

# INTERNATIONAL JOURNAL OF ADVANCED BIOLOGICAL RESEARCH

© 2004-2014 Society For Science and Nature (SFSN). All Rights Reserved.

www.scienceandnature.org

# SNUFFLES DISEASE IN RABBITS: 4- PATHOLOGICAL STUDY

Khalil H. Al-Jeboori & Saif S. Rasheed

Dept. of Pathology, College of Veterinary Medicine, University of Baghdad, Iraq.

## ABSTRACT

Snuffles disease is a major problem in rabbits, for this reason this study aimed to identify all the pathological findings associated with this disease process experimentally. Pasteurella multocida is the causative agent their LD50 was  $10^7$ CFU/ml, infective dose was 10<sup>2</sup> CFU/ml. Then two groups of rabbits. 1<sup>st</sup> group was control (7), the 2<sup>nd</sup> group was infected group (27) were injected with 10<sup>2</sup> CFU/ml I/P whereas control group injected with pbs. All the pathological lesions were recorded at 4<sup>th</sup>, 8<sup>th</sup>, 12<sup>th</sup>, 16<sup>th</sup> and 20<sup>th</sup> days post infection. During these periods different pathological lesions grossly seen such as congestion, hemorrhage edema, all over organs especially liver, lungs, kidneys, spleen, lymph nodes and heart enlargement. Microscopically, multiple abscesses in liver and chollangitis and extensive fibrinous and suppurative bronchopneumonia and fibrino purulent pleuritis. Kidneys showed extensive embolic nephritis together with bacterial colonies distributed in the suppuration. Mesenteric and prefemoral lymph nodes and spleen showed reactive lymphoid hyperplasia and micro abscess in white pulp of spleen and in lymphoid follicles of the lymph nodes giving suppurative and edematous lymphadenitis feature. P. multocida induces extensive fatal pathological lesions in the rabbits organs.

**KEYWORDS:** Snuffles in rabbits – pathological findings.

# **INTRODUCTION**

Snuffles disease a major problem in rabbits caused by various Pasteurella multocida serotypes, the disease occurs during the stress factor such as shipping, mating, experimental handling and malnutrition<sup>[1,2]</sup>. P. multocida already present in upper respiratory tract and tympanic bullae of rabbits and during seasonal influences and other stresses factor the P. multocida multiply rapidly and induce the disease in the lungs and upper respiratory tract such as pneumonia, otitis media and rhinitis together with septicemia<sup>[3]</sup>. For the importance of this disease problem, this study aimed to identify pathological parameters associated with this experiment disease process in rabbits.

# MATERIALS AND METHODS

A local strain of *Pasteurellamultocida* were reidentified <sup>[4]</sup> and the LD50 and infective lethal dose were indentified <sup>[5]</sup>, the LD50 was  $10^7$  CFU/ml and infective dose was  $10^2$ 



FIGURE :Lungs grossly showed 1 consolidation and emphysema (12th -16th ) days of foci with rounded borders of liver parenchyma. infection.

CFU/ml. Two groups of rabbits were taken, first group 7 (control group) injected I/P with pbs, the second group 27 (infective group) injected I/P with  $10^2$  CFU/ml P. multocida and 4 rabbits sacrificed at 4 days intervals (at 4<sup>th</sup>, 8<sup>th</sup>, 12<sup>th</sup>, 16<sup>th</sup> and 20<sup>th</sup>) days post infection. The other rabbits<sup>[7]</sup> were died during the experiment period. All the dead and sacrificed rabbits organs were examined and pieces for histopathological study were taken, fixed routinely and processed and examined under light microscope<sup>[6]</sup>.

#### **RESULTS & DISCUSSION Gross Pathology**

The Lungs: Showed extensive congestion, hemorrhage, fibrin and edema in pulmonary tissue at the  $4^{th} - 12^{th}$  days post infection, the consolidation and emphysema appeared at  $12^{th} - 20^{th}$  days post infection (Fig – 1) together with pleural adhesions.



sever gray **FIGURE 2:** Liver grossly showed multifocal pale yellowish

The Liver: Showed extensive congestion, edema and hemorrhage at the  $4^{th} - 12^{th}$  days post infection, multiple pale yellow foci appeared on liver tissue at  $12^{th} - 20^{th}$  days post infection together with swelling of gall bladder (Fig – 2)

The Kidney: Showed extensive congestion of the cortex and medulla, edema and hemorrhage and the kidney is soft in consistency along the period of experiment(Fig - 3).



**FIGURE 3:** Kidney and mesenteric lymph node grossly showed enlargement and multi edematous areas.



**FIGURE 5** : Heart grossly showed enlargement and adhesion of the lung with thoracic cavity



**FIGURE 7**: Lung tissue microscopically showed streaming macrophages exudate together with fibrin .(H&E) X100

# HISTOPATHOLOGY

The Lungs: It showed congestion of blood vessels, edema, fibrin and emphysema at the beginning and thickening of pulmonary interstitial tissue at 8<sup>th</sup> day post infection, also

The Lymph Nodes: All the mediastinal, mesenteric and prefemorallymph nodes showed edematous enlargement, congestion and hemorrhage along the experiment period (Fig - 3).

The Spleen: Showed sever congestion and hemorrhage together with edema in the whole splenic tissue (Fig - 4). The Heart: Showed extensive hemorrhage and congestion between the muscle fibers and cardiac hypertrophy (Fig - 5).



**FIGURE 4:** Spleen, grossly showed congestion and enlargement at 4<sup>th</sup> day of infection.



**FIGURE 6:** Lung tissue microscopically showed suppurative bronchopneumonia et the  $12^{th}$  - $16^{th}$  days of infection (H&E) X100



**FIGURE 8:** Liver tissue microscopically showed suppurative cholangitis , thrombus and perivascular leukocyte cuffing (H&E)X100

there is fibrinous pneumonia and suppurative bronchopneumonia (Fig - 6) characterized by infiltration of neutrophils in bronchi, bronchioles and alveoli together with fibrin and edema. Steaming macrophages exudates

(Fig - 7) were seen in some alveolar spaces, hyperplasia of peribronchial lymphoid tissue. Also there is fibrino purulent pleuritis later on.

The Liver: The lesions began as congestion, perivascular leukocytes cuffing, thrombus formation and suppurative

**FIGURE 9:** Liver tissue microscopically showed diffuse abscess formation (H&E)X100



**FIGURE 11** : Lymph node microscopically showed suppurative lymphadenitis together with localized microabscess in the lymphoid follicles (H&E)X100

The Kidneys: The lesions began as congestion of renal blood vessels, infiltration of neutrophils between the renal tubules and glomeruli lead to embolic nephritis (Fig - 10). Also bacterial colonies were seen in the neutrophils exudates in the renal tissue. At the advance stage atrophies of glomerular tuft and dilation of Bowman's space.

The Lymph Nodes: The lesions began as a reactive hyperplasia of lymphoid follicles and congestion of sub capsular and medullary sinuses and edema all over the lymph node tissue together with infiltration of neutrophils and macrophages lead to supportive and edematous lymphadenitis later on (Fig - 11).

The Spleen: The lesions began as congestion, reactive hyperplasia of white pulp and at the  $12^{th} - 20^{th}$  days post infection there is abscessation inside white pulp and reticuloendothelial cells hyperplasia in red pulp together with hemorrhage (Fig – 12).

cholangitis (Fig - 8).Also multinecrotic areas in liver tissue lead to liver abscess formation (Fig - 9). Hypertrophy of hepatocytes lead to sinusoidal disappearance later on.



**FIGURE -10 :** Kidney tissue microscopically showed embolic nephritis with abscess formation adjacent glomeruli , also there is bacterial colonies in glomeruli and periglomerular tissue (H&E)X100



**FIGURE 12**: Spleen microscopically showed extensive reactive lymphoid hyperplasia together with localized microabscess in white pulp. (H&E)X100

The Heart: The lesions began as congestion and hemorrhage between muscle fibers together with thrombus. Also clotted blood was seen in heart cavities during the experiment period.

All the pathological lesions were associated with extensive neutrophils infiltration occurred as a result of bacterial effects that lead to leukocytes recruitment mainly neutrophils and some lymphocytes and macrophages and these inflammatory cells induce parenchymal cell injury in the body organ <sup>[7,8,9 10]</sup>. Thrombus is mostly common in all the organs due to effect of endotoxins increase the adherence of platelets on the endothelial cells with the expression of adhesion molecules<sup>[111]</sup>. Neutrophils are the main inflammatory cells infiltrated the all over the organs, these inflammatory cells have powerful effects on production and release of cytokines and have also be shown to exert potent proinflammatory and anti-

inflammatory effects of animal model of inflammatory disease<sup>[12].</sup> Lipopolysaccharide (endotoxins) of *P*. multocida have a role in endothelial damage of blood vessels and increase their permeability and activate the alternative pathway of the complement as well as to increase chemotactic activity of all the inflammatory cells into the organs which more evident in this experiment. The degree of neutrophils activation increased with the degree of tissues injury occurred as a result of effect of bacterial endotoxins<sup>[12]</sup> which lead to endothelial damage so hemorrhage and fibrin together with edema infiltrated in all the organs, other lesion, focal abscessations occurred in all organs later on as a result of tissue injury caused by bacterial endotoxins and bacterial proliferation, since necrosis and neutrophils infiltration lead to abscess formation in the lungs, liver, spleen as well as cholanitis which more observed in this study, similar finding reported by<sup>[13,14]</sup>. Necrosis of the tissue in all examined organs was occurred also as a result of ischemia due to partial obstruction of blood vessels at inflammatory areas in all the organs<sup>[15]</sup>. Emphysema in the lungs tissue occurred as a result of neutrophils infiltration caused damage of alveoli and developed in to emphysema which is observed in all pulmonary tissues in this study similar to <sup>[16]</sup>finding. Streaming macrophages were seen in all pulmonary tissue, this feature considered a pathognomonic findings for pneumonic pasteurellosis<sup>[17]</sup>. Peribronchial lymphoid tissue hyperplasia in lungs, lymphoid hyperplasia of cortical resion in lymph node and in white pulp of the spleen occurred as a result of persistence stimulation of these lymphoid tissues hv lipopolysaccharide and outer membranes protein of P. *multocida*<sup>[18,19]</sup>. Other lesions were observed in this study dilation of Bowman's space, shrinkage of glomeruli, whereas, other glomeruli showed increase cellularity, these lesions were related to immune complex deposition (*P. multocida* endotoxins and antibodies)<sup>[14]</sup> which more evident in this study together with the embolic nephritis which occurred as a result of hematogenous route of dissemination of *P. multocida* so reach into glumeruli and infect periglomerular interstitial tissue so localized abscesses seen between glomeruli and renal tublules. The abscesses and necrosis occurred in all the organs as a result of several mechanisms of tissue injuries such as reactive oxygen species, lytic enzymes, inflammatory cytokines and modulators of hemostasis <sup>[20, 21].</sup>

## REFERENCES

- DiGiacoma, R.F., Garlinghonse, L.E. and Van Hoosier, G.L. (1983) Natural history of infection with *Pasteurella multocida* in rabbits. J.Am. Vet. Med. Assoc. 183:1172-1175.
- [2]. DiGiacoma, R.F., Deeb, B.J., Giddens, Jr.W.E., Bernard, B.L. and Chengappa, M.M. (1989) Atrophic rhinitis in New Zealand white rabbits infected with *Pasteurella multocida*. Am. J. Vet. Res. 50:1460-1465.
- [3]. DiGiacoma, R. F., Xu, Y. M., Allen, V., Hinton, M. H. And Pearson, G.R. (1991) Naturally acquired *Pasteurellamultocida* infection in rabbits. Clinical pathological a spects. Can. J. Vet. Res. 55:34-38.

- [4]. Quinn, P.J., Carter, M.E, Narkey, B.K. and Carter, G.R. (2004) Clinical Veterinary Microbiology, Wolf Publishing Mosby. Year Book Inc. Europe Limited 6<sup>th</sup> edition P. 250.
- [5]. Dixon, W.J. (1980) Efficient analysis of experimental observation. Ann. Res. Pharmacol. Toxicol. 20:441-462.
- [6]. Luna, L.G. (1968) Manual of histological staining methods of the Armed Forces Institute of pathology, 3<sup>rd</sup>. ed. McGraw-Hill Book Company, USA.
- [7]. Uchiba, M., Okajima, K., Murakami, K., Mohri, M., Okabe, H. and Takatsuki, K. (1997) Rhs-TM prevent ET-induced increase in pulmonary vascular permeability through protein C Activation Am. J. Physiol. Lung cell Mol. Physiol. 273:889-894.
- [8]. Lechner, A.J., Velasquez, A., Knudsen, K.R., Johanns, C.A., Thomas, F., Tracy, J. and Matuschak, G.M. (1998) Cholestatic liver injury increases circulating TNF- and IL6 and mortality after *Escherichia coli* endotoxemia. Am. J. Resp. Crit. Care Med 157(5):1550-1558.
- [9]. Crespo, E., Macias, M., Pozo, D. Escames, G., Martin, M., Vives, F., Guerrero, J. M. and Acuna-Castro Viejo, D. (1999) Melatonin inhibits expression of inducible No Synthase II in liver and lung and prevents endotoxemia in lipopo lysaccharide induced multiple organ dysfunction syndrome in rats. The FASEB Jour. 13:1537-1546.
- [10]. Lawson, J.A., Burn, A. R., Farhood, A., Lynn-Bajt, M., Collins, R. G., Smith, C. W., Jaeschke, H. (2000) Pathophysiologic importance of E and L selection for neutrophils – induced liver injury during endotoxemia in mice. Hepatology 32(5)990-998.
- [11]. Pawlinski, R., Pedesen, B. and Mackman, N. (2003) Role of the tissue factor – thrombin pathway in endotoxemia. J. Thrombosis Haemotasis. 1:116-127.
- [12]. Kapoor, V., Katoch, R. C., Sharma, M., and Asrani, R. K. (2004) Pathogenicity test of *P. multocida* in mice A: 1. Indian J. Anim. Sci. 74:495-496.
- [13]. Cunningham, J.G. (2002) Textbook of Veterinary physiology, 3<sup>rd</sup>. ed. Saunders, USA.
- [14]. Plumlee, K.H. (2004) Clinical Veterinary toxicology, Mosby, USA.
- [15]. Reamer, K.A. and Ideker, R. E. (1987) Myocardial ischemia and infection – Human Pathol. 18:462-470.
- [16]. Kinoshita, M., Mochizuki, H. and One, S. (1999) Neutrophils accumulation following human endotoxemia. Chest. 116:1707-1715.

- [17]. Al Jeboori, K. H. (1984) Study of some Aspects of pneumonia in local goats, M. Sc. Thesis submitted to the college of Veterinary Medicine, Univ. of Baghdad, Iraq.
- [18]. Olivera, S. (2006) Essential knowledge: respiratory disease in the growing pig – innate immune response. Veterinary diagnostic laboratory, University of Minnesota.
- [19]. Nor –Satinati, S., Zuki, A. B. Z Zamri Saad, M., Awang – Hazmi, A. J. and Saw Popo (2006) The response of gut associated lymphoid tissue (GALT)

following intranasal administration of *P. multocida* B2 in Rats. Journal of Animal and Veterinary advaces. Medweel Journal 5(11):1029-1034.

- [20]. Klebanoff, S. J. (1992) Oxygen metabolites from phagocytes In: Gallin, J. I. and Goldstein, I. M. (eds) Inflammation: Basic Principles and Clinical Correlates. Ravan Press, New York P:541-588.
- [21]. Jaeschke, H. and Smith, C. W. (1997) Mechanism of neutrophile – induced parenchymal cell injury. J. leuko. Biol. 61:647-653.