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SCREENING OF BARLEY (HORDEUM VULGARE L.) GENOTYPES FOR PHYSIOLOGICAL TRAITS UNDER DROUGHT STRESS

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ABSTRACT

Barley (*Hordeum vulgare* L.) is one of the earliest coarse cereal crops. Stress due to drought condition is one of the major factors responsible for low crop productivity. In the present investigation, thirty barley genotypes comprised of two and six row type including checks (AZAD, K 560, K 603, LAKHAN, RD 2624 and RD 2660) were studied for physiological traits *viz.*, Normalised Difference Vegetative Index (NDVI), canopy temperature and SPAD chlorophyll content, at anthesis and 15 days after anthesis of the growth period, under drought stress conditions. The physiological parameters *viz.*, NDVI (0.40-0.76 and 0.49-0.82); SPAD chlorophyll content (17.2-21.4 and 14.5-18.7) and canopy temperature (19.6-43.4 and 17.1-46.8 °C), at the time of anthesis and 15 days after anthesis, respectively, showed significant variation among the genotypes for all the characters studied. On the basis of physiological parameters, the promising genotypes for drought tolerance can be selected and utilized as elite breeding material to transfer drought tolerance through crossing programs. Hence, these physiological traits could be considered as suitable selection criteria for the development of high yielding barley varieties under drought condition.

KEYWORDS: NDVI, canopy temperature, SPAD chlorophyll content.

INTRODUCTION

Barley (Hordeum vulgare L.) is first domesticated and an important coarse cereal crop ranks fourth among cereal crops after rice, maize and wheat. It is widely adapted crop which is grown throughout tropical and temperate regions of the world. Drought stress due to limited water supply is one of the most important environmental factors which affect crop productivity worldwide. Major losses in crop production are due to drought stress conditions. The drought tolerance of a crop is essentially linked to its ability to access soil water and to use it most productively (Richards et al., 2010). Severity of water stress during the grain-filling period decreased the net photosynthetic rate of the flag leaf of barley (Sanchez et al., 2002; Farooq et al., 2017). Barley is considered to be the most drought tolerant grain crop and a simple genetic model for evaluating mechanisms of drought tolerance and associated agronomic and physiological traits (Forster et al., 2004). Chlorophyll content is one of the major factors affecting photosynthetic capacity and indicator of photosynthetic capability of plant tissues (Nageswara et al., 2001). Reduction in chlorophyll content of plant under drought stress has been dependence on different genotypes and magnitude of stress and stress duration (Jagtap et al., 1998). Chlorophyll content in both resistant as well as sensitive genotypes reduced under drought stress condition. Genotypes showed resistance behavior to drought stress conditions had high in chlorophyll content (Sairam et al., 1997). Chlorophyll was one of the major components of chloroplasts, and positively correlated with photosynthesis rate. The reduction of chlorophyll content in drought conditions was considered a typical symptom of oxidative stress that could be the result of pigments photo-bleaching and chlorophyll degradation (Anjum *et al.*, 2011).

According to Blum (1996) under water-limited conditions comparison of canopy temperature depression relative to air temperature makes it possible to detect genotypic differences related to the genetic improvement of cereals. Balota et al. (2007) have proposed low canopy temperature (CT) is the most important mechanism for drought adaptation. Roohi et al. (2015) observed that CT showed negative relationship with grain yield and the triticale, wheat and barley genotypes with low CT, produced more grain yield, suggested that the cooler canopy results in better adaptation to water stress. Chaudhari et al. (2017) reported that the wheat lines with low CT in grain filling produced more grain yields. Staygreen in the post anthesis phase is reported to be associated with drought tolerance in several crops (Campos et al., 2004).

NDVI is calculated using wavelengths within the NIR (near infrared) and VIS (visible) regions of the electromagnetic spectrum. NDVI relates to leaf chlorophyll content, leaf nitrogen and ultimately the photosynthetic capacity of the plant (Tattaris *et al.*, 2016). CT, which is measured from emitted infra- red radiation, can be used as a tool to indirectly evaluate the transpiration rate, water status and stomatal conductance of a plant (Penuelas *et al.*, 1997) while NDVI can estimate relative crop biomass at different growth stages (Babar *et al.*, 2006) as well as nitrogen deficiency and crop

senescence rate (Olivares-villegas *et al.*, 2007). CT, chlorophyll content and NDVI (Normalized Difference Vegetative Index) have been effectively combined for rapid screening of drought and heat tolerance in wheat (Reynolds *et al.*, 2007); hence these parameters were studied in barley genotypes under drought stress conditions. Therefore, the current study examined the response of yield and physiological traits to drought occurred in barley plants, according to the expected situations of climatic change.

MATERIALS & METHODS

Thirty barley genotypes comprised of two and six row type including six checks viz. AZAD,K 560, K 603, LAKHAN, RD 2624 and RD 2660 were studied for physiological parameters at anthesis and 15 days after anthesis. The experiment was conducted during *rabi* season of 2016-17 in a randomized block design (RBD) with three replications under drought conditions at research crop area of Wheat and Barley section, CCS Haryana Agricultural University, Hisar.

Each plot consisted of four rows of 2.5 m length with 23 cm row to row spacing. These barley genotypes were evaluated for physiological parameters and grain yield per plot given below as:

- **1. Normalized difference vegetation index (NDVI)**: NDVI was recorded using optical handheld Instrument Green SeekerTM sensor (Trimble industries, Inc.).
- **2. Canopy temperature (CT)**: CT was measured during 12.00 and 14.00 hrs with hand-held infrared thermometer Sixth Sense LT300 IRT and three readings for each plot were averaged to get true representative values.
- **3.** Chlorophyll content index (SPAD): Mean chlorophyll of three tagged plants flag leaves were determined by a SPAD-502 chlorophyll meter (Konica Minolta Sensing, Osaka, Japan).
- **4. Grain yield/plot** (**g**): Grain yield was recorded after harvesting and thrashing the plot. The thrashed grains were cleaned and yield was recorded in gram.

The data was subsequently analyzed to determine the variability and phenotypic correlation coefficient using the OP STAT software of CCS Haryana Agricultural University, Hisar.

RESULTS

Variability for physiological and grain yield parameters

The data was collected for the physiological parameters at the time of anthesis and 15 days after anthesis of the thirty

genotypes including six checks. The data obtained was analyzed for one factor analysis using the online OPSTAT software. Physiological parameters and yield was significantly influenced by drought at anthesis and 15 days after anthesis but the reduction was found more at 15 days after anthesis. The data for NDVI (Normalised Difference Vegetative Index), canopy temperature and SPAD chlorophyll content, at anthesis and 15 days after anthesis of the growth period showed huge variation (Table 1). NDVI ranged from 0.40 \pm 0.01 (2nd GSBSN 02) to 0.76 \pm 0.02 (LAKHAN) at anthesis and 0.49 \pm 0.04 (2nd GSBSN 02) to 0.82 ±0.001 (2nd GSBSN 94) at 15 days after anthesis. Canopy temperature ranged from 14.5±0.19 °C (2nd GSBSN 66) to 18.7±0.46 °C (2nd GSBSN 02) at anthesis and 17.2±0.27 to 21.4±0.32 °C at 15 days after anthesis for LAKHAN and PL 890, respectively, whereas, SPAD chlorophyll content varied between 19.6±0.93 (2nd GSBSN 02) to 43.4±1.03 (2nd GSBSN 66) SPAD unit at anthesis and 17.1±0.49 to 46.8±1.39 SPAD unit at 15 days after anthesis for 2nd GSBSN 02 and RD 2660, respectively.

Drought condition showed a great variation between grain yield per plot; ranged between 401.7 ± 33.71 g (2nd GSBSN 23) to 741.7±46.67 g (NDB 3) at the time of physiological maturity after harvesting and threshing. The genotypes NDB 3(741.7±46.67 g), KB 1326 (720.0±20.82 g), JB 485 (686.7±46.04 g), 2nd GSBSN 28 (676.7±63.86), KB 1317 (656.7±60.58 g) and 2nd GSBSN 93 (648.3±46.76 g) showed maximum grain yield per plot among all genotypes under drought conditions.

Correlation coefficient analysis

Correlation coefficients for all possible pairs of characters were calculated as per the procedure by Fisher and Yates (1963) using OPSTAT software of CCS Haryana Agricultural University, Hisar (Table 2). NDVI at anthesis and 15 days after anthesis showed negative correlation with canopy temperature at anthesis and 15 days after anthesis. The plants with higher biomass i.e. more NDVI have less canopy temperature. Hence it revealed that the plants with cooler canopy were found to be more drought stress tolerant. Canopy temperature at 15 days after anthesis also showed negative correlation with SPAD chlorophyll content at anthesis and 15 days after anthesis, while there is no significant correlation between canopy temperature at anthesis and SPAD chlorophyll content at anthesis and 15 days after anthesis (Table 2). SPAD chlorophyll content was positively correlated with SPAD chlorophyll content at 15 days after anthesis.

TADLE 1. Variation for	-	monomeetens and a	main reislel in hanlars	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	duarraht and dition
TABLE 1: Variation for	physiological	parameters and g	grann yleid in barley	genotypes under	arought condition

	At anthesis			15 Days aft	er anthesis		Grain
Genotypes	NDVI-I	CT-I (°C)	SPAD-I	NDVI-II	CT-II (°C)	SPAD-II	yield/plot (g)
BL 1122	0.55 ± 0.02	16.0 ± 0.48	35.8 ± 1.92	0.71 ± 0.07	19.3±0.69	29.9 ± 0.84	481.7±37.68
BL 1163	0.64 ± 0.03	15.1±0.47	32.7±1.78	0.77 ± 0.02	18.8 ± 0.58	27.7±0.69	625.0±42.53
JB 481	0.57 ± 0.02	16.2 ± 0.19	35.9 ± 1.26	0.78 ± 0.02	19.1±0.38	34.5±0.89	636.7±40.96
JB 482	0.59 ± 0.03	15.8 ± 0.62	28.1 ± 2.06	0.76 ± 0.03	18.6 ± 0.14	28.9 ± 3.51	493.3±23.15
JB 483	0.51 ± 0.05	17.3±0.64	20.5±1.03	0.69 ± 0.02	19.2 ± 1.11	30.1±1.48	463.3±20.28
JB 484	0.43 ± 0.02	16.8 ± 0.45	27.4 ± 1.71	0.58 ± 0.02	19.7±0.89	26.8 ± 2.71	613.3±28.48
JB 485	0.49 ± 0.02	16.5±0.15	29.4±1.28	0.62 ± 0.03	21.2±0.49	29.6±1.93	686.7±46.04
KB 1055	0.53 ± 0.07	17.9±0.27	27.6±2.76	0.65 ± 0.05	19.8±0.26	26.9±1.72	526.7±40.55
KB 1302	0.67 ± 0.02	16.7±0.48	29.4±1.34	0.80 ± 0.02	18.1±0.26	25.1±1.77	628.3±70.97
KB 1317	0.55 ± 0.03	15.0±0.45	23.0±1.12	0.74 ± 0.03	18.0 ± 0.14	24.5±1.23	656.7±60.58
KB 1326	0.62 ± 0.03	15.6±0.25	38.7±0.94	0.73 ± 0.03	18.7±0.23	39.2±1.11	720.0±20.82
KB 1401	0.58 ± 0.04	16.5±0.33	26.6±0.67	0.80 ± 0.01	18.8 ± 0.21	26.5 ± 2.09	618.3±42.07
NDB 1	0.66 ± 0.03	14.5±0.30	32.7±1.10	0.78 ± 0.01	18.4 ± 0.46	26.7±0.67	583.3±18.56
NDB 2	0.48 ± 0.02	16.0±0.31	31.1±1.14	0.69 ± 0.03	21.2±0.18	29.5±1.56	431.7±13.64
NDB 3	0.51 ± 0.02	15.6±0.46	23.7±1.98	0.62 ± 0.02	19.1±0.34	18.9 ± 1.21	741.7±41.67
PL 751	0.40 ± 0.02	17.4±0.21	24.7 ± 2.24	0.60 ± 0.03	20.4±0.67	25.6±1.82	551.7±34.44
PL 890	0.45 ± 0.01	18.6 ± 0.87	27.7±1.48	0.53 ± 0.02	21.4±0.32	26.8±1.96	481.7±10.93
2 nd GSBSN 02 (2015)	0.40 ± 0.01	18.7±0.46	19.6±0.93	0.49 ± 0.04	21.0±0.81	17.1±0.49	521.7±34.92
2 nd GSBSN 23 (2015)	0.60 ± 0.04	17.3±0.51	23.2±1.06	0.80 ± 0.02	20.0±0.44	23.6±1.09	401.7±33.71
2 nd GSBSN 28 (2015)	0.66 ± 0.02	15.3±0.23	29.5±1.23	0.74 ± 0.02	17.9±0.68	37.0±0.51	676.7±63.86
2 nd GSBSN 60 (2015)	0.48 ± 0.04	16.6±0.26	37.9±0.66	0.72 ± 0.05	19.6±0.15	22.2±0.64	510.0±47.26
2 nd GSBSN 66 (2015)	0.49 ± 0.02	14.5±0.19	43.4±1.03	0.70 ± 0.05	18.9±0.48	31.4±2.31	575.0±59.23
2 nd GSBSN 93 (2015)	0.52 ± 0.02	15.6±0.27	24.0±1.57	0.69 ± 0.08	19.6±0.77	21.1±1.19	648.3±46.76
2 nd GSBSN 94 (2015)	0.62 ± 0.03	15.2±0.67	29.6±1.74	0.82 ± 0.00	19.4±0.19	26.7±0.81	520.0±55.08
AZAD	0.48 ± 0.01	15.5±0.37	34.1±2.07	0.61 ± 0.01	19.4±0.09	30.1±2.56	555.0±60.07
K 560	0.54 ± 0.04	15.7±0.15	30.9±1.43	0.69 ± 0.02	19.5±0.24	30.5±0.90	540.0±45.83
K 603	0.65 ± 0.01	16.1±0.46	31.3±1.95	0.77±0.02	17.6±0.19	22.6±1.36	501.7±39.41
LAKHAN	0.76 ± 0.02	15.6±0.27	27.8±0.88	0.79 ± 0.01	17.2±0.27	26.3±1.53	536.7±49.10
RD 2624	0.47 ± 0.05	17.0±0.53	32.5±1.92	0.59 ± 0.02	20.1±0.61	21.6±1.28	550.0±28.87
RD 2660	0.52 ± 0.03	15.2±0.74	35.0±2.00	0.67 ± 0.04	20.3±0.76	46.8±1.39	523.3±40.96
C.D.	0.08	1.02	4.19	0.09	1.13	3.90	120.22
E(m)	0.03	0.36	1.49	0.03	0.40	1.39	42.36
SE(d)	0.03	0.51	2.11	0.04	0.57	1.96	59.90
C.V.	10.77	4.46	10.00	8.66	4.16	9.96	12.95

TABLE 2: Correlation coefficient between grain yield and physiological parameters in barley under drought stress

	environm	ient			
NDVI	NDVI	CT	CT	SPAD	SPAD
at anthesis	15 days after	at anthesis	15 days after	at anthesis	15 days after
	anthesis		anthesis		anthesis
1.000					
0.838^{**}	1.000				
-0.789**	-0.692**	1.000			
-0.490**	-0.553**	0.562^{**}	1.000		
0.140^{NS}	0.225 ^{NS}	-0.136 ^{NS}	-0.486**	1.000	
0.178^{NS}	0.187 ^{NS}	-0.050^{NS}	-0.376*	0.514^{**}	1.000
	at anthesis 1.000 0.838 ^{**} -0.789 ^{**} -0.490 ^{**} 0.140 ^{NS}	NDVI at anthesis NDVI 15 days after anthesis 1.000 0.838** 0.0789** -0.692** -0.490** -0.553** 0.140 ^{NS} 0.225 ^{NS}		$\begin{array}{c ccccc} NDVI & NDVI & CT & CT & cT \\ at anthesis & 15 days after at anthesis & 15 days after \\ anthesis & 15 days after at anthesis & 15 days after \\ anthesis & 15 days after \\ 0.838^{**} & 1.000 & & \\ 0.789^{**} & -0.692^{**} & 1.000 & & \\ -0.789^{**} & -0.692^{**} & 1.000 & & \\ -0.490^{**} & -0.553^{**} & 0.562^{**} & 1.000 & \\ 0.140^{NS} & 0.225^{NS} & -0.136^{NS} & -0.486^{**} & \\ \end{array}$	$\begin{array}{c ccccc} NDVI & NDVI & CT & CT & SPAD \\ at anthesis & 15 days after & at anthesis & 15 days after & at anthesis \\ anthesis & anthesis & 15 days after & at anthesis \\ anthesis & anthesis & anthesis \\ \hline 1.000 \\ 0.838^{**} & 1.000 \\ -0.789^{**} & -0.692^{**} & 1.000 \\ -0.490^{**} & -0.553^{**} & 0.562^{**} & 1.000 \\ -0.490^{**} & 0.225^{NS} & -0.136^{NS} & -0.486^{**} & 1.000 \\ \hline \end{array}$

DISCUSSION

The huge range of variation was found between genotypes for Normalised Difference Vegetative Index (NDVI), canopy temperature and SPAD chlorophyll content, at anthesis and 15 days after anthesis (Table 1). The data showed that the SPAD chlorophyll content declined at 15 days after anthesis when compared to SPAD chlorophyll content at the time of anthesis. Gonzalez *et al.* (2010) also studied that the loss of chlorophyll induced by water deficit reached 11%. One of the reasons for the reduction of the amount of chlorophyll is chlorophylase enzyme activity, which is the expression of this enzyme induced in stress conditions (Ranjan *et al.*, 2001). The loss of chlorophyll in the final stages of grain filling was as a result of oxidative stress and remobilization of nitrogen to grains (Oncel *et al.*, 2000). Physiological parameters such as and the Normalized Difference Vegetation Index (NDVI) and chlorophyll content (SPAD) are directly related with nitrogen contents in leaves of plant and indicate plant fitness under stress; useful indicators for selection of superior genotypes under stress condition to enhance stress tolerance (Anithakumari *et al.*, 2012, Cabello *et al.*, 2013).

Bilge *et al.* (2008) reported pattern of changes in CTD values during pre-anthesis, anthesis and grain filling stages. Our results are in agreement with the finding of Talebi (2011), water stress affect positively canopy temperature. Previous studies on wheat presented a positive phenotypic correlation of Normalized Difference Vegetation Index, chlorophyll content and canopy

temperature with biomass production and grain yield (Gutierrez et al., 2010; Bowman et al., 2015).

It is well established that drought stress impairs numerous metabolic and physiological processes in plants (Levitt, 1980). Drought stress leads to reduction in growth, grain yield per plot, chlorophyll content, NDVI and canopy temperature (Li *et al.*, 2006; Yang *et al.*, 2006; Balota *et al.*, 2007). Grain yield and physiological traits associated with drought tolerance are suitable indicators for selection of drought tolerant genotypes and can minimize the losses in crop productivity due to drought stress. It was concluded that NDVI, canopy temperature and SPAD chlorophyll content could be considered as reliable indicators for screening barley genotypes for drought tolerance.

CONCLUSION

The genotypes NDB 3, KB 1326, JB 485, 2^{nd} GSBSN 28 (2015), KB 1317 and 2^{nd} GSBSN 93 (2015) showed maximum grain yield per plot among all genotypes under drought conditions and performed better than the checks. The parameters such as canopy temperature, chlorophyll content and Normalized Difference Vegetation Index can prove to be beneficial as non-destructive methods of measuring plant drought stress and can screen field grown barley genotypes under stress condition.

REFERENCES

Anithakumari, A.M., Nataraja, K.N., Visser, R.G. and Vander Linden, C.G. (2012) Genetic dissection of drought tolerance and recovery potential by quantitative trait locus mapping of a diploid potato population. Mol. Breed. 30, 1413–1429.

Anjun, S.A., Xie, X., Wang, L., Saleem, M.F. and Man Lei (2011) Morphological, physiological and biochemical responses of plants to drought stress. African J. Agri. Res. 6(9), 2026-2032.

Babar, M.A., Reynolds, M.P., Van Ginkel, M., Klatt, A.R., Raun, W.R. and Stone, M.L. (2006) Spectral reflectance indices as a potential indirect selection criterion for wheat yield under irrigation. Crop Sci. 46, 578–588.

Balota, M., Payne, W.A., Evett, S.R. and Lazar, M.D. (2007) Canopy temperature depression sampling to assess grain yield and genotypic differentiation in winter wheat. Crop Sci. 47, 1518-1529.

Bilge, B., Yildirim, M., Barutcular, C. and Genc, I. (2008) Effect of canopy temperature depression on grain yield and yield components in bread and durum wheat. Notulae Botanicae Horti Agrobotanici. 36, 34-37.

Blum, A. (1996) Crop responses to drought and the interpretation of adaptation. Plant Growth Regul. 20, 135–148.

Bowman, B.C., Chen, J., Zhang, J., Wheeler, J., Wang, Y., Zhao, W. (2015) Evaluating grain yield in spring wheat with canopy spectral reflectance. Crop Sci. 55, 1881–1890.

Cabello, R., Monneveux, P., De Mendiburu, F. and Bonierbale, M. (2013) Comparison of yield based drought tolerance indices in improved varieties, genetic stocks and landraces of potato (*Solanum tuberosum* L.). Euphytica 193, 147–156.

Campos, H., Cooper, M., Habben, J.E., Edmeades, G.O. and Schussler, J.R. (2004) Improving drought tolerance in maize: a view from industry. Field Crop Res. 90, 19-34.

Chaudhari, S.K., Muhammad, A. and Noshin, I. (2017) Physiological and biochemical responses of hexaploid wheat cultivars to drought stress. Pure and Applied Bio. Quetta 6(1), 60-71.

Farooq, M., Gogoi, N., Barthakur, S., Baroowa, B., Bharadwaj, N., Alghamdi, S.S. (2017) Drought stress in grain legumes during reproduction and grain filling. J. Agron. Crop Sci. 203(2), 81-102.

Fisher, R.A. and Yates, F. (1963) Statistical tables for biological, agricultural and medicinal research. (6^{th} ed) Oliver and Boyd. Edinburgh. pp 63.

Forster, B., Ellis, R., Moir, J., Talame, V., Sanguineti, M., Tuberosa, R., Teulat, M., Ahmed, I., Mariy, S., Bahri, H., El-Ouahabi, M., Zoumarou-Wallis, N., El-Fellah, M. and Salem, M. (2004) Genotype and phenotype associations with drought tolerance in barley tested in North Africa. Annuals of Applied Bio. 144, 157-168.

Gonzalez, A., Bermejo, V. and Gimeno, B.S. (2010) Effect of different physiological traits on grain yield in barley grown under irrigated and terminal water deficit conditions. J. of Agri. Sci. 148, 319–328.

Gutierrez, M., Reynolds, P., Raun, R., Stone, L. and Klatt, R. (2010) Spectral water indexes for assessing yield in elite bread wheat genotypes under well-irrigated, waterstressed, and high-temperature conditions. Crop Sci. 50, 197–214.

Jagtap, V., Bhargava, S., Sterb, P. and Feierabend, J. (1998) Comparative effect of water, heat and light stresses on photosynthetic reactions in *Sorghum bicolor* (L.) moench. J. Exp. Bot. 49, 1715-1721.

Levitt, J. (1980) Responses of plant to environmental stress: water, radiation, salt and other stresses. Academic Press, New York, pp. 365.

Li, R., Guo, P., Michael, B. and Stefania, G. (2006) Evaluation of chlorophyll content and fluorescence parameters as indicators of drought tolerance in barley. Agri. in Sci. in China. 5, 751–757.

Nageswara Rao, R.C., Talwar, H.S. and Wright, G.C. (2001) Rapid assessment of specific leaf area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using a chlorophyll meter. J. Agron. Crop Sci. 189, 175-182.

Olivares-Villegas, J.J., Reynolds, M.P. and McDonald, G.K. (2007) Drought-adaptive attributes in the Seri/Babax

hexaploid wheat population. Funct. Plant Biol. 34, 189–203.

Oncel, I., Keles, Y. and Ustun, A.S. (2000) Interactive of temperature and heavy metal stress on the growth and some biological compounds in wheat seeding. Environmental Pollution 107, 315-320.

Penuelas, J., Isla, R., Filella, I. and Araus, J.L. (1997) Visible and near infrared reflectance assessment of salinity effects on barley. Crop Sci. 37, 198–202.

Ranjan, R., Bohra, S.P. and Jeet, A.M. (2001) Plant senescence: Physiological, Biochemical and Molecular Aspects. Agrobios. New York. pp. 248.

Reynolds, M., Dreccer, F. and Trethowan, R. (2007) Drought-adaptive traits derived from wheat wild relatives and landraces. J Exp Bot. 58, 177–186.

Richards, R.A., Rebetzke, G.J., Watt, M., Condon, A.G., Spielmeyer, W. and Dolferus, R. (2010) Breeding for improved water productivity in temperature cereals: phenotyping, quantitative trait loci, markers and the selection environments. Funct. Plant Bio. 37, 85-97.

Roohi, E., Tahmasebi-Sarvestani, Z., Sanavy, S.A.M.M. and Siosemardeh, A. (2015) Association of some photosynthetic characteristics with canopy temperature in three cereal species under soil water deficit condition. J of Agri. Sci. and Tech. 17, 1233-1244.

Sairam, R.K., Deshmukh, P.S. and Shukla, D.S. (1997) Tolerance of drought and temperature stress in relation to increased antioxidant enzyme activity in wheat. J Agron and Crop Sci. 178, 171–178.

Sanchez, D., Garcia, J. and Antolin, M. (2002) Effects of soil drought and atmospheric humidity on yield, gas exchange, and stable carbon isotope composition of barley. Photosynthetica. 40, 415-421.

Talebi, R. (2011) Evaluation of chlorophyll content and canopy temperature as indicators for drought tolerance in durum wheat (*Triticum durum* Desf.). Australian J of Basic and App Sci. 5, 1457-1462.

Tattaris, M., Reynolds, M.P. and Chapman, S.C. (2016) A direct comparison of remote sensing approaches for high-throughput phenotyping in plant breeding. Front. Plant Sci. 7, 1131-1139.

Yang, X., Chen, X., Ge, Q., Li, B., Tong, Y., Zhang, A., Li, Z., Kuang, T. and Lu, C. (2006) Tolerance of photosynthesis to photo inhibition, high temperature and drought stress in flag leaves of wheat: a comparison between a hybridization line and its parents grown under field conditions. Plant Sci. 171, 389–397.