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ISOLATION AND IDENTIFICATION OF NATIVE ANTAGONISTIC TRICHODERMA SPP. FROM RHIZOSPHERE OF GROUNDNUT, REDGRAM AND TOMATO

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ABSTRACT

A total of 27 rhizosphere soil samples were collected from healthy plants present in groundnut, redgram and tomato fields of Chittoor district. A total of nine *Trichoderma* spp. were obtained from 27 soil samples on *Trichoderma* selective medium on 7th day after inoculation. The isolates were designated as GRT-1 to GRT-5 for the five isolates of *Trichoderma* collected from groundnut rhizosphere soils, RRT-1 to RRT-2 for the two isolates collected from redgarm and TRT-1 to TRT-2 for the two isolates collected from tomato crop. *Trichoderma* spp. were characterized based on growth rate, GRT-2, GRT-3, RRT-2 and TRT-2 were categorized as very fast growing; GRT-1, RRT-1 and TRT-1 as fast growing and GRT-4, GRT-5 as medium growing. These nine *Trichoderma* isolates were grouped. *Trichoderma* isolates GRT-2, GRT-4, GRT-5 were grouped under *Trichoderma virens*, RRT-1 and GRT-3 as *Trichoderma harzianum*. TRT-2 and RRT-2 as *Trichoderma asperillum*. Isolates GRT-1 and TRT-1 identified individually as *Trichoderma longibrachiatum*, *Trichoderma pseudokoningii*

KEY WORDS: Rhizosphere, Trichoderma spp., antagonistic.

INTRODUCTION

Trichoderma spp. commonly available in soil and root ecosystems has gained immense importance since last few decades due to biological control ability against several plant pathogens. The potential of Trichoderma spp. as biocontrol agents of plant diseases was first recognised in early 1930's. Fungal spp. belonging to the genus Trichoderma is worldwide in occurrence and belongs to division Ascomycota, subdivision Pezizomycotina, class Sordariomycetes and order Hypocreales. Trichoderma is characterized as rapidly growing colonies bearing tufted or postulate, repeatedly branched conidiophores with lageniform phialides and hyaline or green conidia born in slimy heads (Shalini and Kotasthene, 2007) and can be easily isolated from soil preferably in acidic soils with pH 3.5 to 4.5. The success of Trichoderma strains as biocontrol agents is due to their high reproductive capacity, ability to survive under very unfavourable conditions, efficiency in the utilization of nutrients, capacity to modify the rhizosphere, strong aggressiveness against phytopathogenic fungi, and efficiency in promoting plant growth and defense mechanisms. These properties have made Trichoderma a ubiquitous genus present in any habitat and at high population densities. Several advantages of using Trichoderma in managing soil borne plant pathogens are reported by different workers such as, ecofriendly (Gaur et al., 2005), effective in managing diseases caused by soil borne plant pathogens which cannot be easily controlled by chemicals (Manoranjitham et al., 2001), ease and cost effective mass Culturing of antagonists (Gaur et al., 2005 and Das et al.,

2006), growth promoting effect (Pan and Bhagat, 2007) and long lasting effective disease management (Sarojini et al., 2007). Several strains of T. viride had a significant reducing effect on plant diseases caused by pathogens such as Rhizoctonia solani, Sclerotium rolfsii, Pythium aphanidermatium, Fusarium oxysporum, F. culmorum and Gaeumannomyces graminis var. tritici under greenhouse and field conditions (Bassim et al., 1999). Soil borne pathogens causing diseases like collar rot, stem rot, root rot, damping off and wilt in crops like Ground nut, Redgram and Tomato. Root colonization ability of Trichoderma spp, is another important properties which make them strong competitive colonizer in the rhizosphere of plant and protect the root infection by pathogens. Thus in present study we concentrated on isolation, identification of Trichoderma isolates existing in Ground nut, Redgram and Tomato fields in few regions of Chittoor.

MATERIALS & METHODS

Isolation and Identification of Trichoderma Species

Soil samples were collected from 27 rhizosphere soil samples from different cropping systems such as groundnut, redgram and tomato fields in Chittoor district, Andhra Pradesh. Among 27 soil samples, 13 samples from groundnut, 7 samples from redgram and 7 samples from tomato rhizosphere region were collected. For rhizospheric soil, plants were gently uprooted, soil tightly adhering the roots was collected, mixed and composite mixture of soil of the region was obtained. The pH of soil was determined in 1:2 (soil: water) ratio, keeping 30 minutes of equilibration. Collected soil samples were air dried for 4 hour and isolation was done by serial dilution technique. Trichoderma Selective Medium (TSH) was used for identification of the isolates of *Trichoderma* (Elad and chet, 1983). One ml of soil suspension was taken with the help of 5ml sterilized pipette and poured on the Petri- plate seeded with TSM. The plates were incubated at $28 + 2^{\circ}$ C for 5 days. Observation on the appearance of colonies was recorded from 3^{rd} to 5th day. Individual colonies were picked up and maintained in pure culture for further study. *Trichoderma* species were identified and examined under compound microscope on the basis of their cultural and morphological character (Park *et al.*, 2005) and were maintained on PDA slants at 4°C for subsequent studies.

RESULTS & DISCUSSION

A total of 27 rhizosphere soil samples were collected from healthy plants present in groundnut, redgram and tomato fields of Chittoor district. Among 27 soil samples, 13 samples from groundnut, 7 samples from redgram and 7 samples from tomato rhizosphere region were collected. From these soil samples, five *Trichoderma* isolates were isolated from soils collected from groundnut rhizosphere region, two from redgram and two from tomato (Table 1 and 2, Fig1). The isolates were designated as GRT-1 to GRT-5 for the five isolates of *Trichoderma*

FABLE 1. Soil Samples collected from	rhizosphere region of	of groundnut, redgram	and tomato fields from	different				
and a late in Children and interior								

S. No	Name of the mandal	Name of the village	crops	No of soil samples collected
1 Irala	Tellagundlapalli	Groundnut	2	
		Redgram	1	
		Groundnut	1	
2	2 Rompicherla	Peddagottigallu	Redgram	1
		Tomato	1	
3	3 Vadamalpeta	Devaraju kandriga	Groundnut	2
5			Redgram	1
4	Pakala	Pachipala palli	Groundnut	3
5	5 Ramachandrapuram	Domonurom	Groundnut	1
5		Ramapuram	Tomato	2
			Groundnut	1
6 Rompicharla	Bommaiahgari palle	Redgram	1	
		Tomato	1	
	7 Pulicherla	Gaddamvaripalle	Groundnut	1
7			Redgram	1
			Tomato	1
8 Pileru	Agraharam	Tomato	1	
		Groundnut	2	
0	0 Timupoti murol	Peruru	Redgram	2
	i nupati tutai		Tomato	1
		Total		27

TABLE 2. List of Trichoderma spp. isolated from soil samples collected from different mandals in Chittoor Dist.

S. No	Isolate No	Name of the Mandal	Name of the Village
1	GRT-1	Irala	Tellagundlapalli
2	GRT-2	Ramachandrapuram	Ramapuram
3	GRT-3	Rompicharla	Bommaiahgari palle
4	GRT-4	Pileru	Agraharam
5	GRT-5	Pulicherla	Gaddamvaripalle
6	RRT-1	Vadamalpeta	Devaraju kandriga
7	RRT-2	Rompicharla	Bommaiahgari palle
8	TRT-1	Rompicherla	Peddagottigallu
9	TRT-2	Tirupati rural	Peruru

GRT = Groundnut Rhizosphere Trichoderma, RRT = Redgram Rhizosphere Trichoderma, TRT = Tomato Rhizosphere Trichoderma



FIGURE 1. Pure cultures of Trichoderma isolates from rhizosphere region of groundnut, tomato and redgram

Pure cultures of Trichoderma isolates from rhizosphere region of groundnut, tomato and redgram collected from groundnut rhizosphere soils, RRT-1 to RRT-2 for the two isolates collected from redgarm and TRT-1 to TRT-2 for the two isolates collected from tomato crop. In similar study several scientists reported the isolation of Trichoderma spp. from rhizosphre soils of different crops. Gajera et al. (2011) isolated 12 isolates of three strains (T. virens, T. viride and T. harzianum) from rhizosphere soils of groundnut crop. Sundaramoorthy and Balabaskar (2013) isolated fungal native antagonists from tomato rhizosphere soils by serial dilution technique using Trichoderma selective medium and identified as T. hamatum, T. harzianum, T. koningi, T. longiconis and T. viride. Depending on the growth rate on PDA medium, nine isolates of Trichoderma spp. were categorized into three groups, viz., very fast, fast, and medium in radial growth. It is evident from Table 2 that the isolates GRT-2, GRT-3, TRT-2, RRT-2 with 90 mm diameter radial growth within 48 h were categorized as very fast growing and the isolates, GRT-1, RRT-1, TRT-1 which exhibited 90 mm in diameter within 72 h were categorized as fast growing, whereas the isolates GRT-4 and GRT-5 exhibited 90 mm in diameter within 96h were categorized as medium growing isolates. Singh et al. (2006) obtained twenty seven isolates of T. harzianum from soil samples collected randomly from fallow agricultural fields throughout the Punjab and studied their growth rate on PDA medium and were categorised as medium, fast and very fast growing. Nine Trichoderma isolates were identified according to the identification key (Rifai, 1969) based on branching of conidiophores, shape of the phialides, emergence of phialospores, and shape of phialospores.. These nine Trichoderma isolates were grouped in to different isolates. Trichoderma isolates GRT-2, GRT-4, GRT-5 were grouped under Trichoderma virens, RRT-1 and GRT-3 as Trichoderma harzianum. TRT-2 and RRT-2 as Trichoderma asperillum. Isolates GRT-1 and TRT-1 identified individually as Trichoderma Trichoderma longibrachiatum, pseudokoningii respectively. Muthukumar and Pratibha Sharma (2011) used morphological description for characterization and grouping of Trichoderma isolates.

REFERENCES

Basim, H., Stark, S.B. and Yegen, O. (1999) Efficacy of biological fungicide against seedling root rot of pathogens of cotton. Phytopathology. 70: 137-144.

Das, B.C., Das, B.K., Pranab, D. and Sarmah, D.K. (2006) Bioformulation of *Trichoderma harzianum* Rifai for management of soyabean stem rot caused by *Rhizoctonia solani* Khunn. Journal of Biological Control. 20 (1):57-64. Elad, Y and Chet, I. (1983) Improved selective media for isolation of *Trichoderma* or *Fusarium* spp. Phyto parasitica, 11, 55-58.

Gajera, H., Kalu, R. and Dinesh, V. (2011) Bioefficacy of *Trichoderma* isolates against *Aspergillus niger* Van Tieghem inciting collar rot in groundnut (*Arachis hypogaea* L.). Journal of Plant Protection Research. 51: 240-247.

Gaur, R.B., Sharma, R.N., Sharma, R.R and Singh, V.G. (2005) Efficacy of *Trichoderma* for *Rhizoctonia* root rot control in chickpea. Journal of Mycology and Plant Pathology, 35 (1):144-150.

Manoranjitham, S.K., Prakasam, V. and Rajappan, K. (2001) Biocontrol of damping off of tomato caused by *Pythium aphanidermatum*. Indian Phytopathology, 54:59-61.

Muthukumar, A. and Prathibha Sharma (2011) Molecular and morphological characters: An appurtenance for antagonism in *Trichoderma* spp. African Journal of Biotechnology, 10(22):4532-4543.

Park, M.S., Seo, G.S., Lee, K.H., Bae, Yu, S.H. (2005) Morphological and Cultural characteristics of *Trichoderma* spp. associated with green mold of Oyster mushroom in Korea. Plant Pathology Journal 21, 221-228.

Pan, S. and Bhagat, S. (2007) Antagonistic potential of *Trichoderma* and *Gliocladium* spp. from West Bengal. Journal of Mycology and Plant Pathology, 37(2):235-239.

Rifai, M.A. (1969) Revision of the Genus *Trichoderma*. Mycological Paper. 166: 1-56.

Sarojini, K., Chakravarthy and Nagamani, A. (2007) Efficacy of non-volatile and volatile compounds of *Trichoderma* species on *Rhizoctonia solani*. Journal of Mycology and Plant Pathology. 37(1): 82-86.

Singh, H.B. (2006) Management of plant pathogens with microorganisms, Journal of Mycology and Plant Pathology, 36: 373-384.

Shalini, S. and Kotasthane, A.S. (2007) Parasitism of *Rhizoctonia solani* by strains of *Trichoderma* spp. Electronic Journal of Environmental, Agriculture and Food Chemistry, 6 (11): 1220-1225.

Sundaramoorthy, S. and Balabaskar, P. (2013) Biocontrol efficacy of *Trichoderma* spp. against wilt of tomato caused by *Fusarium oxysporum* f. sp. *Lycopersici*. Journal of Applied Biology and Biotechnology.1 (3): 36-40.