

INTERNATIONAL JOURNAL OF SCIENCE AND NATURE

© 2004 - 2013 Society For Science and Nature(SFSN). All Rights Reserved

www.scienceandnature.org

EFFECT OF CARBON AND NITROGEN ON THE GROWTH OF LIGNICOLOUS FUNGI FROM RATHANMAHAL WILDLIFE SANCTUARY, GUJARAT, INDIA

¹Praveen Kumar Nagadesi & ²Arun Arya

¹Department of Botany, P.G. section, Andhra Loyola College, Vijayawada -520008, Andhra Pradesh, India. ²Department of Botany, Faculty of Science, The Maharaja Sayajirao University of Baroda, Vadodara – 390002, Gujarat, India. Corresponding Author: Email: nagadesipraveenkumar@yahoo.com

ABSTRACT

The lignicolous fungi like *Lenzites sterioides, Tremates pini, Hexagonia apiaria* and *Navisporus floccosa* were studied for the effect of carbon and nitrogen on the growth. The effect of different carbon nitrogen sours on the growth of *N. floccosa* was studied for the first time. *H. apiaria* showed the maximum growth of mycelium in D – arabinose and D-xylose. *L. sterioides* showed maximum growth in D-xylose, sucrose, and malt extract, in *T. pini* maximum growth of mycelium was observed in case of teak wood sawdust supplemented in the medium as carbon source. All the four test fungi faild to consume D-xylose completely up to 15 days of incubation. Sucrose was slowly breakdown into monosaccharides, it remained present up to 10 days in case of *T. pini* and *N. floccosa* while it was present up to 8 days in *L. sterioides* and 12 days in *H. apiaria*. Maltose was utilized by the present fungi through a hydrolytic pathway. The effect of five different nitrogen sources was observed in case of four wood rotting fungi. The results indicate that potassium nitrite showed better growth for *T. pini* and *N. floccosa*, the sodium nitrate showed better growth in *L. sterioides*, and the ammonium nitrate as sole nitrogen source produced better growth in *H. apiaria*. Based upon growth supporting ability the inorganic nitrogen compounds are grouped as calcium nitrate >Sodium nitrate >Potassium nitrate > Potassium nitrite > ammonium nitrate (35%) for *L. sterioides*.

KEY WORDS: Carbon, Nitrogen, Growth, Lignicolous fungi, Navisporus floccose

INTRODUCTION

On land, lignicolous fungi are important decomposers and major recyclers of nutrients. In forest ecosystems the decomposition of forest litter is essential to nutrient recycling. So lignicolous fungi are able to decompose material like wood in forest and reduce it to a soft almost paper-like substance. Lignicolous fungi able to use wood as a carbon source form a relatively small and specialized group that may be subdivided into brown-rot, white-rot or soft-rot organisms according to the type of decay they cause. Brown-rot fungi degrade only the polysaccharide fraction of wood, whereas the white-rot and soft-rot organisms utilize both the lignin and the polysaccharide fractions (King 1966). Wood-decaying basidiomycetes are able to develop very large colonies with wood, a carbon-rich, but nitrogen poor material, as their sole source of nutrients. Their growth presumably requires sensitive control of their nitrogen economy, involving regulation of proteinase activity both for the extracellular digestion of the protein in wood, and for the intracellular turnover and spatial reallocation of nitrogen from mycelial protein (Wadekar et al., 1995). Very few studies have been made of the relative rates of removal of the structural components of wood during decay by white rot fungi, and brown rot fungi (Kirk, 1973) the quantitative determination of changes in the individual types of structural

sugar polymers (glucan, mannan and xylan) during the decay of conifer woods by either white rot or brown rot fungi have not been made. For heart woods such detailed analyses seem to have been done only by Cowling (1961). The quantitative changes in lignin, glucan, mannan and xylan during decay of five conifer woods by three white rot and three brown rot fungi were determined by Kirk and Highly (1973). Studies on the relative rates of utilization of the structural components of wood by white and brown rot fungi are quite frequent (Santra and Nandi, 1981). Cowling (1961) has reported preferential removal of mannan by brown rot fungi from hardwoods, while neither the major hemicellulose xylan nor the mannan is consistently removed before the glucan by the white rot fungi. Seifert (1968) has reported depletion of cellulose and xylan from pine almost simultaneously by *Cpniophora cerebella* a brown rot fungus. Nitrogen nutrition is a critical factor in the growth of wooddecay fungi. Growth on woody substrates makes special demands on nitrogen metabolism, because wood is a poor source of nitrogen. Wood contains only 0.03% to 0.1% nitrogen, mostly in organic forms (Laidlaw and Smith, 1965). The carbon: nitrogen ratio of wood can easily reach 1250:1 (Merrill and Cowling, 1966). Wood-decay basidiomycetes have evolved extremely efficient mechanisms for assimilating the nitrogen available in wood

and soil and then recycling the nitrogen by autolysis into new, actively growing portions of mycelium (Levi et al., 1968). Nitrogen derived from wood is accumulated in the mycelium to levels well above those in surrounding wood, for example Serpzlla lacymans mycelium utilizing wood as a sole nutrient source contains 3.7% nitrogen compared with 0.07 % in the wood substrate (Watkinson et al., 1981). Fermor and Wood (1981) showed that a number of fungi could degrade the walls of heat killed Bacillus subtilis cells and use these as a sole of carbon for growth. They suggest that microbial nitrogen might be particularly valuable to fungi such as wood decay basidiomycetes, which grow in low nitrogen substrates. Grant et al., (1986) added Phanerochaete Chrysosporium and three unidentified basidiomycetes to the list of decay fungi degrading heat killed Bacillus cell walls. In the present paper the effect of different carbon and nitrogen sources on growth of L. sterioides, T. pini, H. apiaria and N. floccose was studied.

MATERIALS & METHODS

Effect of carbon sources on timber degrading fungi

For the study of the effect of carbon the amount of individual substance in the basal medium was calculated, and a quantity equivalent to that was singly substituted in the basal medium by replacing the original corresponding substance *viz*, Sucrose. The amount of polysaccharides was similar to the amount of sucrose present in the basal medium. The medium devoid of sucrose served as control for carbon.

To study the effect of carbon sources on growth of timber degrading fungi, the Xylose, Arabinose, Maltose sugars were used. These sugars will acts as carbon sources which are supplemented in Czapak Dox medium. Wood degrading fungi like *L. sterioides, T. pini, H. apiaria* and *N. floccosa* were grown on basal medium containing 1% Malt extract as carbon source. After inoculation with test fungi the flasks were incubated in dark for 21 days. After completion of incubation period the fungi were filtered with Whatman filter paper No.1 and dried for 48 h at 60 °C in oven. The dried filter papers were weighed to calculate the growth of each test fungi. The filtrate was used to determine the final pH.

Effect of Nitrogen on growth of wood decay fungi

It was used for the growth of test fungi *i.e. L. sterioides, T. pini, H. apiaria* and *N. floccosa.* The basal medium supplemented with four nitrogen sources was used for growth of these fungi. Flasks containing 25 ml of basal medium were autoclaved at 121°C temperature for 20 min, inoculated with test fungi and incubated for 5, 10, and 15days. After completion of incubation period, each test fungus was filtered by using Watman filter paper no 1. The filtrate was used to determine final pH. The filter papers were dried in oven and weighed to calculate the growth of wood decay fungi.

Utilization of the sugars by wood decay fungi

Utilization of different mono-oligo, and poly sachharides as well as the hydrolytic products of di- and tri sachharides was studies. Paper chromatography was used for this purpose. The quantity of various sugars was similar to that used in experiment dealing with carbon requirements. Dry weight of mycelial mat and pH of the medium was recorded after incubation period of 5, 10 and 15 days and filtrates were analyzed daily to detect the presence of various sugars. Drops of known volume (0.05 ml) were taken from the filtrates every day and were placed on the chromatogram by micropipette at a position located for this purpose. The running solvent was n-butonol-acetic acid- water (4:1:5) v/v). In order to separate glucose and galactose the running solvent was n-butonol-pyridine-water (6:4:3 v/v). a mixture of 5 vol of 4% aniline, 5 vol of 4% diphenylamine and 1 vol of orthophosphoric acid (Buchan and Savage, 1952) was used as spraying reagent for the detection of sugars. Chromatograms were developed after drying at room temperature by heating in an electric oven at 100°C for 90 sec. the Rf values were calculated by the following formula:

Rf = <u>Distance traveled by the solute</u> Distance traveled by the solvent

RESULT & DISCUSSION

Effect of different carbon sources

Frie and Mcloughlin (2000) reported that mycelial growth of Agraicus bisporus was enhanced by malt extract a key component in PMP medium. Among the different carbon sources, mannitol and sorbitol stimulated the best mycelial growth of 110.15 and 100.45 mg/30 cm3, respectively in S. commune isolates (Adejove et al. 2007). While studying S. commune found glucose as best source followed by fructose, xvlose and mannose. Sugar alcohols like manitol also produced good growth of S. commune (Adejoye et al., 2007). Sugar alcohols and polysaccharides get hydrolyzed to monosaccharide before they will enter into respective pathways (Mahier and Cordes 1971). Bealing (1953) observed that invertase preparations catalyzed the transfer of fructofuranosyl groups not only to water, but also to various alcohols and sugars by transglucosidation Glucose and manintol have been reported as good substrates for vegetative growth (Hammond, 1978). Scientists have tried to study the decomposition of Sphagnum fuscum plants and spruce wood chips in vitro. It was found that most taxa degraded cellulose and starch via the synthesis of cellulases and amylase, respectively (Thormann et al., 2002). In order to study the effect of fungi on different wood block, it was thought desirable to use teak sawdust as one of the carbon substance.

In the medium containing D – arabinose as carbon source the wood rotting basidiomycetes member *H. apiaria* showed the maximum growth of mycelium, whereas, least growth was observed in case of *L. sterioides* and *T. pini* (Table 1). In the medium containing D-xylose as carbon sources the maximum growth was shown by *H. apiaria* and *L. sterioides*, whereas, lowest mycelial growth was shown by *T. pini*. In the medium containing sucrose as the carbon source the maximum growth of mycelium was observed in case of *L. sterioides* whereas, lowest growth of mycelium was observed in *N. floccosa*. The final pH of the medium varied from 3.31 to 5.93. In the medium containing malt extract as carbon source the maximum growth of mycelium was observed in *L. sterioides* whereas, lowest growth was shown by *N. floccosa*. In the medium containing teak wood

sawdust as carbon source the maximum growth of mycelium was observed in case of *T. pini* whereas, the lowest growth of mycelium was shown by *N. floccosa*. The final pH of the medium varied from 2.54 to 5.24.

TABLE 1: Effect of Different	Carbon source on my	celial dry weig	ght and change in	pH of four wood decay fungi
------------------------------	---------------------	-----------------	-------------------	-----------------------------

		Lenzites s	terioides	Trametes p	oini	Hexagonia	apiaria	Navisport	us floccosa
Treatment	Carbon source	Dry wt*	Final	Dry wt*	Final	Dry wt*	Final	Dry wt*	Final pH
No			pН		pН		pН		
1	D- Arabinose	56±2.8	3.80	56±1.8	3.96	78±1.8	3.98	58±1.4	4.43
2	D- xylose	101 ± 2.5	3.80	49±1.5	3.98	110 ± 1.5	3.59	69±2.6	4.80
3	Sucrose	614±1.8	5.93	306±1.2	3.31	270±2.6	4.98	220±2.8	5.45
4	Maltose	321±1.5	4.26	243±1.8	2.92	250±2.8	5.52	198 ± 3.4	5.85
5	Malt extract	456±1.0	6.24	356±1.4	2.13	289±2.4	4.63	276±3.8	5.34
6	Teak sawdust	325±2.8	4.67	423±2.5	2.54	256±1.7	4.21	253±2.3	5.24

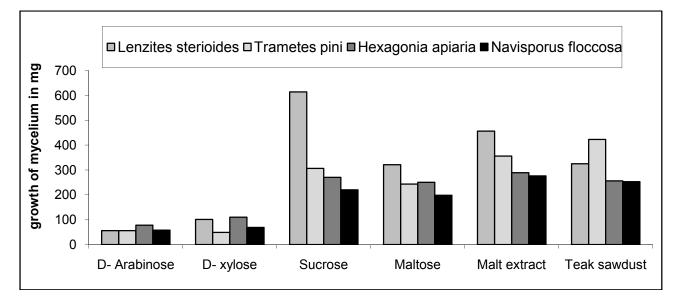
* indicates each component values are based on the three replicates.

 \pm Results were significant at P < .05 level by one way ANOVA.

The maximum mycelial dry weight of *L. sterioides* was obtained (614 mg) when sucrose was used as carbon sources, whereas, the lowest growth was 56 mg when D - arabinose was used as sole carbon source. The maximum dry weight of *T. pini* was 423 mg when teak sawdust was used as carbon sources whereas D-xylose produced lowest dry weight. The maximum growth of *H. apiaria* was obtained on malt extract, Malt extract as a complex source was found suitable for 2 strains of *Stereum hirsutum* (Jonathan *et al.* 2009) and the lowest on D-arabinose. The maximum growth of *N. floccosa* was obtained on the malt extract as a carbon source while it was lowest on the D-arabinose.

The mycelial mass increased till 60^{th} day after incubation and then it declined. The reason might be in deficiency of nutrients after two months of fungal feeding and consequently appearance of phenomena of autolytic degradation of hyphae. The similar process may happen in the wood as well. Swift (1978) found that mycelial mass increased till the weight loss of inoculated wood of some 40% but declines in later stages of decay, which was realized by using Hexosamine test methods. In the present study also after reaching to the maximum mycelial growth of wood rotting fungi, the Hyphae undergoing autolytic degradation. In the present study the best mycelial growth in *L. strioides*, *T. pini*, *H. apiaria* and *N. floccosa* were observed in case of sucrose, teak sawdust, and malt extract were used as carbon source in the medium.

In the present study the different carbon sources were used to see the maximum mycelial growth of the wood rotting fungi. As maximum growth of the mycelium is observed in all wood rotting fungi, they have the capacity to secret lignocellulolytic enzymes. The best growth of *H. apiarai* and *N. floccosa* was obtained on malt extract while it was maximum for *L. sterioides* and *T. pini* on sucrose and teak sawdust respectively (Histogram 1).



HISTOGRAM 1: Effect of different carbon sources on growth of four fungi

		Lenzites sterioides	terioides			Trametes pini	pini			Hexagon	Hexagonia apiaria			Navispo	Navisporus floccosa	P	
Mono- saccharides	Days	Dry wt [±] (mg)	Rate of growth	Final pH	Presence (days)	Dry wt [±] (mg)	Rate of growth	Final pH	Presence (days)	Dry wt* (mg)	Rate of growth	Final pH	Presence (days)	Dry* wt (mg)	Rate of growth		Final pH
D-arabinose	5	37±1.3	37	3.50		22±1.8	22	4.00		28±2.6	28	4.21		25±1.0	25		4.50
	10	48±1.8	н	3.82		35±1.5	13	4.03		54±2.8	26	4.24		42±1.5	17		4.45
	15	56±1.5	12	3.80	ц	56±1.2	21	3.96	ы	78±3.2	24	3.98	IJ	58±2.5	16	A	4.43
D-xylose	S	38±2.5	38	3.50		29±2.3	29	3.50		46±3.5	46	3.85		30±2.3 30	30	4	4.20
	10	45±2.9	13	3.84		38±2.5	9	3.95		67±2.8	21	3.76		51±2.6 21	21	4	4.39
	IJ	101±1.0	56	3.80	ы	49±1.3	11	3.98	12	110±1.5	43	3.59	IJ	69±2.8	18	4	4.80

Effect of Carbon and Nitrogen on the growth of Lignicolous fungi

Utilization of various sugars by four wood degrading fungi

Utilization of monosaccharides

Research findings have revealed that disaccharides get hydrolyzed to maonosaccharides before they enter into different pathways. The range of carbon source utilized for mycelial growth of different fungi is very wide. Monosaccharides, disaccharides and polysaccharides can be used as suitable carbon sources. Monosacharides (such as fructose, glucose etc.) or maltose among the disaccharide are the most suitable carbon sources for A. auricula (Luna et al 2004). Monosaccharides play an important role in the carbohydrate metabolism of fungi. Complex carbohydrates usually first split-up into monosaccharide units or their derivatives, which subsequently enter into the various metabolic pathways. Apart from occurring freely in various parts of the plants, these sugars are also present as component units of polysaccharides. oligosaccharides and different Monosaccharide also takes part in the synthesis of reserve carbohydrates of fungal mycelium and as such, various fungal polysaccharides are there which consist of monosaccharide units like glucose, mannose and galactose etc. In the present study daily chromatographic analysis was undertaken to determine the presence of various sugars in the culture medium. The results of mycelial growth, drift in pH and utilization of monosaccharides by different wood rotting fungi under study have been summarized in Tabel 2.

D-Xylose (Rf 0.62)

This is aldopentose occurs in nature in the form of xylans and as a constituent of polysaccharides of cell wall i.e., in hemicellulose and plant gums. It is evident from the table 2 that D- xylose was present in the culture filtrates of all the four test fungi up to 15 days of incubation. The growth rate increased in first 5 days and later on decreased. The fungal growth rate was increased up to first 5 days and later on decreased up to 10 days however a slight increase was observed up to 15 days in *L. sterioides* and *H. apiaria*. The final pH of the medium was acidic in *L. sterioides*, *T. pini* and *H. apiaria* and *N. floccosa* respectively.

D – arabinose (Rf 0.70)

This aldopentose occurs in nature in the form of arabans as common constituents of plant polysaccharides and various gums especially gum arabica. Chromatographic analysis of the medium showed that none of the wood rotting fungi under the study could assimilate this sugar completely within 15 days of incubation. The growth rate was increased in first 5 days in *L. sterioides* and *N. floccosa*, and decreasing later days of incubation. The growth rate is constantly going in *H. apiaria*. The growth rate was increased in first 5 days, later decreased up to 10 days and then increase up to 15 days in *T. pini*. The growth of mycelium was increasing as the incubation period is increasing. The final pH of the medium is acidic in *L. sterioides*, *T. pini* and *H. apiaria* and slightly acidic in *N. floccosa* respectively.

Both isomers of arabinose were employed in this nutritional investigation of *Calvatia* species but neither of them promoted satisfactory growth. According to Lilly and Barnett (1956), from the distribution of the isomers of arabinose in organisms, it would be expected that Larabinose would be utilized readily by more fungi than Darabinose. In the case of Calvatia species, two of the strains, 1019 B and 766, grew better on L-arabinose, and two strains, 1018 F and 1020, were superior on D arabinose (Sedlmayr et al., 1961). In the present study the D - arbinose showed 56mg, 78mg and 58mg of mycellial growth for L. sterioides, and T. pini, H apiaraia, and N. floccosa respectively. A study has been made of the carbon nutrition of Coriolopsis occidentalis using carbohydrates in liquid growth medium. Of the simple carbohydrates tested, the oligosaccharides supported growth best followed by xylose. (Fawole, 1973). In the present study the oligosaccharides showed better growth than the monosaccharides

Utilization of oligosaccharides

The oligosaccharides are complex sugars composed of two or more monosacharides units linked together by glycosidic bonds. They occur freely in nature or as the units of polysaccharides. These water soluble compounds yield monosaccharide components on hydrolysis.

The present studies were conducted in order to ascertain the pathway of utilization of different oligosaccharides and probable effect of their hydrolytic products on the growth of the wood rotting fungi under study. The rate if assimilation of the component sugars by the four organisms was detected chromatographically. The details of the results have been summarizes in table 3.

	Average d	ry wt. (mg)*		Final p	θH		Presence of	f sugars (days	5)
organism	5	10	15	5	10	15	Sucrose / Maltose	Glucose "	Fructose
Lenzites sterioides	170±1.8	197±2.8	614±1.8	6.15	6.36	5.93	8	1-5	1-10
Trametes pini	130±1.0	162 ± 2.2	306±1.0	4.15	3.50	3.31	10	2-8	2-11
Hexagonia apiaria	148±2.5	185±1.8	270±1.5	5.35	5.10	4.98	12	1-8	1-14
Navisporus floccosa	70±3.5	100±3.6	220±1.2	6.13	5.93	5.45	10	1-8	1-12
Lenzites sterioides	150±3.0	187±2.7	321±2.5	4.30	4.70	4.26	6	6 -12	
Trametes pini	120±3.8	159±1.6	243±2.8	3.58	3.13	2.92	15	4 -15	
Hexagonia apiaria	138±2.4	174±2.5	250±1.7	5.80	5.74	5.52	10	6 -10	
Navisporus floccosa	112±2.8	145±2.8	198±2.5	6.25	6.14	5.85	15	6-15	

TABLE 3: Utilization of Sucrose and Maltose sugars by different wood decay fungi

* indicates each component values are based on the three replicates.

 \pm Results were significant at *P* <.05 level by one way ANOVA.

Disaccharides

Sucrose (Rf 0.43)

This disaccharide is of common occurrence in plants. A large number of workers have shown that most of the fungi are able to hydrolyse sucrose into glucose and fructose and thus it is assimilated through a hydrolytic pathway. Table 3 indicates that all the wood rotting fungi under the study utilized sucrose after hydrolysis, which indicates that they were capable of producing sucrase or trans -fructosidase enzyme in sufficient amount. It yields maximum mycelial yield of wood rotting fungi i.e., L. sterioides, T. pini, H. apiaria and N. floccosa after 15 d. As the incubation period is increased the growth of the wood rotting fungi also increased. It is evident from table was slowly 3 that sucrose breakdown into monosaccharides, it remained present up to 10 days in case of T. pini and N. floccosa while it was present up to 8 days in L. sterioides and 12 days in H. apiaria. This shows that hexagonia was able to utilize this disaccharide with much slower rate.

Maltose (Rf 0.40)

It does not usually occur in the free form in chlorophyllus plants but this disaccharide is obtained as an intermediate product during the digestion of starch to glucose. It consists of two glucose units which are held together by $\alpha - 1$, 4 glucoside linkage. Maltose is utilized by a majority of fungi through a hydrolytic pathway. It yields two molecules of glucose when hydolysis is accomplished by the enzyme α – glucosidase.

Maltose was utilized by the present fungi through a hydrolytic pathway. Its presence was detected upto 6, 15, 10,15 days respectively in L. sterioides, T. pini, H. apiaria, and N. floccosa. Its hydrolytic products were detected in wood rotting fungi i.e., L. sterioides, T. pini, H. apiaria, and N. floccosa. Between 6 and 12, 4 and 15, 6 and 10, and 6 and 15 days respectively. The dry weight of all the wood rotting organisms was maximum on this sugar after15th days. The better yield may be due to hydrolysis of maltose by α – glucosidase enzyme which yields two glucose units. The glucose units were used efficiently by all the wood rotting organisms. The better growth of mycelium is also due to the slow and steady growth of all wood rotting fungi. The final pH of the medium was slightly acidic in case of L. sterioides. The pH of the medium is acidic in case of T. pini. The final pH of the medium was slightly acidic in case of H. apiaria. The pH of the medium shifted towards neutral side in case of N. floccosa.

Out of 21 carbon sources, *Lobivia laterititia* strains exhibited maximum mycelial growth on maltose followed by raffinose, starch and lactose with variable preference of different strains (Jana and Purkayastha 1987). Swartz (1933) was the only one who reported on the carbon requirement of *Calvatia* species. He found maltose the best sugar for producing mycelium of *Calvatia saccata, C. caelata* and *C. gigantean.* In the present study the maltose showed good growth of mycelium in wood rotting fungi i.e., *L. sterioides, T. pini, H apiaraia, and N. floccosa* respectively.

The mycelial growth of *Cystoderma amianthinum* was checked in the media supplemented with 11 different carbon sources. Fructose was found best screened as

carbon source for the mycelial growth of C. amianthinum (Shim et al. 2005). But in the present study It was completely utilized in 13 days. Shim et al. (1997) reported that glucose, one of monosaccharides was exceedingly good for promoting a mycelial growth of Grifola umbellata. However, it was observed that glucose was unsuitable for promoting the mycelial growth of C. amianthinum (Shim et al. 2005) submerged culture of Nigerial mushroom Pleurotus florida grow well on glucose containing medium (Gbolagade et al. 2006). In the present study the wood rotting fungi under study showed fair growth of mycelium when maltose was broken down to two glucose molecules, it was utilized with in 4 to15 days of incubation. Chi et al. (1996) reported that though each of some monosaccharides was supplemented in the basal medium to check a mycelial growth of Phellinus linteus, its mycelial growth was dissimilar among monosaccharides. In the present study also the growth of mycelium depend on the nature of the wood rotting fungi under study.

Sucrose was not a good carbon source for *Calvatia* species. It could be that the organisms produced the hydrolyzing enzyme very slowly or in a small quantity, as both components of this disaccharide (glucose and fructose) produced a satisfactory growth when used separately. Apparently the glucose to fructose linkage was not easily broken (SedImayr *et al.*, 1961). In the present study the sucrose yielded the glucose and fructose which were utilized in 8 to 10 days by wood rotting fungi under study.

Effect of different nitrogen sources

This essential element is used by fungi for functional as well as structural purposes. Chitin, the chief component of cell wall in most of the fungi, is a linear polymer of Dglucoseamine. Similarly proteins, the basis of protoplasm are composed of nitrogenous substance. Purines, pyrimidines, some vitamins and other essential metabolites are also nitrogen containing compounds. In nature both the organic and inorganic forms of nitrogen are available to fungi but as far as their utilization is concerned they fundamentally differ from each other in their metabolic potentialities. A few utilize atmospheric nitrogen, many utilize nitrate nitrogen and a still greater number utilize ammonium nitrogen. All species are able to utilize some form of organic nitrogen. Owing to the specific response of various fungi towards different nitrogenous substances numerous investigators have classified them into different groups on the basis of their abilities to utilize these sources.

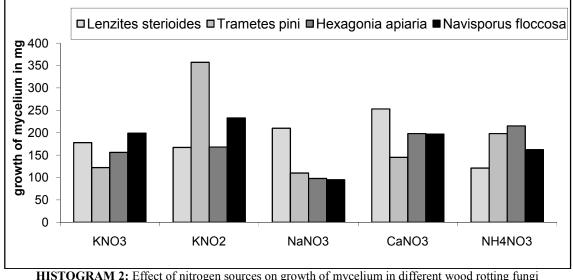
For the few wood rot fungi previously studied both qualitative and quantitative difference have been found in the utilization of known organic and inorganic nitrogen compounds including certain amino acids (Hacskaylo *et al.* 1954 and Yusef 1953). Generally ammonium nitrogen is assimilated (Fries 1950, Lilly and Barnett 1951, Yusef 1953). A few wood rot fungi utilize nitrate nitrogen slowly in stationary culture (Lilly and Barnett 1951, Hacskaylo *et al.*, 1954). Discussing the possible pathway of protein synthesis Lilly and Barnett (1951) have mentioned, "with the exception of certain amino acids (primary amino acids) and ammonia, most nitrogen sources undergo modifications before entering the synthetic metabolic

pathways. Nitrates, nitrites and hydroxylamine are presumably reduced to ammonia before assimilation. Those amino acids (secondary amino acids) which do not enter directly into the metabolic pathways leading to the synthesis of protein are probably deaminated."

In urea there was substantial increase in the average amount of growth from the third to ninth serial subculture, with both the brown rot and the white rots. Presumably this increase reflected some degree of adaptation of certain organisms to the nutrients. M. americanus showed a marked increase in growth in casein hydrolysate and in ammonium sulfate and F. fomentarius in ammonium carbonate from the third to the ninth serial transfer. The negative data for growth in ammonium chloride, Potassium nitrate and Potassium nitrite were not included, since T. serialis was the only culture which could utilize

even one of the compounds under the conditions used (Jennison et al., 1955).

The effect of 5 different nitrogen sources was observed in case of 4 wood rotting fungi, the results are depicted in Table 4. The potassium nitrite showed better growth for T. pini and N. floccosa. The final pH of the medium changed from acidic to slightly neutral nature. The sodium nitrate showed better growth in L. sterioides and lowest growth was shown by N. floccosa. The calcium nitrate as sole nitrogen source showed better growth in L. sterioides, as compared to other fungi. The ammonium nitrate as sole nitrogen source showed better growth in H. apiaria and lowest growth in L. sterioides respectively (Histogram 2). Based upon growth supporting ability the inorganic nitrogen compounds are grouped as calcium nitrate > Sodium nitrate > Potassium nitrate > Potassium nitrite > ammonium nitrate for L. sterioides.



HISTOGRAM 2: Effect of nitrogen sources on grow	wth of mycelium in different wood rott	ing fungi
---	--	-----------

		Lenzites s	Lenzites sterioides		pini	Hexagonia	a apiaria	Navispor	us floccosa
Treatment	Nitrogen	Dry wt*	Final	Dry wt*	Final	Dry wt*	Final	Dry wt*	Final
No	source	-	pН	-	pН	-	pН	-	pН
1	KNO3	178±2.8	4.53	122±2.2	4.65	156±1.8	5.70	199±1.5	5.54
2	KNO2	167±3.4	6.30	357±1.0	4.70	168±1.2	6.76	233±2.5	7.12
3	NaNO3	210±1.5	7.10	110±1.5	6.86	98±1.8	7.26	95±2.8	7.60
4	CaNO3	253±2.5	6.20	145±2.5	5.10	198±1.9	5.23	197±3.4	6.18
5	NH4NO3	121±2.6	3.58	198±1.7	4.50	215±2.4	6.50	162±3.9	5.18

TABLE 4: Effect of different Nitrogen sources on growth of wood decay fungi

*indicates each component values are based on the three replicates.

 \pm Results were significant at P <.05 level by one way ANOVA

The brown rot fungi as a group and for the white rot species, there was a consistent decrease in average growthsupporting ability for ammonium nitrate (Jennison et al., 1955). In the present study the wood rotting fungi showed the good mycelial growth in *H. apiaria* and lowest growth in L. sterioides for ammonium nitrate as sole nitrogen sources. The Basidiomyctes with the exception Polyporus distortus, grew very slowly on nitrate nitrogen. The species of basidiomycetes tested utilize nitrate nitrogen slowly or not at all, with the time of incubation (Hacskaylo et al., 1954). In the present study the wood rotting fungi

showed good growth in nitrate nitrogen. The phenomenon of slow utilization of nitrate nitrogen appears to an exaggerated degree in some of the Basidiomycetes tested (Hacskaylo et al., 1954). In the present study the nitrate nitrogen shown better growth in 15 days of incubation as it take less time for stabilization

and the wood rotting fungi *i.e.*, L. sterioides, T. pini, H. apiaria and N. floccosa showed good growth of mycelial mass in nitrate, nitrite, ammonium nitrogen sources which were incubated for short time. On the other hand none of these strains could grow on Sodium nitrite. All strains

responded moderately to the ammonium compounds studied. However the nitrates were found to support moderate to poor for growth of all strains (Singh and Verma, 1996). In the present study the sodium nitrite showed better growth in T. pini and N. floccosa and good growth in L. sterioides and also in H. apiaria.. Similarly sodium nitrite has been shown as a non available nitrogen source to several fungi including Agaricus bisporus (Hsu and Hu 1967) and Lentinus edodes (Tokimoto and Kumatsu 1979). In the present study L. sterioides, T. pini, H. apiaria and N. floccosa showed better to good growth of mycilial mass when sodium nitrite was used as sole nitrogen source. In the series of complex and inorganic nitrogen sources, it was observed that inorganic compounds supported moderate biomass production. The best biomass yield was found with ammonium nitrate closely followed by potassium nitrate. This result is contrary to that obtained by (Jonathan and Fasidi, 2001) for P. atroumbonata where ammonium nitrate supported insignificant mycelial yield. (Gbolagade et al., 2006). In the present study also the ammonium nitrate showed good growth for *H. apiaria* and *T. pini* to moderate growth in *N*. floccosa and L. sterioides. So ammonium nitrate is good source of nitrogen. As the incubation period increases the growth mycelium also increased in Sterium hirsutum on Ammonium sulphate and Potassium nitrate as sole nitrogen source (Minc 2005). The potassium nitrate also showed good growth in N. floccosa and L. sterioides and moderate growth in T. pini and H. apiaria. The differences in nitrogen sources requirements may suggest that biomass production in different fungi may be influenced by different nutritional requirements. It was also observed that very low concentration of nitrogen compounds generally supported little biomass yield while low concentration and above were supportive to high biomass yield (Yajie and Zhong, 2002). In the present study also the wood rotting fungi showed good growth in higher concentration of nitrogen as the free nitrogen was available to the fungi. The mycelial growth of Cystoderma amianthinum in Calcium nitrate was 5.6, Potassium nitrate was 16.4, and Sodium nitrate was 30.6 mg (Shim et al., 2005). In the present study the mycelial mass of the L. sterioides in calcium nitrate was 25 3mg, potassium nitrate was 178 mg and sodium nitrate was 210mg. Nitrogen sources (NH₄)₂HPO₄, NH₄Cl, NH₄NO₃ (NH₄)₂SO₄ were good source of ammonium nitrogen sources whereas the KNO₃ and KNO₂ are poor source of nitrate nitrogen (Niederipruem, et al., 1964). In the present study the ammonium nitrogen sources showed good to moderate growth whereas the inorganic nitrogen sources *i.e.* KNO₃ and KNO₂ showed better to good growth in all wood rotting fungi under study. These fungi, Polyporus adustrzs and Liberfella befulincr, grew better on ammonium nitrate. Utilization of nitrate led to an increase of pH in the medium. The changes in pH of media in which Polyporrrs adusfus had grown on ammonium nitrate indicates that the ammonium ion was taken up before the nitrate ion (Henningsson, 1967). In the present study also the pH of the medium is decreasing as incubation period is increasing. This decrease may be due to the release of cations and organic acids into the medium.

ACKNOWLEDGEMENTS

The authors are thankful to the Head, Department of Botany, The M. S. University of Baroda for laboratory facilities. This study is funded by DST, New Delhi.

REFERENCES

Adejoye D., Adebayo-Tayo B. C., Ogunjobi A. A., and Afolabi O. O. (2007) Physicochemical studies on *Schizophyllum commune* (Fries) a Nigerian Edible Fungus. World Applied Sciences Journal 2 (1), 73-76.

Bealing, F. J. (1953) Mould "Glucosaccharase": a fructosidase. Biochem. J. (London) 55, 93

Chi, J. H., Ha, T. M., Kim, Y. H. and Rho, Y. D. (1996) Studies on the main factors affecting the mycelial growth of *Phellinus linteus*. Kor. J. Mycol. 24(3), 214-222.

Cowling E.B. (1961). Comparative biochemistry of the decay of sweetgum by white rot and brown rot fungi. U.S. Dep. Agric. For. Serv.Tech. Bull. 1259. Washington D.C. 79pp.

Fawole, M. O. (1973) Studies on the carbon nutrition of *Coriolopsis occidentalis* (Klotzsch) Murr. Zeitschrift für allgemeine Mikrobiologie 13, 395–403.

Fermor, T.R. and Wood, D.A. (1981) Degradation of bacteria By *Agaricus Bisporus* and other fungi. J. Gen. Micobiol. 126, 377-387.

Fries, N. (1950) Growth factor requirements of some higher fungi. Svensk bot. Tid. 44, 379-386.

Gbolagade J., Sobowale A. and Adejoye D. (2006) Optimization of sub-merged culture conditions for biomass production in *Pleurotus florida* (mont.) Singer, a Nigerian edible fungus. African Journal of Biotechnology 5 (16), 1464-1469.

Grant W.D., Rhodes L.R., Prosser B.A. and Asher R.A. (1986) Production of baceriolytic enzymes and degradation of bacteria by filamentous fungi. J. Gen. Micobiol. 132, 2253-2258.

Hacskaylo J., Lilly L.G., and Barnett H.L. (1954) Growth of fungi on three sources of nitrogen. Mycologia 46, 691-701.

Hammond, W.B.J. (1978). Changes in composition of harvested mushrooms (*Agaricus bisporus*). Phytochemistry 18, 415-418.

Henningsson B. (1967) Physiology of fungi attacking birch and aspen pulpwood. submitted to Srogshogskolan, Royal College Of Forestry Stockholm Printed in Sweden by Esselte Ab, pp 54

Hsu, H.T. and Hu K.J. (1967) Nitrogen nutrition of *Agaicus* bisporus.I form of inorganic nitrogen utilized by *Agaricus* bisporus. Nung Yen Chiu 16, 25-28.

Jana K.K. and Purkayastha R.P. (1987) Nutritional effects on the growth of two edible fungi and theor nutritive values. Indian Mush. Sci. 2, 305-308.

Jonathan S.G., and Fasidi I.O. (2001) Effect of carbon, Nitrogen and Mineral Sources on growth of *Psathyenella atroumbonata* (Pegler), a Nigerian Edible Mushroom. Food Chem. 72, 479-483.

Jonathan S.G., Bawo D.D.S., Adejoye D.O. and Briyai O.F. (2009) Studies on Biomass Production in *Auricularia polytricha* collected from Wilberforce Island, Bayelsa State, Nigeria. American Journal of Applied Sciences 6 (1), 182-186,

Kirk T.K. (1973) The chemistry and biochemistry of decay. p. 149-181.in D. Nicholas (ed.) wood deterioration and its prevention by preservative treatments. Syracuse University press. Syracuse N.Y.

Kirk T.K. and Highley T.L. (1973) Quantitative changes in structural components of conifer woods during decay by white and brown rot fungi. Phytopathology 63, 1338 -1342.

Laidlaw, R. A., and Smith G. A. (1965) The proteins of the timber of Scots pine (*Pinus sylvestris*). Holzforschung 19, 129-134.

Levi, M. P., Merrill, W. and Cowling. E. B. (1968) Role of nitrogen in wood deterioration. VI. Mycelial fractions and model nitrogen compounds as substrates for growth of *Polyporus versicolor* and other wood-destroying and wood-inhabiting fungi. Phytopathology 58, 626-634.

Lilly V. G. and Barnett H. L. (1951) Physiology of fungi. Mcgrauhill, New York 464pp

Lilly, V. G., and Barnett, H. L. (1956) The utilization of D- and L-arabinose by fungi. Am. Jour. Bot. 43, 709.

Luna M. L., Murace M. A., Keil G. D. and Otaño M. E. (2004) Patterns of decay caused by *Pycnoporus sanguineus* and *Ganoderma lucidum* (Aphyllophorales) in poplar wood. IAWA 25 (4), 425–433

Mahier, H.R. and Cordes, H.E. (1971) Biological Chemistry. 2nd Edn. New York.

Merrill, W., and Cowling E. B. (1966) Role of nitrogen in wood deterioration: amount and distribution of nitrogen in fungi. Phytopathology 56, 1083-1090.

Minc M. (2005) Decay of oak wood provoked by fungus *Atereum hirsutum* (Wild Ex Fr.) S.F. Gray. and it's essential physiological requirements. Biblid 91, 179-192

Niederipruem D. J., Hobbs H., and Henry L. (1964) Nutritional studies of development in *Schizophyllum commune*. Journal of Bacteriology 88 (6), 1721-1729

Sedlmayr M. E. S., Beneke J. A. and Stevens (1961) physiological studies on *Calvatia* Species. II. Carbon Utilization. Mycologia 53 (6), 558-565

Seifert K. (1968) Zur Systematik der holzfaulen ihre chemischen and physikalischen kennzeichen. Holz rohwerks 26, 208-215

Shim, J. O., Son, S. G., Kim, Y. H., Lee, Y. S., Lee, J. Y., Lee, T. S., Lee, S. S. and Lee, M. W. (1997) The cultural conditions affecting the mycelial growth of *Grifola umbellate*. Kor. J. Mycol. 25(3), 209-218.

Shim, S. M. Oh Y. H., Lee, K. R Kim, S. H. Im K. H., Kim, J. W. Lee U Y., Shim J. O., Shim M. J., Lee M. W., Ro H. S, Lee H. S. and Lee T. S. (2005) Culture Conditions Affecting the Optimal Mycelial Growth of *Cystoderma amianthinum*. Mycobiology 33(1), 65-67.

Shim, S. M. Oh Y. H., Lee, K. R Kim, S. H. Im K. H., Kim, J. W. Lee U Y., Shim J. O., Shim M. J., Lee M. W., Ro H. S, Lee H. S. and Lee T. S. (2005) Culture Conditions Affecting the Optimal Mycelial Growth of *Cystoderma amianthinum*. Mycobiology 33(1), 65-67.

Singh T.G. B. and Verma R.N. (1996) Studies on carbon and nitrogen nutrition of *Leetinula lateritia* (Berk.) Pegler strains from northeastern India. Mushroom biology and mushroom products. Royse (ed.) Penn State University.

Swartz, D. (1933) Some development characters of species of Lycoperdaceae. Am. Jour. Bot. 20, 440.

Swift M. J. (1978) Growth of *Stereum hirsutum* during the longterm decomposition of oak branch wood. Soil biolog and Biochemistry 104, 335-337

Thormann M. N., Currah R. S. and Bayley S. E. (2002) The relative ability of fungi from *Sphagnum fuscum* to decompose selected carbon substrates. Can. J. Microbiol. 48(3), 204–211

Tokimota K. and Komastu M. (1978) Biological nature of Lentinus edodes. In the "Biology and cultivation of edible mushrooms (eds. S.T. Chang and W.A. Hayes). Academic press. Inc. New York 445-459pp.

Wadekar R. V., North M. J. and Watkinson S. C. (1995) Proteolytic activities in two wood-decaying basidiomycete fungi, *Serpula lacrymans* and *Coriolus versicolor*. Microbiology 141, 1575-1583

Watkinson. S.C., Davison, E. M. and Bramah J. (1981) The effect of nitrogen availability on growth and cellulolysis by *Serpula lacymans*. New Phytol 89, 295-305.

Yajie T. and Zhong, J. T. (2002) Fed batch fermentation of *Ganoderma lucidum* for hyperproduction of polysaccharides and ganoderic acid. Enzyme Micr. Tech. 31, 20-28.

Yusef H.M. (1953) The requirements of some hymenomycetesfor essential metabolites. Bull. Torrey Bot. Club. 80, 43-64.