i>et al.</i> , 1996	Quarles, 1999	Zoberi, 1995	Wang and Powell, 2004		Chouvenc et al., 2009	Chouvenc et al., 2009	Chouvenc et al., 2008	Chouvenc et al., 2010	CHOUNCING to the, FOLL	Chouvenc et al., 2009			2003	Shimizu and Yamaji,	Vishwakarma et al., 2011	Rishi <i>et al.</i> , 2012	RongYing et al., 1995		1988 Chouvenc <i>et al.</i> , 2009 Romero <i>et al.</i> , 2012

				Larva	Strong mortality in the insects treated at the larval	Outtar et al., 2011
131.	Scolytus multistriatus	Coleoptera	Curculionidae	Grub	Recorded 100% mortality	Houle <i>et al.</i> , 1987
132.	Scolytus scolytus	Coleoptera	Curculionidae	Adult	Limited mortility observed	Doberski, 1981
133.	Sitophilus oryzae	Coleoptera	Curculionidae	Adult	Percentages of infestation were significantly	Sabbour, 2012
134.	Spilarctia obliqua	Lepidoptera,	Acrididae	Adult	Tested isolates were virulent	Remadevi et al., 2010
135.	Spodoptera litura	Lepidoptera	Noctuidae	Larva	73% mortality and 77% weight reduction Pathogenic relationship with varying degree of	Rao <i>et al.</i> , 2011 Pegu <i>et al.</i> , 2013
					infectivity.	,
136.	Spondylis buprestoides	Coleoptera	Ceramby cidae	Grub	Effectively controlled.	ChengRun, 2005
137.	Stigmella populnea	Coleoptera	Nepticulidae	Grub	Recorded 70.8% control	Fan <i>et al.</i> , 1990
138.	Strophosoma capitatum	Coleoptera	Curculionidae	Adult	Average survival time at 20° C ranged between 23 and 28 days.	Nielsen <i>et al.</i> , 2006
				Adult	80% mortality in lab and 60% mortality in the field.	Nielsen et al., 2007
139.	Strophosoma melanogrammum	Coleoptera	Curculionidae	Adult	Direct inoculation caused over 80% mortality. Indirect inoculation resulted in 72% mortality.	Kram, 2010
				Adult	80% mortality in lab and 60% mortality in the field.	Nielsen <i>et al.</i> , 2007
140.	Taeniothrips inconsequens	Thysanoptera	Thripidae	Larva	The highest rates of infection were seen in larvae	Brownbridge et al., 1999
	,	,			recovered from soil samples (11.9%)	(
141.	Termites	Isoptera	Rhinotermitidae	Adult	Efficient for controlling termites	Nyeko <i>et al.</i> , 2010
142.	Termites	Isoptera	Rhinotermitidae	Adult	Behavioural defence mechanisms by termites can limit the effectiveness of conidia applications	Lenz , 2005
143.	Thaumetopoea pityocampa	Lepidoptera	Thaumetopoeidae	Larva	Mortality was significantly high	Er et al., 2007
144.	Toxoptera aurantii	Hemiptera	Aphididae	Adult	Observed pathogenic	Pegu <i>et al.</i> , 2013
145.	Trialeurodes vaporariorum	Hemiptera	Aleyrodidae	Maggot	Recorded 75% mortality	Pena <i>et al.</i> , 2011
					Recorded 98% efficacy	Prishchepa et al., 2005
146.	Tuta absoluta	Lepidoptera	Gelechiidae	Egg and Larva	Pathogenic to eggs and larvae	Pires <i>et al.</i> , 2010
147.	Vatiga illudens	Hemiptera	Tingidae	Adult	Recorded 74% Mortality	Oliveira et al., 2001
148.	Zaprionus indianus	Diptera	Drosophilidae	Adult	Recorded 100% mortality	Svedese et al., 2012
149.	Zeuzera pyrina	Lepidoptera	Cossidae	Larva	Recorded 95-99% mortality	Deseo and Docci, 1985

## CONSERVATION OF ENTOMOPATHOGENIC FUNGUS METARHIZIUM

Biological control is defined as the reduction of pest populations by natural enemies and typically involves an active human role. Conservation biological control (CBC) is a biological control strategy in which farming practices and environmental manipulations are adopted to enhance the living conditions for specific natural enemies of pests (Meyling and Eilenberg, 2007). Avoiding measures that are harmful to natural enemies and adapting measures that can benefit them may provide successful biological control. Probably, the first comprehensive review of the relationships of crop pests and beneficial insects with uncultivated land (van Emden, 1965) was able to cite examples of all the components of Conservation Biological Control by habitat modification used in practice today. There are three basic types of biological pest control strategies, importation (sometimes called classical biological control), augmentation and conservation. Importation is the introduction of a species in some area with the intention of biological control. Augmentation defines the actions or processes by which population of biocontrol agent can be increased. Conservation means act of preserving or guarding a biocontrol agent. Natural enemies of insect pests, also known as biological control agents, include predators, parasitoids, and pathogens. A predator is an organism which naturally preys on another organism, a parasitoid is an insect whose larvae lives as parasite and eventually kills the host, and a pathogen is an agent that causes illness in the host or kills it. Metarhizium as an entomopathogen is a natural enemy of pests found in forest and agricultural land and is a candidate for future conservation biological control in temperate regions. However, compared with classical and augmentation biological control, it has received relatively little attention as a method of arthropod pest suppression (Ehler, 1998; Landis et al., 2000). It is generally acknowledged that biological control alone is unlikely to provide adequate pest control, but through careful integration with other pest suppression tactics, it could represent a significant source of sustainable control (Dowell, 1990; Gerling, 1992; Cock, 1994; Heinz, 1996). Here question arises what can be done for conservation of this natural enemy of pests, and the answer is hidden somewhere in the biology and ecology of this entomopathogen. Like other organisms, these natural enemies too require food, water, and shelter, and protection from adverse conditions. One the best way could be avoiding use of insecticides and pesticides which has both direct and indirect effect of eliminating natural enemies. Insecticides not only kill harmful insects but also significantly affects biological control agent. In soil, this species can withstand drought or any adverse climatic condition by forming sclerotium and can start multiplying under favourable conditions. Since it is a soil-borne fungus, it is perfectly protected from UV radiation. It is important to consider biological and ecological need of a natural enemy for the success of any biological control method. Environmental modifications may be made to increase natural enemy effectiveness. Some of these modifications include, A. Construction of artificial structures, B. Provision of supplementary food, C. Provision of alternative hosts, D. Improvement of pestnatural enemy synchronization, E. Modification of adverse

agricultural practices.

#### CONCLUSION

The forest is a dynamic ecosystem constituting principal natural renewable resources of multifarious uses which fulfill the requirement of the society and sustainability of the earth. Insect pests are major biological determinants of forest productivity, and integrated pest management is not new to Indian forestry (Sundararaj, 2014). Biological control is a self sustaining measure that could be adopted for long term results in forest habitat as well as in field conditions. It can also be used in formulation form on timber along with timber polish, or can be sprayed and left as such. Though biological control with entomopathogens like Metarhizium is most frequently encouraged to be practiced, there are some limitations with this method. Not all the target pests are affected up to the expectation. Field and lab trials results some times vary greatly. Some insects get infected when they are in their larval stage and some in their adult stage. In case of some insects, results were almost 100% whereas for others mortality was as low as 15% only. Death time also varies with different pests, which vary from 3 to several days. Infection rates were high in freshly cultured isolates as compared to old and stored cultures. Temperature and humidity plays vital role in attachment and growth of conidia. If temperature is lower than 20°C it slows the infection rate of the fungus. When Metarhizium conidia were applied with sunflower oil or IMI, comparatively higher mortality rate was observed. Pests exposed to Metarhizium also showed shortened longevity and decreased oviposition. It also reduces hatching percentage of laid eggs. Wet inoculum is more effective as compared to dry inoculum. Biopesticide has been mostly used in agricultural field compared to forests. Introduction of an effective bioconrol agent in the forest plantaion may provide a solution for the damage caused by notorious insect pests.

#### REFERENCES

Ambethgar, V. (2001) Indigenous entomofungi for biological control of stem and root borer, *Plocaederus ferrugineus* L. infesting cashew. *South Indian Horticulture*, 49(Special),281-284.

Ambethgar, V., Lakshmanan, V., Dinakaran, D and Selvarajan, M. (1999) Mycosis of cashew stem and root borer, *Plocaederus ferrugineus* L. (Coleoptera, Cerambycidae) by *Metarhizium anisopliae* (Metsch.) Sorokin (Deuteromycotina, Moniliales) from Tamil Nadu (India). *Journal of Entomological Research*, 23(1),81-83. Ambethgar, V. (2010) Field assessment of delivery methods for fungal pathogens and insecticides against cashew stem and root borer, *Plocaederus ferrugineus* L. (Cerambycidae, Coleoptera). *Journal of Biopesticides*,3(1),121-125.

Anjana Patial Bhagat, R. M. (2005) Field evaluation of some insecticides against white grub in maize under midhill conditions of Himachal Pradesh. *Journal of Entomological Research*, 29(2), 123-125.

Ansari, M. A., Shah, F. A., Whittaker, M., Prasad, Mand Butt, T. M. (2007) Control of western flower thrips (*Frankliniella occidentalis*) pupae with *Metarhizium* 

anisopliae in peat and peat alternative growing media. *Biological Control*, 40(3),293-297.

Araujo Junior, J. M., de Marques, E. J and Oliveira, J. V. de. (2009) Potential of *Metarhizium anisopliae* and *Beauveria bassiana* isolates and neem oil to control the aphid *Lipaphis erysimi* (Kalt.) (Hemiptera, Aphididae). *Neotropical Entomology*, 38(4),520-525.

Augustyniuk-Kram, A. (2010) Mortality of the nut-leaf weevil *Strophosoma melanogrammum* (Forster) and damage rate of needles after treatment with entomopathogenic fungi. *Journal of Plant Protection Research*, 50(4), 545-550.

Azevedo, F. R., de Guimaraes, J. A., Braga Sobrinho, R and Lima, M. A. A. (2005) Efficiency of natural products to control *Bemisia tabaci biotype B* (Hemiptera, Aleyrodidae) on melon plant. *Arquivos do Instituto Biologico* (Sao Paulo), 72(1), 73-80.

Bagheri, S., Tajvand, B., Khajehzadeh, Y and Askary, H. (2007) Tree trunk spraying as an insecticidal control method of oak short-wing grasshopper, *Esfandiaria obesa* (Orth., Acrididae). *Iranian Journal of Forest and Range Protection Research*, 5(1), 26-38.

Balachander, M., Remadevi, O. K and Sasidharan, T. O.(2013). Dissemination of *Metarhizium anisopliae* infection among the population of *Odontotermes obesus* (Isoptera, Termitidae) by augmenting the fungal conidia with attractants. *Journal of Asia-Pacific Entomology*, 16(3), 199-208.

Balachander, M., Remadevi, O. K., Sasidharan, T. O and Bai, N.S. (2012) Virulence and mycotoxic effects of *Metarhizium anisopliae* on Mahogany shoot borer, *Hypsipyla robusta* (Lepidoptera, Pyralidae). *Journal of Forestry Research*, 23(4), 651-659.

Barman, H. K and Nath, R. K. (2002) Effectiveness of certain insecticides and bio-pesticides against *Catopsilia crocale* Cram. (Pieridae, Lepidoptera) on Cassia fistula. *Insect Environment*,8(1),28-29.

Batalla-Carrera, L., Morton, A., Santamaria, S and Garciadel-Pino, F. (2013) Isolation and virulence of entomopathogenic fungi against larvae of hazelnut weevil *Curculio nucum* (Coleoptera, Curculionidae) and the effects of combining *Metarhizium anisopliae* with entomopathogenic nematodes in the laboratory. *Biocontrol Science and Technology*, 23(1), 101-125.

Bedford, G. O.(2013)Biology and management of palm dynastid beetles, recent advances. *Annual Review of Entomology*, 58, 353-372.

Braza, R. D. (1990) Laboratory evaluation of Metarhizium anisopliae (Metsch.) Sorokin against *Leucopholis irrorata* (Chevrolat) (Coleoptera, Scarabaeidae). *Philippine Entomologist*, 8(1), 671-675.

Brito, E.S., Paula, A.R., de Vieira, L.P., Dolinski, C and Samuels, R.I. (2008) Combining vegetable oil and sublethal concentrations of Imidacloprid with *Beauveria bassiana* and *Metarhizium anisopliae* against adult guava weevil *Conotrachelus psidii* (Coleoptera, Curculionidae). *Biocontrol Science and Technology*, 18 (7), 665-673. 22.

Broglio, S.M.F., Lopes, D.O.P., Santos, D.S., dos Dias-Pini, N., da S., Giron-Perez, K and Micheletti, L. B. (2014) Control evaluation of *Rhipicephalus* (Boophilus) microplus in vivo with *Metarhizium anisopliae* and extract of *Annona muricata*. *Magistra*, 26 (4), 543-546. Brousseau, C., Charpentier, G and Belloncik, S. (1998) Effects of *Bacillus thuringiensis* and destruxins (*Metarhizium anisopliae* mycotoxins) combinations on spruce budworm (Lepidoptera, Tortricidae), *Journal of Invertebrate Pathology*, 72(3),262-268.

Brousseau, C., Charpentier, Gand Belloncik, S. (1996) Susceptibility of spruce budworm, *Choristoneura fumiferana* Clemens, to destruxins, cyclodepsipeptidic mycotoxins of *Metarhizium anisopliae. Journal of Invertebrate Pathology*, 68(2),180-182.

Brownbridge, M., Adamowicz, A., Skinner, M. and Parker, B.L. (1999) Prevalence of fungal entomopathogens in the life cycle of pear thrips, *Taeniothrips inconsequens* (Thysanoptera, Thripidae), in Vermont sugar maple forests. *Biological Control*, 16(1), 54-59.

Brownbridge, M., Townsend, R. J., Nelson, T. L., Gicquel, B and Gengos, M.(2009) Susceptibility of red-headed cockchafer, *Adoryphorus couloni*, in New Zealand to *Metarhizium anisopliae* strain DAT-F001 (Chaferguard <sup>TM</sup>), *New Zealand Plant Protection*, 62,395.

Bruck, D. J. (2007) Efficacy of *Metarhizium anisopliae* as a curative application for black vine weevil (Otiorhynchus sulcatus) infesting container-grown nursery crops. *Journal of Environmental Horticulture*, 25(3), 150-156.

Burjanadze, M., Nakaidze, E., Arjevanidze, Mand Abramishvili, T. (2013) Effect of local strains of *Beauveria bassiana* (Bb024) and *Metarhizium anisopliae* (M7/2) against the fallweb worm *Hyphantria cunea* (Lepidoptera, Arctiidae) in Georgia. *IOBC/WPRS Bulletin*, 90, 97-101.

Castiglioni, E., Djair Vendramim, J and Batista Alves, S. (2003) Compatibility between *Beauveria bassiana* and *Metarhizium anisopliae* with Nimkol-LReg. in the control of *Heterotermes tenuis*. *Manejo Integrado de Plagas y Agroecologia*, (69), 38-44.

Castiglioni, E., Vendramim, J. D and Alves, S. B.(2005) Effect of the association of Meliaceae by-products and entomopathogenic fungi on the survival of *Heterotermes tenuis* (Hagen, 1858) (Isoptera, Rhinotermitidae). *Agrociencia (Montevideo)*, 9(1/2), 347-356.

Castilho, A. M. C., Fraga, M. E., Aguiar-Menezes, E. de L., Rosa, C. A and da R. (2010) Selection of *Metarhizium anisopliae* and *Beauveria bassiana* isolates pathogenic *to Atta bisphaerica* and *Atta sexdens* rubropilosa soldiers under laboratory conditions. *Ciencia Rural*, 40(6),1243-1249.

Castillo M. A., Moya P., Hernandez E and Primo-Yufera E.(2000) Susceptibility of *Ceratitis capitata* Wiedemann (Diptera, Tephritidae) to entomopatogenic fungi and their extracts. *Biological Control*, 19, 274-282.

Chai, Y. Q. (1995) Preliminary studies on the pathogenicity of some entomogenous fungi to Coptotermes formosanus. *Chinese Journal of Biological Control*. 11(2),68-69.

Channakeshavamurthy, H., Naik, M. I and Manjunatha, M. (2010) Evaluation of certain new chemicals, bio-agents and plant products for the management of arecanut root grub, *Leucopholis lepidophora* Blanch. *Mysore Journal of Agricultural Sciences*, 44(4), 815-817.

Chavan, S. M., Kabade, K. H and Sushil Kumar. (2012) Laboratory evaluation of different chemical insecticides and biopesticides against larvae of teak skeletonizer,

- Eutectona machaeralis (Walker). Journal of Biopesticides, 5 (Supplementary), 196-198.
- Chen, ShiLan ,, Xu , ShiDuo ,, Lai , XinHong., Hu , SiLin and Lin ,LongFu (1994) A study on the biological characteristics and control of *Micadina yingdeensis*. *Forest Research*, 7(2), 187-192.
- Chen DeLan. (2013) Control efficiency and pathogenicity in different strains of *Metarhizium anisopliae* on Apriona germari Hope larvae. *Journal of Nanjing Forestry University (Natural Sciences Edition)*, 37 (1), 78-82.
- Choudhuri, J. C. B.(1973) Mycosis of a stem and root borer of cashewnut, *Plocaederus ferrugineus* L. (Coleoptera, Cerambycidae). *Journal of Plantation Crops, publ.* 1974. 1,164-167.
- Chouvenc, T., Su, N.Y. and Elliott, M.L. (2008) Interaction between the subterranean termite *Reticulitermes flavipes* (Isoptera, Rhinotermitidae) and the entomopathogenic fungus *Metarhizium anisopliae* in foraging arenas. *Journal of Economic Entomology*, 101(3),885-893.
- Chouvenc, T., Su, N.Y and Robert, A. (2009) Cellular encapsulation in the eastern subterranean termite, *Reticulitermes flavipes* (Isoptera), against infection by the entomopathogenic fungus *Metarhizium anisopliae. Journal of Invertebrate Pathology*, 101(3), 234-241.
- Chouvenc, T., Su, N. Y. and Robert, A. (2009) Inhibition of *Metarhizium anisopliae* in the alimentary tract of the eastern subterranean termite *Reticulitermes flavipes*. *Journal of Invertebrate Pathology*, 101(2), 130-136.
- Chouvenc, T., Su, NanYao and Robert, A. (2009) Susceptibility of seven termite species (Isoptera) to the entomopathogenic fungus Metarhizium anisopliae. *Sociobiology*, 54(3), 723-748.
- Chouvenc, T., Su, N.Y. and Robert, A. (2010) Inhibition of the fungal *pathogen Metarhizium anisopliae* in the alimentary tracts of five termite (Isoptera) species. *Florida Entomologist*; 2010. 93(3), 467-469.
- Chouvenc, T., Su, N.Y. and Robert, A. (2011) Differences in cellular encapsulation of six termite (Isoptera) species against infection by the entomopathogenic fungus *Metarhizium anisopliae*. *Florida Entomologist*, 94(3), 389-397.
- Cock, M.J.W. (1994) Integrated management of whiteflypest problems in the Middle and Near East with special emphasis on biological control. *Arab J. Plant Prot.* 12,127–136.
- Daniel, C., Keller, S and Wyss, E. (2008) Susceptibility of Rhagoletis cerasi to entomopathogenic fungi. *IOBC/WPRS Bulletin*, 31,228-233.
- DeBach, P. (1974) Biological Control by Natural Enemies. *Cambridge University Press*, London, 323pp.
- Delate, K. M., Grace, J. K and Tome, C. H. M. (1995) Potential use of pathogenic fungi in baits to control the Formosan subterranean termite (Isopt. Rhinotermitidae). *Journal of Applied Entomology*, 119(6),429-433.
- Demir, I., Kocacevik, S., Sonmez, E., Demirbag, Z and Sevim, A. (2013) Virulence of entomopathogenic fungi against *Plagiodera versicolora* (Laicharting, 1781) (Coleoptera, Chrysomelidae). *African Journal of Agricultural Research*, 8(18), 2016-2021.
- Deseo, K. V and Docci, R.(1985) Microbiological control against *Zeuzera pyrina* L. (Lepidoptera, Cossidae). *Difesa delle Piante*, 8(2), 285-291.

- Doberski, J. W. (1981) Comparative laboratory studies on three fungal pathogens of the elm bark beetle Scolytus scolytus, pathogenicity of *Beauveria bassiana*, *Metarhizium anisopliae*, and *Paecilomyces farinosus* to larvae and adults of *S. scolytus. Journal of Invertebrate Pathology*, 37(2),188-194.
- Dong, ChangJin, Zhang, JiaMin, Huang, Hai Chen, WuGuo, Hu and YuanYang (2009) Pathogenicity of a new China variety of *Metarhizium anisopliae* (M. anisopliae var. dcjhyium) to subterranean termite *Odontotermes formosanus.Microbiological Research*, 164(1), 27-35.
- Dowell, R.V. (1990) Integrating biological control of whiteflies into crop management systems. In, Gerling, D. (Ed.), Whiteflies, their Bionomics, Pest Status and Management. *Intercept, Andover*, UK, pp. 315–335.
- Duarte, A., Menendez, J. M., Fernandez, A and Martinez, J.(1992)Use of a biopreparation of *Metarhizium anisopliae* strain Nina Bonita in plantations of *Pinus caribaea* for control of *Rhyacionia frustrana*. [Spanish] *Revista Baracoa*, 22(2),17-23.
- Dubois, T., Lund, J., Bauer, L. S and Hajek, A. E. (2008) Virulence of entomopathogenic hypocrealean fungi infecting *Anoplophora glabripennis*. *BioControl*, 53(3), 517-528.
- Echeverri-Molina, D and Santolamazza-Carbone, S.(2010)Toxicity of synthetic and biological insecticides against adults of the Eucalyptus snout-beetle gonipterusscutellatus gyllenhal (coleoptera, curculionidae). Journal of Pest Science, 83(3),297-305.
- Ehler, L.E. (1998) Conservation biological control, past, present, and future. In, Barbosa, P. (Ed.), *Conservation Biological Control. Academic Press*, New York, pp. 1–8. El-Latif, N. A. A and Solaiman, R. H. A. (2014) Foraging
- El-Latit, N. A. A and Solaiman, R. H. A. (2014) Foraging activity of the subterranean sand termite, *Psammotermes hybostoma* (Desneux) and its associated fungus *Metarhizium anisopliae* under natural environmental conditions in El-Fayoum Governorate, Egypt. *Egyptian Journal of Biological Pest Control*, 24(2), 321-328.
- El-Salam, A. M. E.A., Salem, H. A and Salem, S. A. (2013) Biocontrol agents against the leafminer, *Liriomyza trifolii* in faba bean fields. *Archives of Phytopathology and Plant Protection*, 46(9), 1054-1060.
- Er, M.K., Tunaz, H and Gokce, A. (2007) Pathogenicity of entomopathogenic fungi to *Thaumetopoea pityocampa* (Schiff.) (Lepidoptera, Thaumatopoeidae) larvae in laboratory conditions. *Journal of Pest Science*, 80(4), 235-239
- Fan, M. Z., Guo, Cand Li, N. C. (1990) Application of *Metarhizium anisopliae* against forest pests. *Proceedings and abstracts*, *Vth International Colloquium on Invertebrate Pathology and Microbial Control, Adelaide, Australia*, 20-24 August 1990, 172.
- Fan, M. Z., Guo, C., Xiao, H. L and Hu, Y. (1988) Pathogenicity of *Metarhizium anisopliae* and its use in forest pest control. *Chinese Journal of Biological Control*, 4(1), 29-32.
- Farashiani, M.E., Askary, H and Hoseini, M. E.(2008) Laboratory investigation on virulence of three entomopathogenic fungi against the larvae of *Aeolesthes sarta* (Col, Cerambycidae). [Arabic]Journal of Entomological Society of Iran, 28(1), 19-34.

Fisher, J. R and Bruck, D. J.(2008)Biology and control of root weevils on berry and nursery crops in Oregon. *Acta Horticulturae*, (777),345-351.

Gerling, D., (1992) Approaches to the biological control of whiteflies. *Fla. Entomol.* 75, 446–456.

Giraldo-Vanegas, H., Vargas, A., Sarmiento, A., Hernandez, E., Amaya, F., Ramirez, M., Ramirez, F and Contreras, E. J.(2006)Evaluation of bioplaguicides to control sugarcane leafhopper *Saccharosydne saccharivora* (Westwood) (Hemiptera, Delphacidae) in the valley of San Antonio-Urena, Tachira, Venezuela. *Agronomia Tropical* (*Maracay*), 56(2), 253-276.

Glare, T. R., Placet, C., Nelson, T. Land Reay, S. D. (2002) Potential of *Beauveria* and *Metarhizium* as control agents of pinhole borers (Platypus spp.). *New Zealand Plant Protection Volume 55*, 2002. Proceedings of a conference, Centra Hotel, Rotorua, New Zealand, 13-15 August 2002, 73-79.

Gottwald, T. R and Tedders, W. L. (1984)Colonization, transmission, and longevityof *Beauveria bassiana* and *Metarhizium anisopliae* (Deuteromycotina, Hypomycetes) on pecan weevil larvae (Coleoptera, Curculionidae) in the soil. *Environmental Entomology*, 13(2), 557-560.

Grimm, C and Guharay, F.(1998) Control of leaf-footed bug *Leptoglossus zonatus* and shield-backed bug *Pachycoris klugii* with entomopathogenic fungi. *Biocontrol Science and Technology*, 8(3),365-376.

Gutierrez G., A. I. Saldarriaga O, Y. (2004) Observation of the pathogenicity of *Metarhizium anisopliae* in Nasutitermes sp. soldiers (Isoptera, Termitidae). *Revista Colombiana de Entomologia*, 30(2),151-156.

Habibpour, B., Cheraghi, A and Mossadegh, M. S. (2011)Evaluation of cellulose substrates treated with *Metarhizium anisopliae* (Metschnikoff) Sorokin as a biological control agent against the termite *Microcerotermes diversus Silvestri* (Isoptera, Termitidae). *Journal of Entomological and Acarological Research*: 43(2),269-275.

Hajek, A. E., Huang, B., Dubois, T., Smith, M. T and Li, Z. (2006) Field studies of control of *Anoplophora glabripennis* (Coleoptera, Cerambycidae) using fiber bands containing the entomopathogenic fungi *Metarhizium anisopliae* and *Beauveria brongniartii*. *Biocontrol Science and Technology*, 16(3/4),329-343.

Hajek, A. E., Lund, J and Smith, M. T. (2008) Reduction in fitness of female Asian longhorned beetle (Anoplophora glabripennis) infected with Metarhizium anisopliae. Journal of Invertebrate Pathology, 98(2),198-205.

Hajek, A., Dubois, T., Lund, J. Smith, M., Bauer, L and Li ZengZhi. (2007) Developing fungal bands for control of Asian longhorned beetle, *Anoplophora glabripennis*, in the U. S. *Journal of Anhui Agricultural University*. 34(2),149-156

Hanel, H and Watson, J. A. L.(1983) Preliminary field tests on the use of *Metarhizium anisopliae* for the control of *Nasutitermes exitiosus* (Hill) (Isoptera, Termitidae). *Bulletin of Entomological Research*, 73(2),305-313.

Hanel, H.(1983) Biological control of termites with the fungus *Metarhizium anisopliae*. *Holz-Zentralblatt*, 109(33),488-489.

Hanula, J. L and Andreadis, T. G.(1988) Parasitic microorganisms of Japanese beetle (Coleoptera,

Scarabaeidae) and associated scarab larvae in Connecticut soils. *Environmental Entomology*, 17(4),709-714.

Haque, J and Ghosh, A. B. (2007) Effect of *Beauveria bassiana* (Bals.) Vuill, *Metarhizium anisopliae* (Metsch.) Sorokin and Nimbecidine on some insect pests. *Environment and Ecology*; 25(1),209-211.

He, XueYou, Huang, JinShui, Cai, ShouPing, Yang, Xi Yu., Pei,Wang and Chen, ShunLi (2009) Survival dynamics of *Metarhizium anisopliae* on cadaver of *Monochamus alternatus* adults. *Scientia Silvae Sinicae*, 45(12),77-82.

He ,XueYou., Chen, ShunLi ,,Yang, Xi Huang., JinShui, Huang ,,BingRong, Cai and ShouPing. (2007) The investigation of *Metarhizium anisopliae* in forest soil in Fujian and Jiangxi provinces and pathogenicity against *Monochamus alternatus*. *Mycosystema*; 26(2),289-294.

He, XueYou, Chen, ShunLi and Huang, JinShui (2005) Preliminary screening of virulent strains of *Metarhizium anisopliae* against *Monochamus alternatus*. *Acta Entomologica Sinica*, 48(6), 975-981.

He Xue, You., Cai ,ShouPing., Yu ,PeiWang., Huang, JinShui., Zhong , JingHui., Chen ,DeLan and Xiong, Yu. (2008) Virulence of a strain of *Metarhizium anisopliae*, MaYTTR-04, against adults of *Monochamus alternatus*. *Acta Entomologica Sinica*, 51(1),102-107.

Heinz, K.M., Zalom, F.G., (1996) Performance of the predator *Delphastus pusillus* on *Bemisia* resistant and susceptible tomato lines. *Entomol. Exp.* Appl. 81, 345–352.

Herrmann, F and Wegensteiner, R.(2011)Infecting *Ips typographus* (Coleoptera, Curculionidae) with *Beauveria bassiana*, *Metarhizium anisopliae or Isaria fumosorosea* (Ascomycota). *IOBC/WPRS Bulletin*, 66, 209-212.

Hidalgo-Salvatierra and O. Berrios, F.(1972) Studies on the shootborer *Hypsipyla grandella Zeller* (Lep., Pyralidae). XI. Growth of larvae reared on a synthetic diet. XII. Determination of the LC50 of *Metarrhizium anisopliae* (Metchnikoff) Sorokin spores on fifth instar larvae. *Turrialba*, 22(4),431-434, 435-438.

Hidalgo-Salvatierra, O and Berrios, F.(1972)Studies on the shootborer *Hypsipyla grandella* Zeller. (Lep., Pyralidae). XII. Determination of the LC50 of *Metarrhizium anisopliae* (Metchnikoff) Sorokin spores on fifth instar larvae. *Turrialba*, 22(4),435-438.

Hoe, PikKheng, Bong, C.F.J., Kadir, Jugah and Amartalingam, Rajan. (2009) Evaluation of *Metarhizium anisopliae* var. anisopliae (Deuteromycotina, Hyphomycete) isolates and their effects on subterranean termite *Coptotermes curvignathus* (Isoptera, Rhinotermitidae). *American Journal of Agricultural and Biological Sciences*, 4(4), 289-297.

Houle, C., Hartmann, G. C and Wasti, S.S. (1987) Infectivity of eight species of entomogenous fungi to the larvae of the elm bark beetle, *Scolytus multistriatus* (Marsham). *Journal of the New York Entomological Society*, 95(1), 14-18.

Howard, A. F. V.(2010)Natural products for malaria vector control, flora, fish and fungi. *Natural products for malaria vector control, flora, fish and fungi*, 267 pp.

Hunter, D. M., Milner, R. J., Scanlan, J. C and Spurgin, P. A.(1999) Aerial treatment of the migratory locust, *Locusta migratoria* (L.) (Orthoptera, Acrididae) with *Metarhizium* 

anisopliae (Deuteromycotina, Hyphomycetes) in Australia. Crop Protection, 18(10), 699-704.

Hussain, A and Tian MingYi (2013) Germination pattern and inoculum transfer of entomopathogenic fungi and their role in disease resistance among *Coptotermes formosanus* (Isoptera, Rhinotermitidae).*International Journal of Agriculture and Biology*, 15(2),319-324.

Hussain, A., Ruan, Lin Tian., Ming,Yi He and Yu,Rong. (2009) Pathogenic effect of *Metarhizium anisopliae* on the larval growth and development of *Ocinara varians* Walker (Lepidoptera, Bombycidae). *Pakistan Entomologist*, 31(2), 116-121.

Hussain, A., Tian, Ming Yi., He, YuRong, Bland, J.M and Gu, WenXiang (2010) Behavioral and electrophysiological responses of *Coptotermes formosanus* Shiraki towards entomopathogenic fungal volatiles. *Biological Control*, 55(3),166-173.

Ihara, F., Toyama, M and Sato, T. (2003) Pathogenicity of *Metarhizium anisopliae* to the chestnut weevil larvae under laboratory and field conditions. *Applied Entomology and Zoology*, 38(4), 461-465.

Ihara, F., Toyama, M., Higaki, M., Mishiro, K and Yaginuma, K.(2009) Comparison of pathogenicities of *Beauveria bassiana* and *Metarhizium anisopliae* to chestnut pests. *Applied Entomology and Zoology*, 44(1), 127-132.

Jaydeep, Halder, Rai, A.B and Kodandaram, M. H. (2013) Compatibility of neem oil and different entomopathogens for the management of major vegetable sucking pests. *National Academy Science Letters*, 36(1),19-25.

Jiang, Ying Cheng (2000) Comparison of infectivity of *Metarhizium anisopliae* and *Beauveria bassiana* to *Dendrolimus punctatus. Journal of Zhejiang Forestry College*, 17(4), 410-413.

Jones, W.E., Grace, J.K and Tamashiro, M. (1996) Virulence of seven isolates of *Beauveria bassiana* and *Metarhizium anisopliae* to *Coptotermes formosanus* (Isoptera, Rhinotermitidae). *Environmental Entomology*, 25(2), 481-487.

Kalidas, P and Prasad, K.V.R. (2001) Efficacy of green muscardine fungus *Metarhizium anisopliae* and its commercial multiplication on maize grains. *International Journal of Oil Palm Research*, 2(2), 51-53.

Kalita, H., Ramesh, K., Rahman, H and Panda, P. K.(2009)Bioefficacy of some biopesticides against insect pests of rice in Sikkim. *Indian Journal of Entomology*, 71(2),168-169.

Keller, S., Epper, C and Wermelinger, B.(2004) *Metarhizium anisopliae* as a new pathogen of the spruce bark beetle *Ips typographus.Mitteilungen der Schweizerischen Entomologischen Gesellschaft,* 77(1/2), 121-123.

Kramm, K.R. and West, D. F. (1982)Termite pathogens, effects of ingested *Metarhizium*, *Beauveria*, and *Gliocladium* conidia on worker termites (*Reticulitermes* sp.). *Journal of Invertebrate Pathology*, 40(1),7-11.

Kramm, K. R., West, D. F and Rockenbach, P. G.(1982)Termite pathogens, transfer of the entomopathogen *Metarhizium anisopliae* between *Reticulitermes* sp. termites. *Journal of Invertebrate Pathology*, 40(1),1-6.

Krassilstschik, *I. M.* (1888)La production industrielle des parasites vegetaux pour la destruction des insects nuisibles. *Bull. Sci. France et Belg.* 19, 461–472.

Krishna Japur, Prabhu, S. T., Balikai, R. A and Mallapur, C. P. (2012) Management of rice blue beetle (Leptispa pygmaea) under rainfed ecosystem using botanicals, entomopathogenic fungi and new insecticides. *Journal of Experimental Zoology, India*, 15(2),655-659.

Krishna Kant Ramanujam, B., Tyagi, S. K., Sharma, Y. K., Meena, S. S., Mishra, B. K., Vishal, M. K and Meena, S. R. (2013)Management of fennel aphids (*Hyadaphis coriandri Das*) through biorational approaches. *Annals of Plant Protection Sciences*, 21(1),21-23.

Kucera, M.(1980) Proteases from the fungus *Metarhizium* anisopliae toxic for *Galleria mellonella* larvae. *Journal of Invertebrate Pathology*, 35(3),304-310.

LaceyL. A., Frutos R., Kaya H. K and Vail P.(2001)Insectpathogens as biological control agents, do they have a future?. *Biological Control*, 21, 230-248.

Landis, D.A., Wratten, S.D., Gurr, G.M., (2000) Habitat management to conserve natural enemies of arthropod pests in agriculture. *Annu. Rev. Entomol.* 45, 175–201.

Leite, M. S. P. E. T., Penteado, S., do R. C., Zaleski, S. R. M., Camargo, J. M. M and Ribeiro, R. D.(2011)Compatibility of the insect pathogenic fungus *Beauveria bassiana* with neem against sweetpotato whitefly, Bemisia tabaci, on eggplant. *Floresta*, 41(3),619-628.

Lenz, M.(2005)Biological control in termite management, the potential of nematodes and fungal pathogens. *Fifth International Conference on Urban Pests*, *Singapore*, 11-13 July 2005, 47-52.

Li, L. Z., Zhou, X. S., Yan, J., Cui, Y. S and Yang, H. P. (1992) Studies on the biological control technique for the white grub *Blitopertha pallidipennis* Reitter at the Bureau of Shangganling. *Collection of achievements on the technique cooperation project of P. R. China and F. R.Germany*, 136-140.

Loureiro, E., de S and Monteiro, A. C.(2005)Pathogenicity of isolates of three entomopathogenic fungi against soldiers of *Atta sexdens* (*Linnaeus*, 1758)(Hymenoptera, Formicidae). Revista Arvore, 29(4),553-561.

Maketon, M., Sawangwan, P and Sawatwarakul, W.(2007)Laboratory study on the efficacy of *Metarhizium anisopliae* (Deuteromycota, Hyphomycetes) in controlling *Coptotermes gestroi* (Isoptera, Rhinotermitidae). *Entomologia Generalis*, 30(3),203-218.

Maketon, M., Orosz-Coghlan, P and Sinprasert, J. (2008) Evaluation of *Metarhizium anisopliae* (Deuteromycota, Hyphomycetes) for control of broad mite *Polyphagotarsonemus latus* (Acari, Tarsonemidae) in mulberry. (Diseases of mites and ticks.) *Experimental and Applied Acarology*, 46(1/4),157-167.

Mendes, A. C. B., Magalhaes, B. P., Ohashi, O. S and Bastos, C. N. (2001) Infection of Conotrachelus humeropictus Fiedler (Coleoptera, Curculionidae) by *Metarhizium anisopliae* (Metsch.) Sor. and *Beauveria bassiana* (Bals.) Vuill. in the soil. [Portuguese] *Acta Amazonica*, 31(4),531-538.

Meyling N.Vand Eilenberg J. (2007) Ecology of the entomopathogenic fungi Beauveria bassiana and

Metarhizium anisopliae in temperate agroecosystems, Potential for conservation biological control. Biological Control. 43, 145–155.

Michael Wyrebek, Cristina Huber, Ramanpreet Kaur Sasan and Michael J. Bidochka (2011) Three sympatrically occurring species of *Metarhizium* show plant rhizosphere specificity. *Microbiology* 157, 2904–2911.

Milner, R. J., Staples, J. A and Lenz, M.(1996)Options for termite management using the insect pathogenic fungus *Metarhizium anisopliae.Document - International Research Group on Wood Preservation*, (IRG/WP/96-10142),5 pp.

Milner, R. J., Staples, J. A and Lutton, G. G. (1997) The effect of humidity on germination and infection of termites by the hyphomycete, *Metarhizium anisopliae*. *Journal of Invertebrate Pathology*, 69(1),64-69.

Milner, R. J., Staples, J. A and Lutton, G. G. (1998) The selection of an isolate of the hyphomycete fungus, *Metarhizium anisopliae*, for control of termites in Australia. *Biological Control*, 11(3),240-247.

Milner, R. J.(2000) Improved formulations of *Metarhizium* for biological control of termites. *Technical Report - CSIRO Division of Entomology*, (86), 37 + 23 + 9 + 17 + 2 + 2 pp.

Miscellaneous. (1992) Studies on the effect of growth inhibitors, active plant constituents and pathogens on larvae and adults of the desert locust *Schistocerca gregaria*. Results of the field and laboratory trials in Agades and Anou Mekkerene, *North Niger* 1991. iii, 55 pp.

Mohi-Uddin, S., Munazah Yaqoob Ahmed, M. D. J and Ahmed, S. B.(2009) Management of apple stem borer, *Aeolesthes sarta* (Coleoptera, Cerambycidae) in Kashmir. *Environment and Ecology*, 27(2A), 931-933.

Moino Junior, A., Alves, S. B., Lopes, R. B., Oliveira, P. M., Neves, J., Pereira, R. M and Vieira, S. A.(2002)External development of the entomopathogenic fungi *Beauveria bassiana* and *Metarhizium anisopliae* in the subterranean termite *Heterotermes tenuis. Scientia Agricola*, 59(2), 267-273.

Montermini, A., Cortellini, W and Deseo, K. V. (1985) Microbiological control of *Hyphantria cunea* Drury (Lepidoptera, Arctiidae) in North Italy. *Difesa delle Piante*, 8(2),345-351.

Moorhouse, E. R., Easterbrook, M. A., Gillespie, A. T and Charnley, A. K.(1993)Control of *Otiorhynchus sulcatus* (Fabricius) (Coleoptera, Curculionidae) larvae on a range of hardy ornamental nursery stock species using the entomogenous fungus *Metarhizium anisopliae.Biocontrol Science and Technology*, 3(1),63-72.

Morales Romero, L., Grillo Ravelo, H., Maza Estrada, N and Grau, R. (2012) Effectiveness of entomopathogenic fungi in the management of *Pseudacysta perseae* (Heid.) (Hemiptera, Tingidae) in avocado (Persea americana Mill.). [Spanish] *Revista Cientifica UDO Agricola*, 12(3),599-608.

Moslim, R., Wahid, M. B. ,Kamarudin, N., Mukesh Sharma and Ali, S. R. A. (1999) Impact of *Metarhizium anisopliae* (Deuteromycotina, Hyphomycetes) applied by wet and dry inoculum on oil palm rhinoceros beetles,

Oryctes rhinoceros (Coleoptera, Scarabaeidae). *Journal of Oil Palm Research*, 11(2),25-40.

Mudroncekova, S., Mazan, M., Nemcovic, M and Salamon, I.(2013)Entomopathogenic fungus species *Beauveria bassiana* (Bals.) and *Metarhizium anisopliae* (Metsch.) used as mycoinsecticide effective in biological control of *Ips typographus* (L.). *Journal of Microbiology, Biotechnology and Food Sciences*, 2(6),2469-2472.

Mukesh, Nitharwal., Kumawat, K. C and Meenu Choudhary Jat, R. G.(2013)Influence of biorational and conventional insecticides on the population of *Chrysoperla carnea* (Steph.) in green gram, *Vigna radiata* (Linn.) in semi-arid conditions. *Biopesticides International*,9(1),83-87.

Murthy, H. C., Naik, M. I and Manjunath, M.(2010)Evaluation of certain new chemicals, bio-agents and plant product for the management of arecanut root grub, (*Leucopholis lepidophora*) Blanch. Mysore Journal of Agricultural Sciences; 44(3),653-655.

Nagaraju, K., Sundararaj, R and Meenakshi, B. C.(2013)Entomopathogenic fungus of termites and their potential in management of forest nurseries and plantations. *Journal of Pure and Applied Microbiology*, 7(1), 631-635.

Nedveckyte, I., D. Dirginciute-Volodkiene, V and Buda, V. (2011)Pine *defoliator Bupalus piniaria* (L.) (Lepidoptera, Geometridae) and its entomopathogenic fungi. 2. Pathogenicity of *Beauveria bassiana*, *Metarhizium anisopliae* and *Isaria farinosa*. *Ekologija*,57(1),12-20.

Neelima, S., Rao, G. M. V. P., Chalam, M. S. V and Grace, A. D. G.(2011)Bio-efficacy of ecofriendly products against cotton leafhopper, *Amrasca devastans* (Dist.).*Annals of Plant Protection Sciences*, 19(1),15-19.

Nielsen, C., Vestergaard, S., Harding, S and Eilenberg, J.(2007) Microbial control of *Strophosoma* spp. larvae (Coleoptera, Curculionidae) in Abies procera greenery plantations. *Journal of Anhui Agricultural University*, 34(2),185-194.

Nielsen, C., Vestergaard, S., Harding, S., Wolsted, C and Eilenberg, J.(2006) Biological control of *Strophosoma* spp. (Coleoptera, Curculionidae) in greenery (Abies procera) plantations using Hyphomycetes. *Biocontrol Science and Technology*, 16(5/6),583-598.

Nyam, VeiTing, Bong, C. F. J and King, J. H. P. (2015) Control of subterranean termite *Coptotermes curvignathus* (Isoptera, rhinotermitidae) by entomopathogen *Metarhizium anisopliae* var. Anisopliae cultured in liquid state fermentation. *American Journal of Agricultural and Biological Sciences*, 10(1),35-40.

Nyeko, P., Gohole, L. S., Maniania, N. K., Agaba, H and Sekamatte, B. M.(2010) Evaluation of *Metarhizium anisopliae* for integrated management of termites on maize and *Grevillea robusta* in Uganda and Kenya. (RUFORUM Working Document Series No.5)Second *RUFORUM Biennial Regional Conference on "Building capacity for food security in Africa", Entebbe*, Uganda, 20-24, 2010, 333-337.

Oddsdottir, E. S., Eilenberg, J., Sen, R., Harding, S and Halldorsson, G.(2010) Early reduction of *Otiorhynchus* spp. larval root herbivory on *Betula pubescens* by beneficial soil fungi. *Applied Soil Ecology*, 45(3),168-174.

- Oliveira, M. A. S., Alves, R. T., Fialho, J., de F and Junqueira, N. T. V. (2001)Pathogenicity of the entomopathogenic fungi on lace bug on cassava in Federal District. [Portuguese] *Comunicado Tecnico Embrapa Cerrados*, (45), 2 pp.
- Ouakid, M. L., Farine, J. P and Soltani, N. (2005) Activity evaluation of a local strain of the entomopathogenous fungus *Metarhizium anisopliae* on Lymantria dispar larvae. *Bulletin OILB/SROP*, 28(8), 185-191.
- Outtar, F., Doumandji-Mitiche, B., Mouhouche, F and Doumandji, S. (2011) Alternative method against locusts. 4eme Conference Internationale sur les Methodes Alternatives en Protection des Cultures. Evolution des cadres reglementaires europeen et francais. Nouveaux moyens et strategies Innovantes, Nouveau Siecle, Lille, France, 8-10, 197-204.
- Pabst, G. S and Sikorowski, P. P.(1980) Susceptibility of southern pine beetle (Dendroctonus frontalis) on oligidic medium to *Paecilomyces viridis* and also *Beauveria bassiana*, and *Metarhizium anisopliae.Journal of the Georgia Entomological Society*, 15(3),235-240.
- Padmaja, V., Bhanuprakash, G. V. S., Shanmukh anand, P.,Sujatha, K. and Gurvinder K. (2005)Evaluation of *Metarhizium anisopliae* (Metsch) Sorokin for the management of *Spodoptera litura* on beans crop. In, Sustainable insect pest management. (eds.) Ignacimuthu, S.J. and Jayaraj, S. 136-139 pp.
- Pan, Yong Sheng., Xu ,FuYuan and Han Zheng Min. (2012) Studies on the screening of high virulent *Metarhizium anisopliae* against *Clostera anachoreta* larva. *Journal of Jiangsu Forestry Science & Technology*, 39(5),13-15, 44.
- Pan, Yong Sheng., Xu, FuYuan, Han, ZhengMin and Liu YunPeng. (2010)Studies on the screening and liquid shake culture condition of high virulent *Metarhizium anisopliae* against *Monochamus alternatus* Hope adults. *Forest Research*, Beijing, 23(1), 102-107.
- Pan, Rong Ying., She, ChunRen and Cai MeiLan (1995) Tests of virulence of *Metarhizium anisopliae* var. anisopliae to *Pseudophacopteron canarium* Yang et Li. *Journal of Fujian Agricultural University*, 24(3), 304-306. Patil, G.R. and Naik, L.K. (2004) Evaluation of mycopathogens against *Helopeltis antonii* Signoret. *Karnataka Journal of Agricultural Sciences*, 17(2), 337-338
- Pegu, J. R., Pranab Dutta Puzari, K. C and Nath, P. D.(2013) Natural incidence of entomopathogenic fungi in North East India. *Journal of Mycology and Plant Pathology*, 43(2), 243-245.
- Pehl, L and Kehr, R.(1994) Biological control of bark beetles, preparations with the fungus *Metarhizium anisopliae*. *AFZ*, *Allgemeine Forst Zeitschrift*, 49 (19), 1065-1067.
- Phukon, M., Sarma, I., Borgohain, R., Sarma, B and Goswami, J. (2014) Efficacy of *Metarhizium anisopliae*, *Beauveria bassiana* and neem oil against tomato fruit borer, *Helicoverpa armigera* under field condition. *Asian Journal of Bio Science*, 9(2),151-155.
- Pires, L. M., Marques, E. J., Oliveira, J. V and de Alves, S. B. (2010) Selection of isolates of entomopathogenic fungi for controlling *Tuta absoluta* (Meyrick) (Lepidoptera, Gelechiidae) and their compatibility with insecticides used in tomato crop. *Neotropical Entomology*, 39(6),977-984.

- Pogetto, M. H. F. A., dal Wilcken, C. F., Christovam, R. S., Prado, E. P and Gimenes, M. J.(2011) Effect of formulated entomopathogenic fungi on red gum lerp psyllid *Glycaspis brimblecombei*. *Research Journal of Forestry*, 5(2),99-106.
- Posos Ponce, P., Martinez Ramirez, J. L., Carreon Amaya, J., Serratos Arevalo, J. C., Peraza Luna, F. A and Guerrero Rodriguez, E.(2004)Susceptibility of *Cyclocephala comata* Bates (Coleoptera, Sacarabaeidae) to different biopesticides. *Resistant Pest Management Newsletter*, 13(2),20-22.
- Prado, E. (1980) Control of black vine weevil larvae (*Otiorhynchus sulcatus*) using the insect-pathogenic fungi Beauveria bassiana, *Metarrhizium anisopliae* and *Metarrhizium flavoviride*. *Vaxtskyddsnotiser*, 44(6), 160-167.
- Premalatha, K., Rajavel, D. S and Baskaran, R. K. M. (2008) Studies on management of sugarcane termite, *Odontotermes obesus* (Rambur). *Indian Sugar*, 57(10),17-21.
- Prishchepa, L., Mikulskaya, N and Sosnowska, D. (2005) Biological activity of entomopathogens derived from Bialowieza Forest. *Progress in Plant Protection*, 45(2), 1012-1014.
- Quarles, W. (1999) Commercial biocontrol for termites. *IPM Practitioner*, 21(10),1-6.
- Rachappa, V. Lingappa, S. Patil, R. K. Tipannavar, P. S. (2004) Utilization of *Metarhizium anisopliae* (Metch.) sorokin for the management of sugarcane rootgrub. *Indian Sugar*, 54(2), 111-115.
- Raja Rishi, R., Borah,R.K., Rajesh Kumar and Shailesh Pandey. (2013) Isolation, identification and mass production of soil microbes and their utility for biocontrol. International Journal of Advanced Life Sciences (IJALS). 6(3), 168-173.
- Rakesha, H. S., Prabhu, S. T and Balikai, R. A.(2012) Laboratory evaluation of fungal pathogens and plant extracts against arecanut root grub, *Leucopholis lepidophora*blanchard. *Journal of Experimental Zoology, India*, 15(2),463-465.
- Ramakrishnan, R., Suiter, D. R., Nakatsu, C. H., Humber, R. A and Bennett, G. W.(1999) Imidacloprid-enhanced *Reticulitermes flavipes* (Isoptera, Rhinotermitidae) susceptibility to the entomopathogen *Metarhizium anisopliae.Journal of Economic Entomology*, 92(5),1125-1132.
- Ramanuj, Vishwakarma. Pool, Chand and Ghatak, S. S. (2011) Potential plant extracts and entomopathogenic fungi against red pumpkin beetle, *Raphidopalpa foveicollis* (Lucas). *Annals of Plant Protection Sciences*, 19(1), 84-87.
- Ramle , Moslim., Norman, Kamarudin and Mohd Basri Wahid. (2009) Pathogenicity of granule formulations of *Metarhizium anisopliae* against the larvae of the oil palm rhinoceros beetle, *Oryctes rhinoceros* (L.). *Journal of Oil Palm Research*, 21,602-612.
- Ramle, Moslim and Kamarudin, N.(2014)The use of palm kernel cake in the production of conidia and blastospores of *Metarhizium anisopliae* var. major for control of *Oryctes rhinoceros. Journal of Oil Palm Research*, 26(1),133-139.
- Ramle, Moslim, Kamarudin, N, Na AngBan, Siti Ramlah, A. A and Mohd Basri Wahid. (2007) Application of

powder formulation of *Metarhizium anisopliae* to control *Oryctes rhinoceros* in rotting oil palm residues under leguminous cover crops. *Journal of Oil Palm Research*, 19,319-331.

Ramle, Moslim., Norman, Kamarudin and Mohd Basri Wahid. (2011)Trap for the auto dissemination of *Metarhizium anisopliae* in the management of rhinoceros beetle, Oryctes rhinoceros. *Journal of Oil Palm Research*, 23(1),1011-1017.

Ranaivo, F., Welling, M., Zimmermann, G and Schmutterer, H. (1996) Fitness reduction by the African migratory locust, *Locusta migratoria*, after application of low concentrations of *Metarhizium flavoviride* blastospores and neem oil. *Bulletin OILB/SROP*, 19(9), 236-239.

Remadevi, O. K., Sasidharan, T. O.,Balachander, M and Bai, N. S. (2010) *Metarhizium* based mycoinsecticides for forest pest management. *Journal of Biopesticides*, 3(2),470-473.

Rethinam, P and Singh, S. P. (2007) Current status of the coconut beetle outbreaks in the Asia-Pacific region. RAP Publication, 2,1-23.

Rishi, R. R., Barthakur, N. D and Borah, R. K.(2012) Evaluation of the entomopathogenic fungi *Beauveria bassiana* (Bals.) Vuill., and *Metarhizium anisopliae* (Metsch.) for the control of Bamboo Leaf Roller *Pyrausta coclesalis* Wlk. (Pyralidae, Lepidoptera). *International Journal of Advanced Life Sciences (IJALS)*,5(1),12-18.

Rumine,P and Barzanti, G. P. (2009) Microbiological control of the leaf-footed bug Leptoglossus occidentalis. *IOBC/WPRS Bulletin*, 45,325-326.

Sabbour, M. Mand E-Abd-El-Aziz, S. (2007) Efficiency of some bioinsecticides against broad bean beetle, *Bruchus rufimanus* (Coleoptera, Bruchidae). *Research Journal of Agriculture and Biological Sciences*, 3(2),67-72.

Sabbour, M. (2012) Evaluations of some bioagents against *Sitophilus oryzae* under laboratory and store conditions. *IOBC/WPRS Bulletin*, 81,135-142.

Sahayaraj, K and Borgio, J. F.(2009) Distribution of *Metarhizium anisopliae* (Metsch.) Sorokin (Deuteromycotina, Hyphomycetes) in Tamil Nadu, India, its biocontrol potential on *Dysdercus cingulatus* (Fab.) (Hemiptera, Pyrrhocoridae). *Archives of Phytopathology and Plant Protection*, 42(5), 424-435.

Sahu, K. R and Sharma, D. (2008) Management of cashew stem and root borer, *Plocaederus ferrugineus* L. by microbial and plant products. *Journal of Biopesticides*, 1(2),121-123.

Sajap, A. S and Kaur, K. (1990) Histopathology of *Metarhizium anisopliae*, an entomopathogenic fungus infection on the termite, *Coptotermes curvignathus*. *Pertanika*, 13(3),331-334.

Sakchoowong, W. (2002) Effects of entomopathogenic fungi (*Beauveria bassiana* and *Metarhizium anisopliae*) on teak defoliator (Hyblaea puera Cramer, Lepidoptera, Hyblaeidae) in laboratory. *FORSPA Publication*, (30),105-110

Sanchez-Pena, S. R., San-Juan Lara, J and Medina, R. F. (2011) Occurrence of entomopathogenic fungi from agricultural and natural ecosystems in Saltillo, Mexico, and their virulence towards thrips and whiteflies. *Journal of Insect Science (Madison)*, 11, Article 1.

Santiago-Alvarez, C., Santos-Quiros, R., Valverde-Garcia, P and Quesada-Moraga, E. (2005) Selection of entomopathogenic Anamorphic fungi isolates for the control of *Reticulitermes grassei* (Isoptera, Rhinotermitidae). *Boletin de Sanidad Vegetal, Plagas*, 31(2),299-307.

Sapna-Bai, N., Remadevi, O. K., Sasidharan, T. O., Balachander, M and Dharmarajan, P.(2013) Bioefficacy of *Metarhizium anisopliae* isolates against teak skeletoniser Paliga machoeralis (Lepidoptera, Pyralidae). *Journal of Tropical Forest Science*, 25(3),310-316.

Schabel, H. G. (1976) Oral infection of Hylobius pales by *Metarrhizium anisopliae*. *Journal of Invertebrate Pathology*, 27(3), 377-383.

Sevim, A., Demir, I., Sonmez, E., Kocacevik, S and Demirbag, Z. (2013) Evaluation of entomopathogenic fungi against the sycamore lace bug, *Corythucha ciliata* (Say) (Hemiptera, Tingidae). *Turkish Journal of Agriculture and Forestry*, 37(5),595-603.

Shah, F. A., Ansari, M. A., Prasad, Man Butt, T. M. (2007) Evaluation of black vine weevil (*Otiorhynchus sulcatus*) control strategies using *Metarhizium anisopliae* with sublethal doses of insecticides in disparate horticultural growing media. *Biological Control*, 40(2), 246-252.

Shah, F.A., Gaffney, M., Ansari, M. A., Prasad, Man Butt, T. M. (2008) Neem seed cake enhances the efficacy of the insect pathogenic fungus *Metarhizium anisopliae* for the control of black vine weevil, *Otiorhynuchs sulcatus* (Coleoptera, Curculionidae). *Biological Control*,44(1), 111-115.

Shamila Kalia and Harsh, N. S. K.(2003) *Metarhizium anisopliae* (Metschnikoff) Sorokin pathogenic to the larvae of teak defoliator, *Hyblaea puera* Cramer. *Journal of Entomological Research*, 27(2), 135-136.

Shanley, R. P and Hajek, A. E .(2008)Environmental contamination with *Metarhizium anisopliae* from fungal bands for control of the Asian longhorned beetle, *Anoplophora glabripennis* (Coleoptera, Cerambycidae). *Biocontrol Science and Technology*, 18(1/2), 109-120.

Shapiro-Ilan, D. I., Cottrell, T. E., Gardner, W. A., Behle, R. W., Ree, B and Harris, M. K.(2009)Efficacy of entomopathogenic fungi in suppressing pecan weevil, *Curculio caryae* (Coleoptera, Curculionidae), in commercial pecan orchards. *Southwestern Entomologist* 34 (2), 111-120.

Shapiro-Ilan, D. I., Cottrell, T. E., Gardner, W. A., Leland, J and Behle, R. W. (2009) Laboratory mortality and mycosis of adult *Curculio caryae* (Coleoptera, Curculionidae) following application of *Metarhizium anisopliae* in the laboratory or field. *Journal of Entomological Science*, 44(1), 24-36.

Shashi, Sharma, Gupta, R.B.L and Yadava, C.P.S. (1999) Effect of certain soil fungi on *Metarhizium* and *Beauveria* spp. and their pathogenicity against *Holotrichia consanguinea*. *Indian Phytopathology*, 52(2),196-197.

Shimazu, M., Mitsuhashi, W., Hashimoto, H., Ozawa, T.(1993) Persistence of *Metarhizium anisopliae* (Deuteromycotina, Hyphomycetes) as a control agent of *Anomala cuprea* (Coleoptera, Scarabaeidae) in a forestry nursery. *Applied Entomology and Zoology*, 28(1), 103-105.

- Shimizu, S and Yamaji, M. (2003) Effect of density of the termite, *Reticulitermes speratus* Kolbe (Isoptera, Rhinotermitidae), on the susceptibilities *to Metarhizium anisopliae.Applied Entomology and Zoology*, 38(1),125-130.
- Silva, E. A. R., Batista Filho, A., Wenzel, I. M., Furtado, E. L and Almeida, J. E. M.(2012) Selection of entomopathogenic fungi strains to Leptopharsa heveae (Hemiptera, Tingidae) control. *Arquivos do Instituto Biologico (Sao Paulo)*, 79(4), 549-556.
- Silva, M. E and da Diehl-Fleig, E. (1988)Evaluation of different strains of entomopathogenic fungi for the control of the ant *Attasexdens piriventris* (Santschi, 1919) (Hymenoptera, Formicidae). *Anais da Sociedade Entomologica do Brasil*, 17(2), 263-269.
- Silveira, R. D., Anjos, N and dos Zanuncio, J. C. (2002)Natural enemies of *Coelomera lanio* (Coleoptera, Chrysomelidae) in the region of Vicosa, Minas Gerais, Brazil. *Revista de Biologia Tropical*, 50(1),117-120.
- Sivapragasam, A and Tey, C. C.(1994)Susceptibility of *Oryctes rhinoceros* larvae (L.) to three isolates of *Metarhizium anisopliae* (Metsch) Sorokin. *MAPPS Newsletter*, 18(2), 13-14.
- Sohail Ahmed, Ashraf, M. R., Abid, Hussain and Riaz, M. A.(2009) Pathogenicity of isolates of *Metarhizium anisopliae* from Gujranwala (Pakistan) against *Coptotermes heimi* (Wasmann) (Isoptera, Rhinotermitidae). *International Journal of Agriculture and Biology*, 11(6),707-711.
- Sohail, Ahmed ., Ashraf, M. R., Hussain, M. A and Riaz, M. A. (2008) Pathogenicity of isolates of *Metarhizium anisopliae* from Murree (Pakistan) against *Coptotermes heimi* (Wasmann) (Isoptera, Rhinotermitidae) in the laboratory. *Pakistan Entomologist*, 30(2),119-125.
- Sohail Ahmed, Ashraf, M. Rand Hussain, M. A. (2008) Pathogenicity of a local strain of *Metarhizium anisopliae* against *Coptotermes heimi* (Was.) (Isoptera, Rhinotermitidae) in the laboratory. *Pakistan Entomologist*, 30(1),43-50.
- Solaiman, R. H. A and El-Latif, N. A. A.(2014) Isolation and pathogenicity of the fungus *Metarhizium anisopliae* (Metschnikoff) against the subterranean termite, *Psammotermes hybostoma* Desneux (Isoptera, Rhinotermitidae). *Egyptian Journal of Biological Pest Control*, 24(2),329-334.
- Song, Zhang., Lu, FengMei and Chen Hui. (2002)Studies on laboratory infection of *Metarhizium anisopliae* to *Pantana phyllostachysae*. *Journal of Fujian College of Forestry*, 22(2), 120-123.
- Staples, J.A. and Milner, R.J. (2000) A laboratory evaluation of the repellency of *Metarhizium anisopliae* conidia to *Coptotermes lacteus* (Isoptera, Rhinotermitidae) Sociobiology, 36(1),133-148.
- Steinhaus, E. A.(1975) Disease in a Minor Chord. Ohio State University Press, Columbus, Ohio.
- Stenzel, K., Holters, J., Andersch, W and Smit, T. A. M.(1992) BIO 1020, granular *Metarhizium* a new product for biocontrol of soil pests. *Proceedings, Brighton Crop Protection Conference, Pests and Diseases, 1992 Brighton*, 363-368.
- Stephan, D and Herker, M.(2011) Artificial hideouts with entomopathogenic fungi, a strategy for biological control

- of the plum fruit moth *Cydia funebrana/IOBC/WPRS Bulletin*, 66,175-178.
- Sterling C. A. Gomez M., C. A and Campo J. A. A.(2011) Pathogenicity of *Metarhizium anisopliae* (Deuteromycota, Hyphomycetes) on *Heterotermes tenuis* (Isoptera, Rhinotermitidae) in *Hevea brasiliensis. Revista Colombiana de Entomologia*, 37(1), 36-42.
- Subramaniam, Gopalakrishnan., Rao, G. V. R., Pagidi Humayun Rao, V. R., Gottumukkala ,Alekhya., Simi, Jacob. Kanala, Deepthi, Vidya.M. S., Srinivas, V., Linga Mamatha and Om Rupela (2011) Efficacy of botanical extracts and entomopathogens on control of *Helicoverpa armigera* and *Spodoptera litura*. *African Journal of Biotechnology*, 10(73), 16667-16673.
- Sun, J. Z., Fuxa, J. R and Henderson, G. (2003) Effects of virulence, sporulation, and temperature on *Metarhizium anisopliae* and *Beauveria bassiana* laboratory transmission in *Coptotermes formosanus*. *Journal of Invertebrate Pathology*, 84(1), 38-46.
- Sun, J. Z., Fuxa, J. R., Richter, A an Ring, D. (2008) Interactions of *Metarhizium anisoplae* and tree-based mulches in repellence and mycoses against *Coptotermes formosanus* (Isoptera, Rhinotermitidae). *Environmental Entomology*, 37(3),755-763.
- Sundararaj, R. (2014) Relevance of Botanicals for the Management of Forest Insect Pests of India. *Basic and Applied Aspects of Biopesticides*, 155-179 PP.
- Sunitha, V., Lakshmi, K. V and Rao, G. V. R.(2008) Laboratory evaluation of certain insecticides against pigeonpea pod borer, *Maruca vitrata* Geyer. *Journal of Food Legumes*, 21(2),137-139.
- Svedese, V. M., Silva, A. P., de A. P. da Lopes, R. da S., Santos, J. F., dos Lima, E. A and de L. A.(2012) Action of entomopathogenic fungi on the larvae and adults of the fig fly *Zaprionus indianus* (Diptera, Drosophilidae). *Ciencia Rural*, 42(11),1916-1922.
- Swaminathan, R and Suman Manjoo Hussain, T.(2010) Anti-feedant activity of some biopesticides on *Henosepilachna vigintioctopunctata* (F.) (Coleoptera, Coccinellidae). *Journal of Biopesticides*, 3(1),77-80.
- Tangthirasunun, N.,Poeaim, S., Soytong, K., Sommartya, P. and Popoonsak, S. (2010)Variation in morphology and ribosomal DNA among isolates of *Metarhizium anisopliae* from Thailand. Journal of Agricultural Technology . 6(2), 317-329.
- Tedders, W. L., Weaver, D. J and Wehunt, E. J.(1973) Pecan weevil, suppression of larvae with the fungi *Metarrhizium anisopliae* and *Beauveria bassiana* and the nematode Neoaplectana dutkyi. *Journal of Economic Entomology*, 66(3),723-725.
- Tey, C. C and Ho, C. T.(1995) Infection of *Oryctes rhinoceros* L. by application of *Metarhizium anisopliae* (Metsch.) Sorokin to breeding sites. *Planter*,71(837),563-567.
- Tol, R. W. H. M. van.(1993)Control of the black vine weevil (*Otiorhynchus sulcatus*) with different isolates of *Heterorhabditis* sp. and *Metarhizium anisopliae* in nursery stock. *Proceedings of the Section Experimental and Applied Entomology of the Netherlands Entomological Society*, 4,181-186.
- Tserodze, M and Murvandze, A. (2009) Effectivity of entomopathogenic fungi *Metarhizium anisopliae* against

the Fall webworm. Artvin Coruh Uiversitesi Orman Fakultesi Dergisi, 10(2),111-112.

Valdes, T. (1976) The pathogenicity of the fungus *Metarrhizium anisopliae* in larvae of *Galleria mellonella*. *CIARCO*, 4(3/4), 5-6.

Van Emden, H. F. (1965) The role of uncultivated land in the biology of crop pests and beneficial insects. *Scientific Horticulture* 17, 121-136.

Veen, K. H. (1968) Recherches sur la maladie, due a' *Metarrhizium anisopliae* croquet pe'lerin. Mededingen Landbouwhoge school Wageningen, Netherland 68, 1–77. Vey, A and Quiot, J. M.(1989)Cytotoxic effect in vitro and in the insect host of destruxins, cyclodepsipeptide toxin products from the entomopathogenic fungus *Metarhizium anisopliae*. *Canadian Journal of Microbiology*,35(11),1000-1008.

Vishwakarma, R., Prasad, P. H., Ghatak, S. S and Mondal, S.(2011) Bio-efficacy of plant extracts and entomopathogenic fungi against epilachna beetle, *Henosepilachna vigintioctopunctata* (Fabricius) infesting bottle gourd. *Journal of Insect Science*, 24(1),65-70.

Wang, C. L and Powell, J. E.(2004)Cellulose bait improves the effectiveness of *Metarhizium anisopliae* as a microbial control of termites (Isoptera, Rhinotermitidae). *Biological Control*, 30(2),523-529.

Wang, Da Yuan., FangFang, Huang ., DaZhuang Liu ., ChunYan ,Bi and HuaMing. (2010) Infection process of *Metarhizium anisopliae* in *Anoplophora glabripennis* larvae observed with transmission electron microscopy. *Scientia Silvae Sinicae*, 46(5), 113-115.

Wang, Peng Zhang., YaBo,Shu., JinPing, Deng., Shun, Wang and HaoJie.(2010) Virulence of *Metarhizium anisopliae* var. anisopliae to the larvae of *Melanotus cribricollis* (Coleoptera, Elateridae). *Chinese Journal of Biological Control*, 26(3),274-279.

Wasti, S. S and Hartmann, G. C. (1982) Susceptibility of gypsy moth larvae to several species of entomogenous

fungi. Journal of the New York Entomological Society, 90(2),125-128.

Wells, J. D., Fuxa, J. R and Henderson, G.(1995) Virulence of four fungal pathogens to *Coptotermes formosanus* (Isoptera, Rhinotermitidae). *Journal of Entomological Science*, 30(2),208-215.

Wright, M. S and Cornelius, M. L. (2012)Mortality and repellent effects of microbial pathogens on *Coptotermes formosanus* (Isoptera, Rhinotermitidae). *BMC Microbiology*, 12(291), (15 December 2012).

Wright, M. S., Raina, A. K and Lax, A. R. (2005) A strain of the fungus *Metarhizium anisopliae* for controlling subterranean termites. *Journal of Economic Entomology*, 98(5),1451-1458.

Xia, Cheng Run ,Ding, DeGui., Liu, Yun Peng, Chai, XinYi., Fan, MeiZhen., Li, ZengZhi and Ni Jian. (2005) Field trials against *Spondylis buprestoides* through combined use of non-woven fabric bands impregnated with *Metarhizium anisopliae* and an attractant. *Journal of Anhui Agricultural University*, 32(4),419-422.

Xu, Hua Chao., Wu, Hong Zhou and YunE Zhang Hui. (2002) Preliminary study on biological characteristics of Pleonomus canaliculatus and toxicity test of *Metarhizium anisopliae* on it. *Journal of Zhejiang Forestry College*, 19(2),166-168.

Ye, Bin Jiang, YingCheng, Lin, WenQing and Song Zhang (2005) The effects of *Metarhizium anisopliae* on the diversity of arthropod community in masson pine stand. *Journal of Fujian Agriculture and Forestry University* (Natural Science Edition), 34(2), 239-243.

Zimmermann, G. (1993) The entomopathogenic fungus *Metarhizium anisopliae* and its potential as a biocontrol agent. *Pestic. Sci.* 37, 375-379.

Zoberi, M. H. (1995) *Metarhizium anisopliae*, a fungal pathogen of *Reticulitermes flavipes* (Isoptera, Rhinotermitidae). *Mycologia*, 87(3), 354-359.