



IMPACT OF ULTRAVIOLET-B RADIATION ON NODULATION AND NITROGEN METABOLISM IN *PHASEOLUS VULGARIS* L. C.V. PREVAIL

Vijayalakshmi, R. & Rajendiran, K.

Department of Botany, K.M. Centre for Post Graduate Studies, Pondicherry - 605 008, India.

Corresponding author email: rajeworks@yahoo.com

ABSTRACT

Common bean, *Phaseolus vulgaris* L. c.v. Prevail was considered as a poor nitrogen fixing plant compared to other legumes. Even though there are several attempts to produce better varieties for improved N₂ fixation, no work has been done to evaluate the sensitivity of this crop to elevated UV-B radiation. Depletion of ozone layer threatens to continue so as the green house gases around the globe increases in thickness, and the heat that normally would escape the troposphere and enter the stratosphere no longer does so, leaving the stratosphere cooler. As this layer cools down from the normal temperature, ozone depletion is enhanced, which an indirect effect of global is warming in addition to the direct depletion by the ozone depleting substances (ODS). As a result, the ultraviolet-B (UV-B) radiation is bound to increase, disturbing the ecosystems. In this study an attempt to assess the UV-B effects on nitrogen metabolism in the leaves, roots and nodules of common bean, *Phaseolus vulgaris* L. c.v. Prevail was made. The nodulation and nitrogen metabolism on 30 and 45 DAS (days after seed germination) of common bean after exposure to supplementary UV-B radiation (2 hours daily @ 12.2 kJ m⁻² d⁻¹; ambient = 10 kJ m⁻² d⁻¹), were monitored. UV-B stress decreased the protein and amino acid contents of *Phaseolus vulgaris* L. c.v. Prevail in the leaves by 33 to 39 % and 17 to 23 % respectively and reduced nitrate and nitrite by 23 to 26 % and 36 to 38 % in the leaves and by 36 to 44 % and 46 to 48 % in the root nodules. UV-B exposure suppressed NRA (nitrate reductase activity) by 60 % in leaves and 45 to 51 % in nodules. Nodulation was suppressed by UV-B as the number of root nodules (42 to 46 %) and their fresh mass (50 %) were far below controls. UV-B stress also inhibited nitrogenase enzyme activity by 44 to 47 % in roots and by 51 to 55 % in root nodules. Present work indicates that *Phaseolus vulgaris* L. c.v. Prevail has very little tolerance to UV-B stress as indicated by the severe depression of the *Rhizobium*-legume symbiotic nitrogen fixing system.

KEY WORDS: Global warming, Ultra Violet-B stress, *Phaseolus vulgaris*, root nodules, nitrogen metabolism.

INTRODUCTION

Phaseolus vulgaris L. c.v. Prevail (Common bean) has been considered as a poor nitrogen fixing crop compared to other legumes and only recently there have been attempts to select host plants of this species and for breeding better varieties of this crop for increased nitrogen fixation (Graham and Halliday, 1977, Fernandes *et al.* 1982). Even though there are several attempts to produce better varieties for improved N₂ fixation, no work has been carried to evaluate the sensitivity of this crop to elevated UV-B radiation. Human activity has dumped CO₂ along with other heat-trapping gases into the atmosphere which act as a blanket holding in the heat around the earth. These gases increase in thickness warming the troposphere and cooling the stratosphere thereby indirectly depleting the ozone layer in addition to the direct method by ozone depleting substances (ODS). The depletion in the ozone layer allows more ultraviolet-B (UV-B) radiation into earth's surface, affecting the growth of legumes and inhibiting biological nitrogen fixation. An elevation in the flux of ultraviolet-B radiation (280-320nm) is an important atmospheric stress and is detrimental to plant growth and development (Caldwell *et al.*, 1998, Rajendiran and Ramanujam, 2000, Rajendiran and Ramanujam, 2003 and Rajendiran and Ramanujam, 2004, Kokilavani and Rajendiran, 2013.). At the metabolism level, it severely

inhibits photosynthesis (Caldwell *et al.* 1998, Kulandaivelu and Lingakumar, 2000) and hampers nodulation and nitrogen fixation (Balakumar *et al.* 1993, Rachel and Santhaguru, 1999, Rajendiran and Ramanujam 2006, Sudaroli Sudha and Rajendiran, 2013a, 2013b) in sensitive plants. Although the test plant selected for the present study was known to be a poor N₂ fixing crop, the objective of the present study was to find out the extent of damage caused by supplementary UV-B on nodulation and nitrogen metabolism to provide the basic information required for breeding a common bean variety with improved N₂ fixation as well as UV-B tolerance.

MATERIALS & METHODS

Common bean, *Phaseolus vulgaris* L. c.v. Prevail plants were grown in pot culture in the naturally lit greenhouse (day temperature maximum 38 ± 2 °C, night temperature minimum 18 ± 2 °C, relative humidity 60 ± 5 %, maximum irradiance (PAR) 1400 μmol m⁻² s⁻¹, photoperiod 12 to 14 h). Supplementary UV-B radiation was provided in UV garden by three UV-B lamps (*Philips TL20W/12 Sunlamps*, The Netherlands), which were suspended horizontally and wrapped with cellulose diacetate filters (0.076 mm) to filter UV-C radiation (< 280 nm). UV-B exposure was given for 2 h daily from 10:00 to 11:00 and 15:00 to 16:00 starting from the 5th

day after sowing. Plants received a biologically effective UV-B dose (UV-B_{BE}) of 12.2 kJ m⁻² d⁻¹ equivalents to simulated 20 % ozone depletion at Pondicherry (12°2'N, India). The control plants, grown under natural solar radiation, received UV-B_{BE} 10 kJ m⁻² d⁻¹. The seedlings (10 days old) in each pot were inoculated with 200 mg of the commercial preparation of *Rhizobium* (cowpea strain) inoculum suspended in 1 cm³ of water and poured on the surface of the soil as suggested by Shriner and Johnston (1981). Ten plants from each treatment and control were carefully uprooted from the soil at 30 and 45 DAS (days after seed germination) and the number and fresh mass of both the stem and root nodules were recorded. The nitrate and nitrite contents, nitrogenase and nitrate reductase activity of the leaf, root and root nodules were recorded at 30 and 45 DAS, since nodulation was at its peak level during this period. The biochemical estimations were made from the compound leaves at 30 and 45 DAS. The amino acid content was determined by the method of Moore and Stein (1948). Soluble proteins were estimated using Folin phenol reagent method (Lowry *et al.*, 1951). Nitrate and nitrite contents were determined using

naphthylamine salt-mixture (Woolley *et al.*, 1960). *In vivo* NRA was assayed by the method of Jaworski (1971) with suitable modifications (Muthuchelian *et al.*, 1993). Nodular nitrogenase activity was determined by the acetylene reduction technique (Stewart *et al.*, 1967). The values were analysed by Tukey's multiple range test (TMRT) at 5 % level of significance (Zar, 1984).

RESULT & DISCUSSION

The protein and amino acid contents of UV-B stressed *Phaseolus vulgaris* L. c.v. Prevail, decreased by 33 to 39 % and 17 to 23 % respectively in the leaves (Table 1). According to Tevini *et al.* (1981), Vu *et al.* (1981), Rajendiran and Ramanujam (2006) and Sudaroli Sudha and Rajendiran (2013a, 2013b) reductions in soluble protein and amino acid contents of leaves are features of UV-B stress. Plants grown in controlled condition accumulated more nitrate and nitrite in the root nodules (Table 1). On the other hand UV-B stressed plants showed reduction in nitrate and nitrite by 23 to 26 % and 36 to 38 % in the leaves and by 36 to 44 % and 46 to 48 % in the root nodules respectively (Table 1).

TABLE 1. Changes in number and fresh mass (g) of nodules per root system, contents of proteins [mg g⁻¹(f.m.)], amino acids, nitrates and nitrites [mg g⁻¹(d.m.)], and the activities of nitrate reductase, NRA [$\mu\text{mol}(\text{NO}_2^-) \text{kg}^{-1}(\text{f.m.}) \text{s}^{-1}$] and nitrogenase, N₂-ase [$\mu\text{mol}(\text{ethylene reduced}) \text{g}^{-1}(\text{f.m.}) \text{s}^{-1}$] in the 30 and 45 DAS (days after seed germination) leaves, roots and nodules of *Phaseolus vulgaris* L. c.v. Prevail exposed to supplementary UV-B radiation. Means followed by different letters are significantly different at $P = 0.05$, $n = 10$.

Organ	Parameter	Control		UV - B	
		30DAS	45DAS	30DAS	45DAS
Leaf	Protein	16.83b	20.16b	11.22a	12.24a
	Amino acid	21.75b	23.46b	16.68a	19.28a
	Nitrate	4.96b	6.23b	3.67a	4.79a
	Nitrite	0.22b	0.29b	0.14a	0.18a
	NRA	2.78b	2.89b	1.12a	1.16a
Root Nodule	Nodule Number	26.5b	28.5b	14.3a	16.3a
	Nodule Fresh Mass per root	0.20b	0.24b	0.10a	0.12a
	Nitrate	3.32b	3.79b	2.11a	2.13a
	Nitrite	0.28b	0.31b	0.15a	0.16a
	NRA	2.32b	2.64b	1.26a	1.29a
Root	N ₂ -ase	23.10b	25.23b	11.27a	11.38a
	N ₂ -ase	0.36b	0.38b	0.19a	0.21a

Ghisi *et al.* (2002) in barley, Rajendiran and Ramanujam (2006) in *Vigna radiata* (L.) Wilczek, Sudaroli Sudha and Rajendiran (2013a) in *Sesbania grandiflora* (L.) Pers. and Sudaroli Sudha and Rajendiran (2013b) in *Vigna unguiculata* (L.) Walp. c.v. BCP-25 have reported significant reductions in nitrate reductase and glutamine synthetase activities both in the UV-B receiving leaves as well as in the root system. However Chimphango *et al.* (2003) found no adverse effect of elevated UV-B radiation on growth and symbiotic function of *Lupinus luteus* and *Vicia atropurpurea* plants. UV-B irradiation suppressed NRA by 60 % in leaves and 45 to 51 % in nodules. The leaves and roots of *Zea mays* L. (Quaggiotti *et al.* 2004), *Vigna radiata* (L.) Wilczek (Rajendiran and Ramanujam 2006) *Sesbania grandiflora* (L.) Pers. (Sudaroli Sudha and Rajendiran 2013a) and *Vigna unguiculata* (L.) Walp. c.v. BCP-25 (Sudaroli Sudha and Rajendiran 2013b) showed decreased values of NRA after exposure to UV-B

radiation in comparison with control seedlings. A decline in NRA was found related to changes in the protein synthesis and degradation (Bardizick *et al.*, 1971) or inactivation of the enzyme (Plaut 1974). However Marek, *et al.* (2008) in *Pinus sylvestris* L. needle reported an enhancement of NRA after exposure to UV-B irradiance. Guerrero *et al.* (1981) observed an accumulation of the nitrate consequent to UV-B induced inhibition of NRA, but was not confirmed by this study. Such a disparity was also reported by Balakumar *et al.* (1993) in UV-B and water stressed *Vigna unguiculata*. According to Ghisi *et al.* (2002), nitrate content of neither the leaf nor root was influenced by elevated UV-B. Nodulation was inhibited severely by UV-B as the number of root nodules (42 to 46 %) size and fresh mass of root nodules (50 %) were drastically reduced under controls. In contrast, nodulation and nitrogen fixation in three tropical grain legumes were not affected by exposure up to 62 % above ambient UV-B

(Samson *et al.* 2004). UV-B stress inhibited nitrogenase enzyme activity by 44 to 47 % in roots and by 51 to 55 % in root nodules. The UV-B irradiated legume on 30 as well as 45 DAS recorded reduction in all the parameters compared to control plants. However, the legume suffered heavily to UV-B on 30 DAS as the values were far below unstressed plants. To conclude, UV-B stress which was known to affect the metabolism of the aerial parts of the plants also disturbs the essential functions of the root system including *Rhizobium*-legume nitrogen fixation in *Phaseolus vulgaris* L. c.v. Prevail. As this legume was known for its poor nitrogen fixing ability, the feature of UV-B tolerance must be identified while breeding an improved nitrogen fixing variety.

ACKNOWLEDGEMENT

The authors thank Prof. Dr. V. Jayachandran, Head, Department of Botany, KMCPGS, Puducherry for providing research facilities.

REFERENCES

- Balakumar, T., Vincent, V.H.B., Paliwal, K., (1993) On the interaction of UV-B radiation (280-315 nm) with water stress in crop plants. - *Physiol. Plant.* **87**: 217-222.
- Bardizick, J.M., Marsh, H.V., Havis, J.R. (1971) Effects of water stress on the activities of three enzymes in maize seedlings. - *Plant Physiol.* **47**: 828-831.
- Caldwell, M.M., Bjorn, L.O., Bornman, J.F., Flint, S.D., Kulandaivelu, G., Teramura, A.H., Tevini, M. (1998) Effects of increased solar ultraviolet radiation on terrestrial ecosystem. - *Photochem. Photobiol.* **46**: 40-52.
- Chimphango, S.B., Musil, C.F., Dakora, F.D. (2003) Response of purely symbiotic and NO₃-fed nodulated plants of *Lupinus luteus* and *Vicia atropurpurea* to ultraviolet-B radiation. - *J.exp. Bot.* **54**: 1771-1784.
- Fernandes, M.S., Neves, M.C.P., Sa, M.F.M. (1982) Effects of supplemental nitrogen on nodulation, assimilation of nitrogen, growth and seed yield of *Phaseolus vulgaris* and *Vigna unguiculata*. In: *Biological Nitrogen Fixation Technology for Tropical Agriculture*, (ed.): Graham, P.H., Harris, J.C. (ed.): Pp. 317-326. CIAT, Cali, Colombia.
- Ghisi, R., Trentin, A.R., Masi, A., Ferretti, M. (2002) Carbon and nitrogen metabolism in barley plants exposed to UV-B radiation. - *Physiol. Plant.* **116**: 200-205.
- Graham, P.H., Halliday, J. (1977) Inoculation and nitrogen fixation in the genus *Phaseolus*. - In: *Exploiting the Legume-Rhizobium Symbiosis in Tropical Agriculture*. (ed.): Vincent, J. M., Whitney, A. S., Bosa, J. Pp 313-334. Univ. Hawaii Coll. Trop. Agric. Misc. Publ. 145.
- Guerrero, M.G., Veg, J.M., Losada, M. (1981) The assimilatory nitrate reducing system and its regulation. - *Annu. Rev. Plant Physiol.* **32**: 169-294.
- Jaworski, E.G. (1971) Nitrate reductase in intact plant tissue. - *Biochem. biophys. Res. Commun.* **43**: 1274-1279.
- Marek, K., Jerzy S., Heli K., Françoise M., Marja-Liisa S., Kaisa L., Minna T. (2008) Influence of solar UV radiation on the nitrogen metabolism in needles of Scots pine (*Pinus sylvestris* L.) *Environmental Pollution.* **156** (3): 1105-1111.
- Kulandaivelu, G., Lingakumar, K. (2000) Molecular targets of UV-B radiation in the photosynthetic membranes. - In: Yunus, M., Pathre, U., Mohanty, P. (ed.): *Probing photosynthesis, Mechanisms, Regulation and Adaptation*. Pp. 364-378. Taylor and Francis Publications, New York.
- Kokilavani, V., Rajendiran, K. (2013) Ultraviolet-B induced changes in the leaf epidermal and anatomical characteristics of *Vigna mungo* L.var. KM-2. *International Journal of Science and Nature.* **5** (1): 126-130.
- Lowry, O.H., Rosebrough, N.J., Farr, A.L., Randall, R.J. (1951) Protein measurement with the Folin phenol reagent. - *J. biol. Chem.* **193**: 265-275.
- Moore, S., Stein, W.H. (1948) Photometric method for use in the chromatography of amino acids. - *J. biol. Chem.* **176**: 367-388.
- Muthuchelian, K., Nedunchezian, N., Kulandaivelu, G. 1993. Effect of simulated acid rain on ¹⁴CO₂ fixation, ribulose-1,5- biphosphate carboxylase and nitrate and nitrite reductase in *Vigna sinensis* and *Phaseolus mungo*. - *Photosynthetica* **28**: 361-367.
- Plaut, Z. (1974) Nitrate reductase activity of wheat seedlings during exposure to and recovery from water stress and salinity. - *Physiol. Plant.* **30**: 212-217.
- Quaggiotti, S., Trentin, A. R., Vecchia, F. D. and Ghisi, R. 2004. Response of maize (*Zea mays* L.) nitrate reductase to UV-B radiation. *Plant Sci.* **167**: 107-116.
- Rachel, D., Santhaguru, K. (1999) Impact of UV-B irradiation on growth, nodulation and nitrate assimilation in *Vigna mungo* L. and *Vigna radiata* L. Wilczek. - In: Srivastava, G.C., Singh, K., Pal, M. (ed.): *Plant Physiology for Sustainable Agriculture*. Pp. 294-300. Pointer Publishers, Jaipur.
- Rajendiran, K., Ramanujam, M.P. (2000) Efficacy of triadimefon treatment in ameliorating the UV-B stress in green gram. - In: Khan, M. (ed.): *National Symposium on Environmental Crisis and Security in the New Millennium*. Pp. 41-42. National Environmental Science Academy, New Delhi.
- Rajendiran, K., Ramanujam, M.P. (2003) Alleviation of ultraviolet-B radiation-induced growth inhibition of green gram by triadimefon. - *Biol. Plant.* **46**: 621-624.
- Rajendiran, K., Ramanujam, M.P. (2004) Improvement of biomass partitioning, flowering and yield by triadimefon

- in UV-B stressed *Vigna radiata* (L.) Wilczek.- Biol. Plant. **48**: 145-148.
- Rajendiran, K., Ramanujam, M.P. (2006) Interactive effects of UV-B irradiation and triadimefon on nodulation and nitrogen metabolism in *Vigna radiata* plants. Biologia Plantarum. 50 (4): 709-712.
- Samson, B.M., Chimphango, F.B., Musil, C.F., Dakora, F.D. (2004) Effects of UV-B radiation on plant growth, symbiotic function and concentration of metabolites in three tropical grain legumes - Functional Plant Biol. **30**: 309-318.
- Shriner, D.S., Johnston, J.W. (1981) Effects of simulated acidified rain on nodulation of leguminous plants by *Rhizobium* spp. - Environ. exp. Bot. **21**: 199-209.
- Stewart, W.D.P., Fitzgerald, G.P., Burris, R.H. (1967) *In situ* studies on nitrogen fixation using the acetylene reduction technique. - Proc. nat. Acad. Sci. USA **58**: 2071-2078.
- Sudaroli Sudha, J and Rajendiran, K. (2013a) Effect of elevated UV-B irradiation on the nodulation and nitrogen metabolism in *Sesbania grandiflora* (L.) Pers. International Journal of Science and Nature. **4** (4): 664 - 667.
- Sudaroli Sudha, J. and Rajendiran, K. (2013b) Effect of elevated UV-B irradiation on the nodulation and nitrogen metabolism in *Vigna unguiculata* (L.) Walp. c.v. BCP-25. International Journal of Food, Agriculture and Veterinary Sciences. **3** (3): 77 - 81.
- Tevini, M., Iwanzik, W., Thoma, U. (1981) Some effects of enhanced UV-B radiation on the growth and composition of plants. - Planta **153**: 388-394.
- Vu, C.V., Allen, L.H., Garrard, L.A. (1981) Effects of supplementary UV-B radiation on growth and leaf photosynthetic reactions of soybean (*Glycine max*).- Physiol. Plant. **52**: 353-362.
- Woolley, J.T., Hicks, G.P., Hageman, R.H. (1960) Rapid determination of nitrate and nitrite in plant material. - J. agr. Food Chem. **8**: 481-482.
- Zar, J.H. (1984) Bio-statistical Analysis. - Prentice-Hall, Englewood Cliffs.