

© 2004 - 2018 Society For Science and Nature (SFSN). All rights reserved

www.scienceandnature.org

### DETECTION OF *GIARDIA DUODENALIS* IN SHEEP BY DIFFERENT LABORATORY METHODS

Hiba Ali Ganim Al-Sead & Haider Mohammed Ali Sadiq El-Rubaie Department of Parasitology, Veterinary College, Baghdad University, Baghdad, Iraq. \*Corresponding author: www.dr.hiba198@gmail.com

#### ABSTRACT

The study was conducted to estimate the prevalence of *Giardia duodenalis* in sheep in Baghdad city (Abu Ghraib, Al-Bayaa, Al-Mahmudiah and field of Veterinary Medicine College -Baghdad University), and the effects of age and sex of the animal in the infection rate by using 200 fecal samples during the period from the beginning of November to the beginning of March, 2017. All fecal samples were examined by conventional microscopic examination {Direct wet smear, lugols iodine smear, Giemsa stain smear and flotation technique (NaCl)}. The total infection rate of microscopic examination was 27.50% and the higher infection rate (34.61%) was recorded in the age group 6 months, while the lower infection rate (23.85%) was recorded in the age group 6-12 months, with a significant (P 0.01) difference. Males had a close infection rate (27.27%) with females (27.72%) with significant difference (P 0.01). A higher infection rate in February (40%), while the low infection rate (80.00%) of *Giardia* in the field Vet. Med. College and the low infection rate (16.66%) in Al-Mahmudiah region with significant differences (P 0.01). A different infection rates of *Giardia* in sheep were recorded by different conventional diagnostic methods. A higher infection rate (9%) was recorded by lugols iodine smear , while a lower infection rate (5%), was recorded by the flotation methods (NaCl) with significant (P 0.01) difference.

KEY WORDS: Giardia, sheep, conventional methods, trophozoites, cyst

#### **INTRODUCTION**

Giardia is a flagellate protozoan parasite causes a disease called Giardiasis (Geurden et al., 2010 and Feng and Xiao, 2011). It is infected numerous hosts that range from different mammals to amphibians and birds (Thompson and Caccio, 2004; Caccio et al., 2005; Monis et al., 2009). Different animal species have been reported worldwide infected by the parasite and considerable economic losses in livestock animals that are associated with the morbidity and mortality (Aloisio et al. 2006; Sweeny et al. 2011). Transmission of Giardia, particularly may occur through either direct contact (farmers, veterinarians, and petting zoos) or through indirect routes such as contaminated water or foods (Dixon, 2009). This parasite have many species such as G. duodenalis (syn. G. lamblia, G. intestinalis ), G. muns, G. microti, G. agillis and G. psicatti (Olson et al., 2004). The organism exist in two forms, vegetative form (trophozoite) capable of causing illness in the host which lives principally in the upper part of small intestine and cyst form that considered as an infective stage of the parasite (Peter and Lisa, 2010). Infected animal revealed symptoms range from asymptomatic to acute or chronic disease (Gardner and Hill, 2001) and the predominant clinical signs in ruminants are diarrhea and reduction in growth rate (Lalle et al., 2005). Diagnosis of the parasite is based upon the demonstration of cysts or motile trophozoites in the feces, duodenal aspirates or serologically (Smith and Mank, 2011).

#### **MATERIALS & METHODS**

Two hundred direct fresh sheep fecal samples were collected from different areas (Al- Bayaa , AL-Mahmudiah , Abu- Ghraib and Field of Veterinary Medicine College, Baghdad University ) in Baghdad city during the period from 1/ November/2016 till 31/ March /2017. The samples were conveys to the parasitic laboratory at Veterinary Medicine College/ Baghdad University. Each fecal sample was examined by direct wet smear (Griffiths, 1978), Lugol's iodine stains (Levine, 1961), Giemsa stain (Coles, 1986) and by concentration method -NaCl flotation (Soulsby, 1982).The information about the age and sex of the animals were recorded.

#### RESULTS

#### Total infection rate of Giardia sp.

The total infection rate of sheep *Giardiasis* by different conventional methods

(Direct fecal wet smear, logul s iodine stain, Giemsa stain and flotation –Nacl) was 27.50% (55/200). (Table, 1)

TABLE 1: The total Infection rate of Giardia in sheep by different conventional methods

Number of samples examined	Positive	Percentage (%)
200	55	27.50

Infection rate of *Giardia* sp. according to age of the animal

The study revealed that all age groups of sheep were infected by *Giardia* sp. with different infection rates, but

with a significant (P 0.01) difference among groups .The higher infection rate was recorded in animals aged more 6 month 34.61 while the lowest infection rate in 6-12 month 23.85% (Table, 2).

TABLE 2:	: Infection rate of	Giardia sp.	in sheet	o according to	the age of th	e animals

	<b>1 1</b>	U	0
Age(Months)	Number of samples examined	Positive*	Percentage (%)
6	26	9	34.61
6-12	109	26	23.85
12-24	65	20	30.76
Total	200	55	27.50
*P 0.01			

#### Infection rate of *Giardia* sp. in sheep according to sex of the animals

Table (3) was showed with significant (P 0.01) difference between both sexes with an infection rate 27.72% in females, and 27.27 % in males.

TABLE 3	: Infection	rate of	Giardia	sp.in	sheep	according	to the	sex of	f the ar	nimals

Sex	Number of samples examined	Positive*	Percentage (%)
Males	99	27	27.27
Females	101	28	27.72
Total	200	55	27.50
*P 0	.01		

Infection rate of *Giardia* sp. in sheep according to Months of the study

The infection rate along the months of the study was variable with significant (P 0.01) difference. The higher

*Giardia* sp. infection rate (40 %) was estimated in February, while the lower infection rate (20%) was recorded in December (Table, 4).

<b>TABLE 4:</b> Infection rates of	Giardia sp.in sheep	p according to the month	hs of the study
------------------------------------	---------------------	--------------------------	-----------------

Months	Number of Samples examined	Positive*	Percentage (%)
November	35	8	22.85
December	35	7	20
January	45	10	22.22
February	45	18	40
March	40	12	30
Total	200	55	27.50
P 0.01			

## Infection rate of *Giardia* sp. in sheep according to the areas of the study

A different infection rate of *Giardia* sp.in sheep was recorded in different areas of the study but with significant

(P 0.01) difference. A higher infection rate (80%) was found in the field of Veterinary Medicine College and the lowest infection rate (16.66 %) was recorded in Al-Mahmudiah area (Table, 5).

<b>TABLE 5:</b> Infection rates of <i>Giardia sp.</i> in sheep according to	the areas of the study
---	------------------------

Areas	Number of samples examined	Positive*	Percentage (%)
Abu Ghraib	33	13	39.39
Al-Biae	85	22	25.88
Al-Mahmudiah	72	12	16.66
Field of Vet. Med. College	10	8	80.00
Total	200	55	27.50
*P 0.01			

# Infection rate of *Giardia* sp.in sheep according to the methods of diagnosis

There was a different infection rate of *Giardia* sp. in sheep recorded by different conventional diagnostic methods with significant (P 0.01) difference. A higher infection rate (9%) was recorded by the Lugol s iodine smear, while the lower infection rate (5%) was found by the flotation - Na Cl) method (Table, 6).

<b>TABLE 6:</b> Infection rate of Giardia sp. i	in sheep by using the different	conventional diagnostic methods
---	---------------------------------	---------------------------------

Methods	No. of Samples *Examined	Positive	Percentage (%)
Direct wet smear		15	7.5
Lugol s iodine smear	200	18	9
Giemsa stain smear		12	6
Flotation -Na Cl		10	5

\*P 0.01

#### DISCUSSION

Giardia is the most commonly intestinal protozoal diagnosed parasite in the world and a numerous methodologies are available to identify the parasite (Olson, 2002), such as the traditional methods for fecal analysis (direct fecal wet smear or fecal concentration techniques by sedimentation or flotation techniques (Cheesbrough, 2005) or by the serological tests (El-Nahas et al., 2013) that were used in the present study. The total infection rate of Giardia sp. in the present was recorded 27.50% from 200 sheep fecal samples that examined by using conventional methods (Direct wet fecal smear, lugols, Giemsa stain and flotation) that was agree or different from reports before in many countries of the world, and the species G. duodenalis has a worldwide distribution (Wade et al., 2000; Degerli and Ozcelik, 2003; Bomfim et al., 2005; Thompson et al., 2008; Winkworth et al., 2008; Muhid et al., 2011; Geurden et al., 2012; Di-Cristanziano et al., 2013 and Wang et al., 2014). Also, it has been reported in sheep in many countries including Australia, Canada, England and USA (Buret et al., 1990; Taylor et al., 1993; Olson et al., 1997; Ryan et al., 2005 and Santín et al., 2007), and in China (Zhang et al., 2012). On the other hand, the result was high than the infection rate (6.26%) of Giardia found in sheep in Southern Spain (Diaz et al., 1996) and AI-Fetly et al. (2010) was estimate an infection rate 13.5% in AL-Diwaniya Province and was less than that identified in sheep from different Canadian farms 38% (Olson et al., 1997). Also a wide range of an infection rates were estimated (6-82%) in North America (Xiao, 1994), while cumulative incidence of Giardia in sheep has been reported in the previous literatures to be nearly 100%. Also, a different Giardia sp. infection rate was estimated in different animal species in different countries of the world; in goats a different incidence of the Giardia infection was recorded that may reach 100% (Castro-Hermida et al., 2005, 2006) and among dairy goats in Brazil (14.3%) by microscopic examination and 42.2% in Spain and 20% among asymptomatic adult goats in France by immunofluorescence technique (Bomfim et al., 2005; Castro-Hermida et al., 2005 and Ruiz et al., 2008), and it has been established in beef and dairy cattle with prevalence reach to 100% (Xiao and Herd, 1994; O'Handley et al., 1999; O'Handley, 2002 and Ralston et al., 2003). The prevalence of infection in cats and dogs was different from <1 to 45% in sheltered dogs (Upjohn et al., 2010) and 4 to 11% in cats (Olson et al., 2010). The difference in the infection rates between the results of present study and other studies may be due to many factors that may be associated with risk of infection that can be narrowed down to the demographic and management factors (Xiao, 1994 and O'Handley et al.,

1999). Demographic factors may include age distribution of animals sampled, size of the farm, geographic location, herd size, and other species of animals present on the farm (Gow and Waldner, 2006). Management factors include general management (type of flooring, calf housing, and frequency and method of cleaning) (Maddox-Hyttel et al., 2006). Also separation of the dam from the calf and administration of colostrum, and don't direct contact with infected animals. Generally, intensive management has been found to favor transmission of Giardia cysts (Hamnes et al., 2006). Previous studies revealed that animals reared indoors especially under group housing were more likely to be infected with the parasite than those housed outside (Quigley et al., 1994 and Reust et al., 1998). Some management practices that reduce direct contact between animals such as separation of new born from the dam immediately after birth may aid in reducing the transmission of the cysts (Wade et al., 2000), because adult animals are a potential source of the parasite especially for neonates as a per parturient rise in cyst excretion has been reported in cattle, sheep, goats and pigs (Xiao and Herd, 1994; Xiao et al., 1994; Wade et al., 2000 and Castro-Hermida et al., 2005). Different studies showed that synanthropic flies are the most important mode of transmission of Giardia duodenal which mechanical transmission of pathogens by flies is intensive and it is achieved through defecation regurgitation or mechanical dislodgment .They carry the viable parasite from unhygienic sites (Graczyk et al., 2001 and Graczyk et al., 2003) and nonbiting flies can deposition of this pathogen on the visited surfaces (Bean et al., 1996 and Wallace et al., 2000). The role of animal infections remains controversial particularly that of livestock and wildlife because of their potential role as zoonotic reservoirs of infection (Cifuentes et al., 2002). The great potential for zoonotic transmission of Giardia is with genotype A and domestic animals, wildlife, and possibly pets act as reservoirs (Guy et al., 2004), and a several studies referred to a variety of birds suggest they may be zoonotic reservoirs (Franssen et al., 2000). Also, the transmission of parasite in humans and animals are restricted largely to the presence of genetic recombination (Ashford and Snowden, 2001, Coope et al., 2007; Teodorovic et al., 2007; Lasek-Nesselquist et al., 2009 and Sprong et al., 2009). There is an effects of age in infection rates of Giardia and cysts of the parasite were found in all age categories, but the high infection rate in this study was recorded in the age group between 1-6 months 9 of 26 (34.61%) and there were an apparent declining infection rate with an increasing age of the animals that was agreement with the previous studies (epidemiological studies) which referred that young

animals were more susceptible to opportunistic parasites than adults in Giardiasis of sheep and goats and the majority focused on the occurrence in lambs and kids. In a longitudinal study of lambs, overall prevalence of G. duodenalis was 23.0% in the first samples and 31.0% in the second samples (Robertson et al., 2010) also, in Belgium, the prevalence was 25.5% in lambs and 35.8% in kids (Geurden et al., 2008), but these results were differ and disagreement with Taylor et al. (1993) who found 68.6 % of lambs excreted Giardia cysts. On the same hands; a higher prevalence of Giardia infections in neonatal lambs than in adult sheep has been reported in Spain (Castro-Hermida et al., 2011). Another study in Australia also revealed that infection with the parasite was higher in lambs aged below twelve months than in adult sheep (Rvan *et al.*, 2005). In the Brazilian study, infections were more in kids from one to three months of age than they were in adult goats (Bomfim et al., 2005).Also, calves aged over nine days were found to be more likely to be infected with Giardia (Gow and Waldner, 2006). On the same way, a study in North America revealed that dairy calves as young as two days of age were harboring the parasite (Mark-Carew et al., 2010) and the burden of infection in dairy cattle has been reported to be low above six months of age (Buret et al., 1990 and Becher et al., 2004). High infection rates in the younger animals may be due to many reasons; It has been suggested through experimental studies that lambs do not rapidly develop high antibody titers against Giardia (Yanke et al., 1998), that develop a specific immunity by the host against the parasite (O'Handley et al., 2003).Wolf (1992) emphasized that the antibodies type IgM and IgA play a major role in the extermination of parasites, and noted that the chronic Giardiasis link with low immune globulin IgG, and acquired immunity after initial infection may emerge as an important protection toward the parasite (Hanevik et al., 2011). As a result, young animals can be considered to be a source of infection for susceptible hosts and high levels of infection with the parasite have been recorded on farms located in areas with poorly drained soils (Tiranti et al., 2011). Poorly drained soils may increase the retention of moisture, which in turn prolongs survival of the cysts in the environment (Barwick et al., 2003). Sex of the animals doesn't affect the infection rate in sheep in the present study that results agreement with Diaz et al. (1996) and Xiao (1994). The infection rates of *Giardia* were variable with significant difference in the different months of the year. A high infection rate was 40 % in February, while a low infection rate (20%) was found in December that agreement with Al-Dulaimi (2016) who refers variation in the infection rates during different months of the year. Furthermore, different infection rates of Giardia were recorded in different areas of the study with significant difference. The infection is diversely dispersed throughout all over Iran, such as East Azerbaijan Province and the incidence in this Province were variable from 15.2% in Tabriz city to 43.8% in Naghadeh District (Saebi, 2005). Although, there is direct evidence of transmission of G. duodenal from small ruminants to the other sheep via contaminated water and it is considered a threat. Also, the prevalence of Giardia in water was significantly higher in the inland area, with higher concentration of livestock and

fewer water treatment plants (Castro-Hermida *et al.*, 2011).On the same hand , the pastures surrounding the drinking water basins are all grazed by small ruminants lead to a substantial public health threat (Tzanidakis *et al.*, 2014).

There were a different infection rates of Giardia in sheep recorded by different conventional diagnostic methods .A high infection rate (9%) was recorded by Lugols iodine smear and a low infection rate (5%) was estimated by flotation methods that was agreement with Zhang et al. (2012) who referred that a different results of each sheep fecal specimen was directly used to smear three slides for iodine wet mount staining and wet smears were examined for the presence of G. duodenalis cysts by light field microscopy (40x magnification). The average prevalence of G. duodenalis infection was 5.0% (34/678) by microscopy after Lugol's iodine staining 5.6% (30/539). Furthermore, the prevalence of G. duodenalis in goats varies (<10% to >40%), depend on the age of animal, geographical locations and diagnostic techniques used in the examination (Robertson, 2009).

#### REFERENCES

Geurden, T., Vercruysse, J. & Claerebout, E. (2010) Is Giardia a significant pathogen in production animals?. *Experimental parasitology*, 124(1), 98-106.

Feng, Y. and Xiao, L. (2011) Zoonotic potential and molecular epidemiology of Giardia species and giardiasis. Clinical microbiology reviews, 24(1), 110-140

Cacciò, S.M., Thompson, R.A., McLauchlin, J. and Smith, H.V. (2005) Unravelling cryptosporidium and giardia epidemiology. Trends in parasitology, 21(9), 430-437.

Monis, P.T., Caccio, S.M. and Thompson, R.C. (2009) Variation in *Giardia*: towards a taxonomic revision of the genus. Trends Parasitol., 25: 93-100

Aloisio, F., Filippini, G., Antenucci, P., Lepri, E., Pezzotti, G., Caccio, S.M. and Pozio, E. (2006) Severe weight loss in lambsinfected with Giardia duodenalis assemblage B. Veterinary Parasitology, 142, 154–158.

Sweeny, J.P.A., Ryan, U.M., Robertson, I.D. and Jacobson, C. (2011) Cryptosporidium and Giardia associated with reduced lamb carcase productivity. Veterinary Parasitology 182, 127–139

Dixon, B.R. (2009) The role of livestock in the food borne transmission of *Giardia duodenalis* and *Cryptosporidium* spp. to humans In: Ortega–Pierres MG, Caccio` SM, Fayer R, Smith H (eds.), *Giardia* and *Cryptosporidium*: from molecules to disease. CAB International, Wallingford, UK. 107 – 122.

Olson, M.R., O'Handley, R.B. and Thompson, R.C.A. (2004) Emerging issues of *Cryptosporidium* and *Giardia* infections in cattle. Trends. Parasitol; 20: 185–191.

Peter, M.R. and Lisa, A.C. (2010) Human-Animal Medicine.Zoonoses.9: 167-171.

Gardner, T.B. and Hill, D.R. (2001)Treatment of giardiasis .Clinical Microbiology Reviews, 14(1), 114-128.

Lalle, M., Pozio, G., Capelli, F., Bruschi, D. and Crotti, S. (2005) Genetic heterogeneity at the \_giardin locus among human and animal isolates of Giardia duodenalisand identification of potentially, zoonotic subgenotypes. Int J Parasitol., 35:207–213.

Smith, H.V. and Mank, T.G. (2011) Diagnosis of human giardiasis. In: Svärd S, Luján HD (eds) *Giardia* :a model organism. Springer, New York. ISBN 978-3-7091-0197-1.

Griffiths, H.J. (1978) A handbook of veterinary parasitology: domestic animals of North America. U of Minnesota Press 212-213.

Levine, N.D. (1961) Protozoan Parasites of Domestic Animals and of Man. Burgess Publishing Company. Minnesota, USA. PP: 118-122.

Coles, E.H. (1986) Veterinary Clinical Pathology .4<sup>th</sup> ed., Press of W.B. Saunders Co:472-545.

Soulsby, E.J.L. (1982) Helminths, Arthropods and protozoa of Domesticated Animals, 7<sup>th</sup> Edition, Lea and Febiger, Philadelphia.

Olson, M.E. (2002) *Giardia* and giardiasis: a zoonotic threat. Compend. Contin. Educ. Pract. Vet; 5(24): 10-14.

Cheesbrough, M. (2005) District laboratory practice in tropical countries (2<sup>nd</sup>ed). Cambridge; New York: Cambridge University Press.

El-Nahas, H.A., Salem, D.A., El-Henawy, A.A., El-Nimr, H.I., Abdel-Ghaffar, H.A. and El-Meadawy, A.M. (2013) *Giardia* diagnostic methods in human fecal samples: A comparative study. Cytometry Part B: Clinical Cytometry, 84B (1), 44-49.

Wade, S., Mohammed, H. & Schaaf, S. (2000) Epidemiologic study of *Giardia* sp. infection in dairy cattle in southeastern New York State. Veterinary Parasitology, 89, 11–21

Bomfim, T.C., Huber, F., Gomes, R.S. and Alves, L.L. (2005) "Natural infection by *Giardia* sp. and *Cryptosporidium* sp. in dairy goats, associated with possible risk factors of the studied properties." Veterinary Parasitology, 134, 9–13.

Thompson, J., Young, R., Power, M., Hufschmid, J., Beveridge, I., Reid, S., Ng, J., Armson, A, and Ryan, U. (2008) "Identification of zoonotic *Giardia* genotypes in marsupials in Australia." Experimental Parasitology, 120, 88-93.

Winkworth, C., Learmonth, J., Matthaei, C. & Townsend, C. (2008) "Molecular characterization of *Giardia* isolates from calves and humans in a region in which dairy

farming has recently intensified. Applied and Environmental Microbiology, 74, 5100-5105.

Muhid, A., Robertson, I., Ng, J., Yang, R, and Ryan, U. (2011) Prevalence of *Giardia spp*. infection in pre-weaned and weaned calves in relation to management factors." The Veterinary Journal, 191, 135-137.

Geurden, T., Vanderstichel, R., Pohle, H., Ehsan, A., von Samson-Himmelstjerna, G., Morgan, E.R. and Claerebout, E. (2012) A multicentre prevalence study in Europe on Giardia duodenalis in calves, with molecular identification and risk factor analysis. Veterinary parasitology, 190(3), 383-390.

Di Cristanziano, V., Santoro, M., Parasi, F., Albonico, M., Shaali, M., Di Cave, D, and Berrilli, F. (2013) Genetic characterization of *Giardia duodenalis* by sequence analysis in humans and animals in Pemba Island, Tanzania." Parasitology International, 63, 438-441.

Wang, H., Zhao, G., Chen, G., Jian, F., Zhang, S., Feng, C., Wang, R., Zhu, J., Dong, H., Hua, J., Wang, M, and Zhang, L. (2014) "Multilocus genotyping of *Giardia duodenalis* in dairy cattle in Henan, China." PloS one, 9, e100453.doi: 10.1371/ journal. pone. 0100453.

Buret, A., Gall, D.G. and Olson, M.E. (1990) Effects of murine giardiasis on growth, intestinal morphology, and disaccharides activity. The Journal of parasitology, 403-409.

Taylor, M.A., Catchpole, J., Marshall, R.N, and Green, J. (1993) 'Giardiasis in lambs at pasture." Veterinary Record, 133, 131–133.

Olson, M.E., Thorlakson, C.L., Deselliers, L., Morck, D.W. & McAllister, T.A. (1997) Giardia and Cryptosporidium in Canadian farm animals. *Veterinary parasitology*, 68(4), 375-381.

Ryan, U.M., Bath, C., Robertson, I., Read, C., Elliot, A., McInnes, L., Traub, R., Besier, B. (2005) Sheep may not be an important zoonotic reservoir for Cryptosporidium and Giardia parasites. Appl Environ Microbiol 71, 4992-4997.

Santin, M., Trout, J.M., Fayer, R. (2007) Prevalence and molecular characterization of Cryptosporidium and Giardia species and genotypes in sheep in Maryland. Vet Parasitol 146, 17-24.

Zhang, D., Iyer, L. M., He, F., and Aravind, L. (2012) Genetic characterizations of *Giardia duodenalis* in sheep and goats in Heilongjiang Province, China and possibility of zoonotic transmission. PLoS neglected tropical diseases, 6(9), e1826.

Díaz, V., Campos, M., Lozano, J., Manas, I., and Gonzalez, J. (1996) Aspects of animal giardiosis in Granada province (southern Spain). *Veterinary parasitology*, *64*(3), 171-176.

AI-Fetly, D.R., AIrodhan, M.A. and Abid, T.A. (2010) Epidemiological and therapeutical study of Giardiasis in sheep in AL-Diwaniya province. College of Veterinary Medicine, AL-Qadisiya Universit Kufa Journal for Veterinary Medical Sciences, 1(1).

Castro-Hermida, J.A., Pors, poupin, B., Ares-Mazas, E. and Chartier, C. (2005) Prevalence of *Giardia duodenalis and Cryptosporidium parvum* in goat kids in western France. Small Ruminant Research, 56(1), 259-264.

Castro-Hermida, J.A., Carro-Corral, C., Gonzalez-Warleta, M. and Mezo, M. (2006) Prevalence and intensity of infection of *Cryptosporidium spp* and *Giardia duodenalis* in dairy cattle in Giardia (NW Spain).Zoonoses and Public Health, 53(5),244-246.

Ruiz, A., Foronda, P., Gonzalez, J.F., Guedes, A., Abreu-Acosta, N., Molina, J.M. and Valladares, B. (2008) Occurrence and genotype characterization of *Giardia duodenalis* in goat kids from the Canary Islands, Spain. Veterinary.

Xiao, L. & Herd, R.P. (1994) Infection patterns of *Cryptosporidium* and *Giardia* in calves. Veterinary Parasitology, 55(3), 257-262.

O'Handley, R.M. (2002) *Giardia* in farm animals. In: *Giardia: The Cosmopolitan Parasite*, eds Olson, B. E., M. E. Olson and P. M. Wallis, CAB International, Wallingford, UK, pp. 97–105.

O'Handley, R., Cockwill, C., McAllister, T.A., Buret, A.G., Jelinski, M. and Olson, M.E. (1999) Duration of naturally acquired giardiasis and cryptosporidiosis in dairy calves and their association with diarrhoea. *Journal of the American Veterinary Medical Association*, 214, 391–396.

Ralston, B.J., McAllister, T.A. and Olson M.E. (2003) Prevalence of *Giardia and Cryptosporidium* and ersoni and their effects on performance in feedlot beef cattle. Canadian journal of animal science, 83(1), 153-159.

Upjohn, M., Cobb, C., Monger, J., Geurden, T., Claerebout, E., and Fox, M. (2010) Prevalence, molecular typing and risk factor analysis for *Giardia duodenalis* infections in dogs in a central London rescue shelter. Veterinary parasitology, *172*(3), 341-346.

Xiao, L. & Herd, R.P. (1994) Infection patterns of *Cryptosporidium* and *Giardia* in calves. Veterinary Parasitology, 55(3), 257-262.

Gow, S. and Waldner, C. (2006) An examination of the prevalence of and risk factors for shedding of *Cryptosporidium spp. and Giardia spp.* in cows and calves from western Canadian cow–calf herds. Veterinary parasitology, 137(1), 50-61

Maddox-Hyttel, C., Langkjær, R.B., Enemark, H.L and Vigre, H. (2006) "Cryptosporidium and Giardia in different age groups of 93 Danish cattle and pigs

Occurrence and management associated risk factors." Veterinary Parasitology, 141, 48–59

Hamnes, I., Gjerde, B, and Robertson, L. (2006) "Prevalence of *Giardia* and *Cryptosporidium* in dairy calves in three areas of Norway." Veterinary Parasitology, 140, 204–216.

Quigley, J., Martin, K., Potgieter, D., Reinemeyer, C., Rohrbach, B., Dowlen, H, and Lamar, K. (1994) "Effects of housing and colostrum feeding on the prevalence of selected infectious organisms in faeces of Jersey calves." Journal of Dairy Science, 77, 3124-3131.

Ruest, N., Faubert, G, and Couture, Y. (1998) "Prevalence and geographic distribution of *Giardia spp.* and *Cryptosporidium spp.* in dairy farms in Quebec." Canadian Veterinary Journal, 39, 697-700

Wade, S., Mohammed, H. and Schaaf, S. (2000) "Epidemiologic study of *Giardia* sp. infection in dairy cattle in southeastern New York State." Veterinary Parasitology, 89, 11–21.

Graczyk, T., Conn, D., Marcogliese, D., Graczyk, H., and De Lafontaine, Y. (2003) Accumulation of human waterborne parasites by zebra mussels (*Dreissena polymorpha*) and Asian freshwater clams (*Corbicula fluminea*). Parasitology research, 89(2), 107-112.

Graczyk, T.K., Knight, R., Gilman, R.H. and Cranfield, M.R. (2001) The role of non-biting flies in the epidemiology of human infectious disease. Microbes infect; 3:231-235.

Cifuentes, E., Gomez, M., Blumenthal, U., TellezRojo, M.M., Romieu, Clark, C.G. and Diamond, L.S. (2002) Methods for cultivation of luminal parasitic protists of clinical importance, Clin. Microbiol Rev., 15(3):329-341.

Franssen, F.F.J., Hooimeijer, J., Blankenstein, B. and Houwers, D.J. (2000) Giardiasis in a white stork in the Netherlands. J. Wildlife Dis; 36(4): 764–766.

Ashford, R.W. and Snowden, K.S. (2001) Dogs and protozoan zoonoses. In Dogs, Zoonoses and Public Health. 3<sup>rd</sup> ed., Macpherson, C. N. L.; Meslin, F. X. and Wandeler, A. I. CABI Publishing, New York: 127–128.

Coope, M.A., Adam, R.D., Worobey, M. and Sterling, C. R. (2007) Population genetics provides evidence for recombination in *Giardia*. Current Biology, 17(22), 1984-1988.

Coope, M. A., Adam, R.D., Worobey, M. and Sterling, C. R. (2007) Population genetics provides evidence for recombination in *Giardia*. Current Biology, 17(22), 1984-1988.

Lasek-Nesselquist, E., Welch, D.M., Thompson, R.C., Steuart, R.F. and Sogin, M.L. (2009) Genetic exchange

within and between assemblages of *Giardia duodenalis*. Journal of Eukaryotic Microbiology, 56(6), 504-518.

Sprong, H., Cacciò, S.M. and van der Giessen, J.W. (2009) Identification of zoonotic genotypes of *Giardia duodenalis*. *PLoS* neglected tropical diseases, 3(12), 558.

Robertson, L.J., Hanevik, K., Escobedo, A.A., Morch, K. and Langeland, N. (2010) Giardiasis--why do the symptoms sometimes never stop? Trends in Parasitology, 26(2), 75-82.

Geurden, T., Thomas, P., Casaert, S., Vercruysse, J., and Claerebout, E. (2008) Prevalence and molecular characterization of Cryptosporidium and Giardia in lambs and goat kids in Belgium. Veterinary parasitology, 155(1), 142-145.

Castro- Hermida, J.A., Garcia- Presedo, L., Gonzalez-Warleta, M. and Mezo, M. (2011) Prevalence of *Cryptosporidium and Giardia* in roe deer (*Capreolus capreolus*) and wild boars (*Sus scrofa*) in Galicia (NW, Spain).Veterinary parasitology,179(1),216-219.

Mark-Carew, M.P., Khan, Y., Wade, S.E., Schaaf, S. and Mohammed, H.O. (2010) Incidence of and risks associated with Giardia infections in herds on dairy farms in the New York City Watershed. Acta Veterinaria Scandinavica, 52 (1), 44.

Becher, K., Robertson, I., Fraser, D., Palmer, D, and Thompson, R. (2004 "Molecular epidemiology of *Giardia* and *Cryptosporidium* infections in dairy calves originating from three sources in Western Australia." Veterinary Parasitology, 123, 1-9.

Yanke, S.J., Ceri, H., McAllister, T.A., Morck, D.W. and Olson, M.E. (1998) Serum immune response to *Giardia duodenalis* in experimentally infected lambs. Veterinary parasitology, 75(1), 9-19.

Wolfe, M.S. (1992) Giardiasis. Clinical . Microbiology . Rev; 5(1): 93–100.

Hanevik, K., Kristoffersen, E., Svard, S., Bruserud, O., Ringqvist, E., Sornes, S. and Langeland, N. (2011) Human cellular immune response against *Giardia lamblia* 5 years after acute giardiasis. Journal of Infectious Diseases, 204(11), 1779-1786.

Tiranti, K., Larriestra, A., Vissio, C., Picco, N., Alustiza, F., Degioanni, A., and Vivas, A. (2011) Prevalence of *Cryptosporidium spp.* and *Giardia spp.*, spatial clustering

and patterns of shedding in dairy calves from Córdoba, Argentina. Revista Brasileira de Parasitologia Veterinária, 20(2), 140-147.

Barwick, R.S., Mohammed, H.O., White, M.E. & Bryant, R.B. (2003) Prevalence of *Giardia spp. and Cryptosporidium spp.* on dairy farms in southeastern New York state. Preventive veterinary medicine, 59(1), 1-11.

Díaz, V., Campos, M., Lozano, J., Manas, I., and Gonzalez, J. (1996) Aspects of animal giardiosis in Granada province (southern Spain). Veterinary parasitology, 64(3), 171-176

Al-Dulaimi, A.A., Al-Bayati, N.Y., Nazal, M.F. and Mahmood, S.M. (2016) Correlation Between Nutritional Status and *Giardia lamblia* Infection of Primary Schoolchildren in Al-khalis City.

Robertson, L.J. (2009) Giardia and Cryptosporidium infections in sheep and goats: a review of the potential for transmission to humans via environmental contamination. Epidemiology and Infection, 137(7), 913-921.

De erli, S., and Özçelik, S. (2003) The First Giardia Infection in Cattle Gall Bladder. Turkish Journal of Veterinary and Animal Sciences, 27(5), 1231-1233.

Xiao, L., and Herd, R.P. (1994) Infection patterns of Cryptosporidium and Giardia in calves. *Veterinary Parasitology*, 55(3), 257-262.

Olson, M.E., Leonard, N.J. and Strout, J. (2010) Prevalence and diagnosis of Giardia infection in dogs and cats using a fecal antigen test and fecal smear. The Canadian Veterinary Journal, 51(6), 640.

O'Handley, R.M., Ceri, H., Anette, C. and Olson, M.E. (2003) Passive immunity and serological immune response in dairy calves associated with natural Giardia duodenalis infections. Veterinary parasitology, 113(2), 89-98.

Tzanidakis, N., Sotiraki, S., Claerebout, E., Ehsan, A., Voutzourakis, N., Kostopoulou, D. and Geurden, T. (2014) Occurrence and molecular characterization of *Giardia duodenalis* and Cryptosporidium spp. in sheep and goats reared under dairy husbandry systems in Greece. Parasite, 21.

Thompson, R.A. Caccio, S.M. (2004) The zoonotic significance and molecular epidemiology of Giardia and giardiasis. Veterinary parasitology, *126*(1), 15-35.