



TOXIC IMPACTS OF INSECTICIDE POLO ON HISTOPATHOLOGY OF GILL, LIVER AND KIDNEY AND GLYCOGEN CONTENTS OF A FRESHWATER FISH, *LABEO ROHITA*

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ABSTRACT

The broad spectrum insecticide Polo was used for this investigation to study the biochemical and histopathological effects in freshwater fish, *Labeo rohita*. The LC₅₀ value of Polo was found to be 1.9ppm for 96 hrs. under laboratory conditions and 1/10 of the corresponding LC₅₀ value was treated as sub lethal concentration. The fish showed decreased amount of glycogen content in liver and gonads (Testis and Ovary) and severe histopathological changes in gills, liver and kidney.

KEY WORDS: *Labeo rohita*, Polo, Biochemical, Histopathology.

INTRODUCTION

The widespread and indiscriminate use of chemicals for controlling the agricultural pests, many aquatic organisms like fishes, bivalves, prawns, crabs *etc.* are getting affected. The damage of different tissues and alteration in biochemical processes as well as disturbances in physiological processes were observed by these pollutants. Insecticide Polo is widely used in agriculture for controlling the crop pests. For this reason Polo was used in this investigation to study its toxic effects in fish. Acute exposure (24 to 96 hrs.) of Polo alters the normal architecture of tissues like gill, liver and kidney and decreases the glycogen contents of fish *Labeo rohita*.

MATERIALS & METHODS

The fish *Labeo rohita* were collected from Ganeshpur and Girna river dam near Chalisgaon city, Dist. Jalgaon, Maharashtra, India and were acclimatized to laboratory conditions for 15 days into 1000 liter capacity tank, previously washed with potassium permanganate and water temperature was 26.3±35degree centigrade and pH 7.0 to 7.2 maintained in aquarium. The acclimated healthy and active fishes were exposed to the acute toxicity (up to 96 hrs.) exposure to an insecticide Polo.

Toxicity assay

Group of 10 healthy and active fishes *Labeo rohita* were kept in a glass tank of de-chlorinated tap water. The fishes were treated with varying concentrations of Polo to determine the LC₅₀ values. 1/10 of the LC₅₀ values were taken as sub lethal concentration for the 24, 48, 72 and 96 hours of the experiment. To observe the histopathological changes, a group of individuals exposed to different concentrations as 3ppm for 24hrs. 2ppm for 48hrs. 1.5 ppm for 72hrs. and 1ppm for 96hrs. (Sub lethal concentrations are 0.3ppm for 24hrs., 0.2ppm for 48hrs., 0.15ppm for 72hrs. and 0.1ppm for 96hrs. that means 1/10 value of LC₅₀.) All individuals in control were maintained in toxicant free de-chlorinated water in the separate tank. During this experiment mortality was recorded for 24, 48,

72 and 96hrs. respectively. After exposure and completion of treatment, *Labeo rohita* were dissected and gills, liver, gonads and kidney were gently separated. The gills, liver and kidney tissues were fixed in Bouin's fluid and were processed by routine micro technique method and blocks were prepared. Then these prepared blocks of the tissues were cut with the thickness of 6µ on microtome. The ribbons of sections were spread on slide and these slides were further processed for double staining method. After the completion of these procedures, the slides were observed under microscope. Slides were observed under oil immersion for histopathological details. The tissues of liver, testis and ovaries were also processed for glycogen estimation by Anthrone reagent method (Dezwann and Zandee, 1972).

RESULTS & DISCUSSION

From the histopathological observations, the insecticide Polo had altered the structure of gills, liver and kidney. In fishes, the gills are the most important organ for respiration and osmoregulation and it is the first organ which the pollutant comes into contact. Gills are the main route through which toxicants enter into the body. Control gill (fig no. 1) showed a gill arch with double row of elongated laterally projecting primary gill filament, which in turn bear leaf like projections, the secondary gill lamellae. Each secondary gill lamellae is delicate flattened structure comprising of a pair of two layered epithelial sheets supported by pillar cells which in rows occupy the whole area of secondary gill lamellae. On the basement membrane of epithelial covering, neighboring pillar cells fuse to complete the lining of lamellae and blood channels, which connect the afferent and efferent lamellar vessels. The gill of the fish *Labeo rohita* exposed to the insecticide Polo (96hrs.) (Fig no.2) had showed marked histopathological changes characterized by swelling and degradation in respiratory epithelial cells and connective tissue cells. Connective tissue cells lost their normal cellular structure.

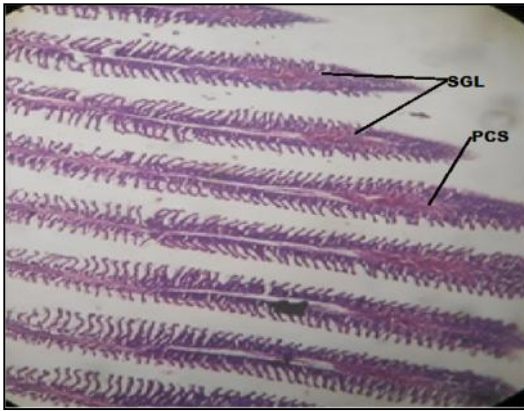


Fig.No 1
Control Gill of *Labeo rohita* at 100x
PCS – Pillar cells
SGL- Secondary gill lamellae

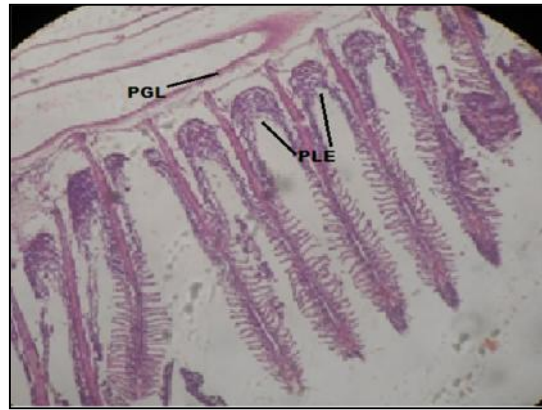


Fig No. 2
Polo treated Gills of *Labeo rohita* at 100x
PGL- Primary Gill lamellae
PLE – Primary lamellar epithelium

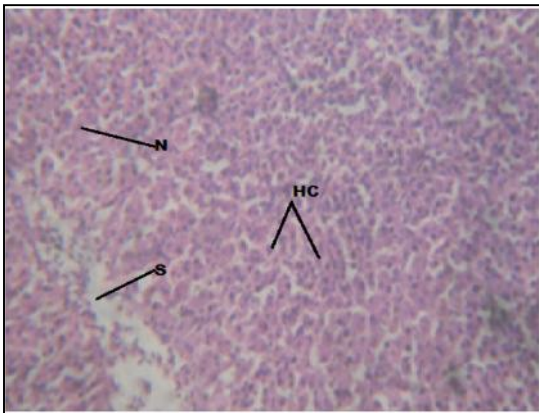


Fig.No 3
Control Liver of *Labeo rohita* at 100x
HC – Hepatocytes
N- Nucleus
S

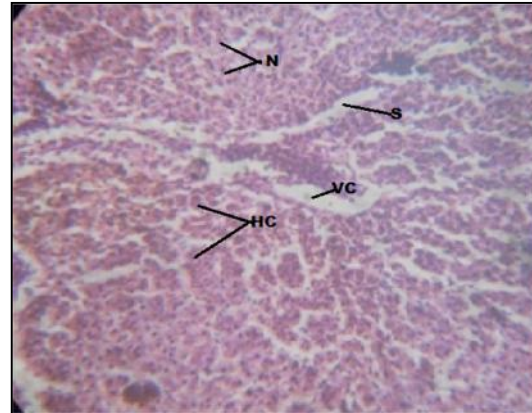


Fig.No 4
Polo treated liver of *Labeo rohita* at 100x
VC - Vacuolization
S - Sinusoid
HC

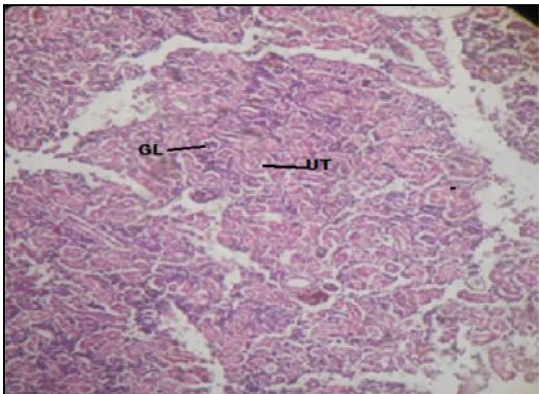


Fig.No 3
Control kidney of *Labeorohita* at 100x
UT – Uriniferous tubules
GL - Glomerulus

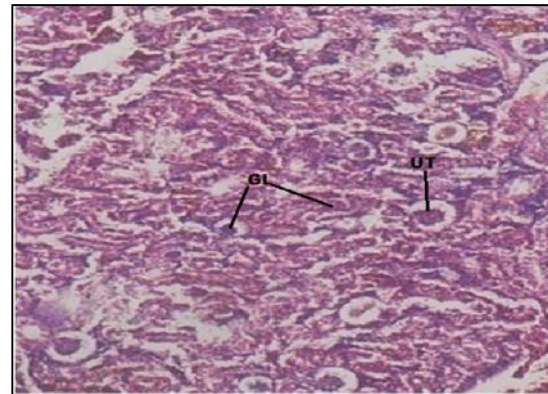


Fig.No 4
Polo treated Kidney of *Labeorohita* at 100x
UT

The secondary gill lamellar cell walls are disappeared, gill lamellae are shortened, and necrotic changes in respiratory epithelium of gills resulting in the development of vacuoles were seen. Cytoplasm showed disintegration at greater degree because of hyperplasia of respiratory

epithelial cells and reduced inter lamellar space. Severity of damage in the gills was found to be dose dependent. The liver is an important organ performing vital functions. Control liver (fig no. 3) showed large polygonal hepatocytes. These hepatocytes are separated by blood

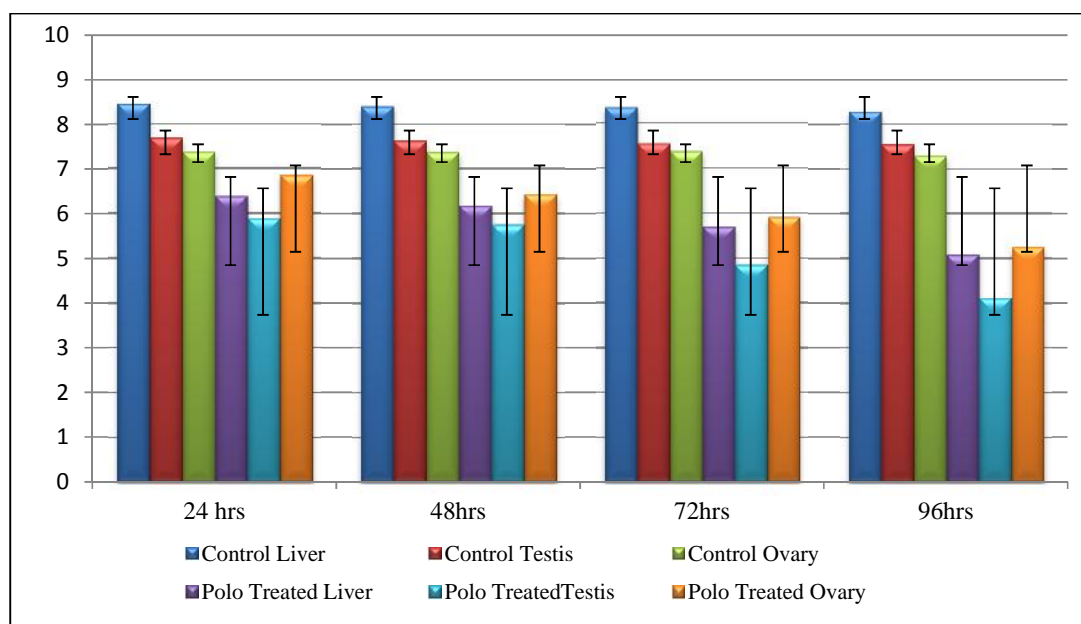
sinusoids. Each hepatocyte showed a distinct round and central nucleus with nucleoli and granular cytoplasm. Polo treated liver (fig no. 4) showed changes in its architecture. Hepatic cell diameter was changed, extensive cytoplasmic vacuolization had seen and nuclei become pyknotic and excentric. Histopathological alterations resulting from an exposure of Polo may affect the functional efficiency of the liver, leading to malfunctioning of several organ systems of the fish. Histology of control kidney (fig no. 5) of fish *Labeo rohita* showed distinct and normal size of proximal and distal conducting tubules of glomerulus with connective tissues. Whereas, Polo treated kidney (fig no. 6) showed damaged proximal and distal tubules as well as sinus appeared in connective tissues. The depletion in glycogen contents in liver and gonads after acute exposure by Polo were increased as the period of exposure increased. The maximum depletion occurred in the liver

followed by gonads. The results are summarized in table no1. There was depletion in glycogen content in liver and gonad as compared to the control. Liver is the vital organ of carbohydrate metabolism. It was affected by Polo. Decrease in glycogen values in liver was noticed by K. Suneetha (2011). Carbohydrates are the primary source of energy in stress condition. Depletion of glycogen may be due to direct utilization for energy generation demand caused by pesticidal stress. Total depletion of glycogen would result in the disruption of enzymes associated with carbohydrate metabolism. Decrease in glycogen content in liver were reported by Muley *et al.* (2007); Balaji and Chockalingam (1991), Amudha and Mahalingam (1999), Maruthi and Rao (2000), Mohammed A. Al. Khatani (2011), Venkatramana *et al.*, (2006) and Logaswamy *et al.* (2009).

Table- 1 Toxic impact of insecticide Polo on Glycogen contents in Liver and Gonads (testis and Ovary) of *Labeo rohita* after acute exposure.

Tissue	Treatment	Acute			
		24 hrs	48hrs	72hrs	96hrs
Liver	Control	8.4505	8.3939	8.3548	8.2829
		±0.008246***	±0.010392***	±0.145602***	±0.000078***
Testis	Control	7.6851	7.6249	7.5536	7.5431
		±0.006164***	±0.009165***	±0.008831***	±0.02607***
Ovary	Control	7.3768	7.3764	7.3762	7.2961
		±0.034058***	±0.03286***	±0.02898***	±0.05639***
Liver	Polo	6.3913	6.1766	5.6962	5.0832
		±0.08485***	±0.03065***	±0.02323***	±0.02236***
Testis	Polo	5.8921	5.7558	4.8438	4.1152
		±0.02898***	±0.02449***	±0.04795***	±0.02792***
Ovary	Polo	6.8571	6.4332	5.9102	5.2567
		±0.006928***	±0.006164***	±0.01077***	±0.008296***

- 1) Values expressed as mg/100g of wet wt. of tissues.
- 2) ± indicate S.D. of five observations.
- 3) Values are significant at $P < 0.001$ ***.



GRAPH: Variation in Glycogen content of Liver and Gonads (Testis and Ovary) of *Labeo rohita* after acute exposure to insecticide Polo

REFERENCES

- Hadi, A.A. and Alwan, S.F. (2012) Histopathological changes in Gills, Liver and kidney of freshwater fish, *Tilapia zillii*, exposed to aluminium IJPLS, 3(11):2071-2078
- Amudha, P. and Mahalingam, S. (1999) Effect of dairy effluent on the body consumption of *Cyprinus carpio* (Cyprinidae). J. Ecotoxicol. Environ. Monit., 9(1):03-08.
- Balaji, A. and Chokalingam, S. (1991) Effect of sublethal concentration of dairy effluent on an air breathing fish, *Channa punctatus*. Acad. Environ. Biol. 75-80.
- Cengiz, E.I. (2006) Gill and Kidney histopathology in the fresh water fish *Cyprinus carpio* after acute exposure to deltamethrin, Environmental Toxicology and Pharmacology, 21:1093-1096
- Das, B. K. and Mukherjee, S. C. (2000) A Histopathological Study of carp (*Labeo rohita*) exposed to hexachlorocyclohexane. Vet. Archiv. 70(4): 169-180.
- Desai, A. K., Joshi, U. M. Ambadkar, P. M. (1984) Histological observations on the liver of *Tilapia mosambica* after exposure to monocrotophos an organophosphorus insecticide Toxicol. Lett. 21:325 – 331
- Dezwann and Zandee (1972) Body distribution and seasonal changes in glycogen content of the common sea mussel *Mytilus edulis*. J. Environ. Biol., 7 (2) ,149-154.
- Dhanapakiam, P. and Juliet Premalatha (1994) Histopathological changes in the kidney of *Cyprinus carpio* to malathion and sevin J. Environ. Biol. 15(4), 283-287.
- Elezaby, M.M., Serafy, S.E., Aeckmann, R., Sharf Eldeen, K.H., Seddek, M.M. (2001) Effect of some toxicants on the fresh water fish *Oreochromis niloticus*, J. Egypt Ger. Soc. Zool. 36: 407-434.
- Velisek, J., Svobodova, Z., Piackova, V. (2009) Effects of acute exposure to bifenthrin on some haematological, biochemical and histopathological parameters of Rainbow trout (*Oncorhynchus mykiss*). Veterinari Medicina, 54(3): 131-137.
- Lloyd, R. (1992) Pollution and fresh water fish. The Buckland foundation. Fishing News Books , Oxford, pp35-40,77-81,107-110:122-124.
- Logaswamy, S. and Remia, K.M. (2009) Impact of cypermethrin and Ekalux on respiratory and some biochemical activities of a freshwater fish, *Tilapia mossambica*. Current Biotica, 3:65-73.
- Prashanth, M.S. (2011) Histopathological changes observed in the kidney of freshwater fish, *Cirrhinus mrigala* (Hamilton) exposed to Cypermethrin, RRST, 3(2): 59-65.
- Majid Askari Hesni, Ahrnad Savari, Ali Dadolahi Sohrab and Mohammad Sediq Mortazvi (2011) Gill Histopathological changes in milkfish (*Chanoschanos*) exposed to Acute toxicity of disel oil, WASJ 14(10): 1487-1492.
- Mandel, P.K., Kulshrestha, A.K. (1980) Histopathological changes induced by the sublethal sumithion in *Clarias batrachus* (Linn). Ind. J. Exp. Biol. 18:547-552.
- Maruthi, Y. A. and Subba Rao, M. V. (2000) Effect of distillery effluent on biochemical parameters of Fish, *Channa punctatus* (Bloch). J. Environ. Pollut. 7(2).111-113.
- Mohammed, A. Al.-Khatani (2011) Effect of an insecticide Abamectin on some biochemical characteristics of Tilapia fish, *Coreochromis niloticus*. Am. J. Agri & Biol. Sci., 6(1):62-68.
- Muley, D.V., Karanjkar, D. M. and Maske, S.V. (2007) Impact of industrial effluents on the biochemical composition of freshwater fish, *Labeo rohita*. Journal of Environ. Bio. 28, (2) 245-249.
- Ram Nayan Singh (2012) Histopathological alterations in the kidney of *Cyprinus carpio* after exposure to Dimethoate (EC 30%). Indian J. Sci. Res. 3(1) : 127-131.
- Sakrand, S.A. Jamal Al lail S.M. (2005) Fenvalerate induced histopathological and histochemical changes in the liver of *the Clarias gariepinus* JASR 1(3) : 263-267.
- Sakr, S.A., Hanafy, S.M. El-Desouky, N. E. (2001) Histopathological, Histochemical and physiological studies on the effect of the insecticide hostathion on the liver of catfish *Clarias gariepinus* Egypt J. Aquatic Biol. Fish 6(2) :103-124.
- Sarkar B. Chatterjee A., Adhikari S., Ayyappan S. (2005) :Carbofuran and cypermethrin induced histopathological alterations in the liver of *Labeo rohita* (Hamilton) and its recovery, Journal of Applied Ichthyology, 21,131-135.
- Sastry, K.V. and Sharma, S.K. (1979) The effect of endrin on the histopathological changes in liver of *Channa punctatus*. Bull. Environ. Contam. Toxicol. 20,674-677.
- Tilak, K.S., Veeraiah K. and Yacoub K. (2001) Studies on histopathological changes in the Gill, Liver and Kidney of *Ctenopharyngodon idella* (Valenciennes) exposed to technical fenvalerate and EC 20% Poll. Res. 20(3), 387-393.
- Velmurugan, B., Selvanayagam M., Cengiz E.I. and Unlu E., (2007) The effects of monocrotophos to different tissues of fresh water fish *Cirrhinus mrigala*. Bull. Environ. Contam. Toxicol., 78: 450-454.
- Venkatramana, G.V. Sandhya Rani, P.N. and Moorthy, P.S. (2006) Impact of Malathion on the biochemical parameters of gobiid fish, *Glossogobius giurinus* (Ham) J. Environ. Biol., 27:119-122.