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SEASONAL VARIATIONS AND PHYSICOCHEMICAL CHARACTERISTICS OF THE HABITATS IN RELATION TO THE DENSITY OF DENGUE VECTOR AEDES AEGYPTI IN THANJAVUR, TAMIL NADU, INDIA

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ABSTRACT

Nadu, India. In each study, the breeding habitats such as coconut shells, waste buckets and tires were examined for the presence of mosquito larvae. The surveys were carried out twice in each month from January, 2013 to December, 2013 and the larval density was correlated with the habitat characteristics. The physicochemical characteristics of breeding water such as pH, conductivity, salinity, total dissolved solids, turbidity and dissolved oxygen were measured. One way analysis of variance $F(_{2,21})= 3.49$, gives p value 0.045, i.e., p.> 0.05. From this observation of the value p, it was clear that there is no significant difference between the larval densities among the three habitats. The correlation between the physicochemical characteristics and the larval density of container- breeding mosquitoes indicated that the pH, calcium and the phosphate showed positive correlation with the larval density, total hardness, chloride, and sulphate were negatively correlated with larval abundance in the containers, with correlation coefficients of -0.65, -0.65, -0.50, -0.003 and -0.66 respectively. Rainfall is an important environmental factor associated with *Aedes* breeding at the study sites.

KEY WORDS: *Aedes aegypti*, physicochemical characteristics, Rainfall

INTRODUCTION

The occurrence of dengue has grown dramatically around the world in recent decades. Some 2.5 billion people - two fifths of the world's population – all now of risk from dengue. WHO currently estimates there may be 50 million dengue infections worldwide every year (WHO 2012). In the Indian scenario, almost due to favorable ecological conditions. Aedes aegypti L., a vector of dengue that carries the arbovirus responsible for these diseases, is widely distributed in the tropical and subtropical zones. The only way to prevent dengue virus transmission is to combat the disease carrying mosquitoes. The global resurgence and expanded distribution of vector borne diseases, such as dengue fever and chikungunya infection has generated a renewed interest in the biology and control of Aedes (Stegomyia) mosquitoes in recent years. Aedes aegypti (Linnaeus) is a principal vector of these diseases in several countries. This mosquito usually breeds in natural habitats especially in tree holes, leaf axils, rock pools and similar sites. The container habitats have unique ecological properties and these habitats could be natural such as tree holes and leaf axils or artificial such as tyres, plastic cups and water tanks (Service, 1995). The household wastes such as earthen, porcelain, plastic and coconut shells were as larval habitats of the dengue vectors (Soumyajit Banerjee et al., 2012). The population of Aedes aegypti fluctuates with temperature, rainfall and humidity. Dengue infectious was generally encountered during or after rain along with the rise in the vector Aedes aegypti population (Pandya et al., 1982; Ananya Bar and Andrew, 2012). In extremes of weather in winter and summer Aedes aegypti larvae die because of low and high temperature. Widespread deforestation, climate change and increase in global trade has forced this mosquito worldwide to adapt to breeding in domestic and semi domestic artificial container habitats (Gubler et al, 2001; Delatte et al, 2008). The species composition of the container-breeding mosquitoes and their habitat characteristics need to be studied in the context of the emergence of dengue and chikungunya in this part of the country. The information on the ecological factors influencing mosquito biology such as the physicochemical properties of breeding water could help in better implementation of the vector management programmes (Rao et al., 2011; Reji Gopalakrishnan et al., 2013). Hence, the present study was carried out to understand the Seasonal variations and physicochemical characteristics of the habitats in relation to the density of dengue vector Aedes aegypti in Thanjavur, Tamil Nadu, India.

MATERIAL & METHODS

Study area

The studies on the mosquito breeding habitats were conducted in and around Thanjavur District of Tamil Nadu, India.

Larval surveys

In each study, the breeding habitats such as coconut shells, waste buckets and tires were examined for the presence of mosquito larvae. The surveys were carried out twice in each month from January, 2013 to December, 2013.

Larval density

The mosquito larvae were collected from the breeding habitats using a net (6cm. width). Incidences of Aedes

aegypti larval collections were recorded and the larval density was calculated using the following formula:

Larval density= Number of larvae collected / Number of dips made

Physicochemical characteristics

Water samples were collected from the breeding habitats, which showed the presence of mosquito larvae. The breeding water characteristics were pH, conductivity (μ S/cm), salinity (ppt), total dissolved solids (mg/l), turbidity (NTU) and dissolved oxygen (%) were recorded using Orion 5-star portable multipara meter (Thermo Scientific). The dissolved micronutrients such as, Calcium, Magnesium, Iron, Ammonia, Nitrite, Nitrate, Chloride, Fluoride, Sulphate and Phosphate was estimated by standard methods APHA (2006).

Data analysis

Means and standard deviations (SD) were used for summarizing the physicochemical factors. The mean larval

density among the readings of each month of 2013 was compared using one-way ANOVA. The relationships between the mean values of the habitat characteristics and the larval densities were derived by Pearson's correlation. The statistical analyses were carried out using IBM SPSS 19 statistical software.

RESULTS

Aedes aegypti population density in Coconut shell, waste bucket and tires in and around Thanjavur District is considerably more in the collected mosquito larval samples. Aedes aegypti larvae are found in water collections throughout the year. The larval collections made from the places during January 2013 to December 2013 (Figure 1, Table 2) show the variation in the larval density from 1.66 ± 0.57 to 18.00 ± 1.00 from the monthly collections. From January to June, the larval index is considerably low. It ranges from 1to 8. Aedes aegypti larval development is found very slow in this season.

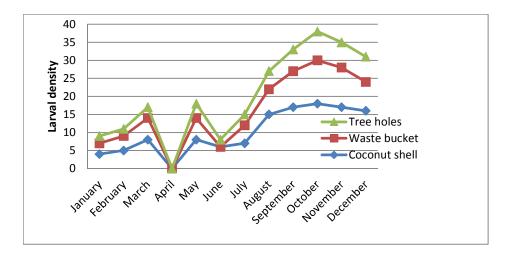


FIGURE 1. Larval density of Aedes aegypti in 2013

Physicochemical parameters	Coconut Shell	Waste Bucket	Tires
Turbidity NTU	29.66 ±1.52	20.66±2.08	25.33+1.52
2			
Total Dissolved Solids mg/l	511.33±1.52	79.83±1.75	693.00±0.5
Electrical Conductivity mho/cm	730.16±0.76	115.3±0.36	989.9±0.73
pH	6.73±0.67	6.38±0.38	6.50 ± 0.02
Total Hardness(as CaCo3)mg/l	40.23±0.68	22.06 ± 0.60	83.83±0.76
Calcium mg/l	55.93±1.10	40.26±1.02	20.16±1.25
Magnesium mg/l	9.93±0.63	3.10±0.36	21.80±0.62
Iron mg/l	1.62 ± 0.22	00	0.28 ± 0.2
Ammonia mg/l	0.13 ± 0.005	0.5 ± 0.01	0.18 ± 0.01
Nitrite mg/l	8.67 ± 0.05	0.62 ± 0.03	11.54 ± 0.20
Nitrate mg/l	50.00 ± 1.52	6.9±0.63	50.0 ± 1.00
Chloride mg/l	159.66±1.52	2.93±0.60	208.16±7.52
Fluoride mg/l	0.38 ± 0.44	0.3±0.1	0.7 ± 0.16
Sulphate mg/l	32.7±0.40	24.00±1.00	99.76±1.36
Phosphate mg/l	4.77 ± 0.07	00	4.31±0.47

TABLE 1. Physicochemical parameters of different breeding habitats

TABLE 2. T	he survey c	of Aede	es aegypti l	larvae from	January	2013 t	o Decem	ber 201	3 in	Kannan I	Nagar,	Thanjavur
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	Mean sample of mosquito larvae			Average Rainfall
Monthly	10 dipp	10 dipping / larval density (\pm)		
catches	Coconut Shell	Waste Bucket	Tires	
January	3.66 ± 1.52	3.00±1.00	1.66 ± 0.57	4
February	5.33 ± 1.52	4.66 ± 2.08	2.33 ± 1.52	6.5
March	6.66 ± 1.52	5.33 ± 2.51	3.00 ± 2.00	13.6
April	$7.00{\pm}1.00$	6.00 ± 1.00	4.33±1.52	00
May	6.00 ± 1.00	4.33±0.57	2.66 ± 0.57	7.7
June	6.66 ± 0.57	5.33±0.57	3.33 ± 1.52	24.6
July	14.66 ± 0.57	6.66±1.52	4.00 ± 1.00	5
August	16.66 ± 1.52	10.16±0.76	$5.66 \pm .52$	26.9
September	18.00 ± 1.00	11.66 ± 1.52	8.5 ± 0.50	22.9
October	17.33±0.57	11.00 ± 1.00	7.00 ± 1.00	17.71
November	16.33±0.57	7.66±0.57	7.33 ± 0.57	15.23
December	15.33±0.57	6.66 ± 1.52	5.66 ± 1.52	16.66

TABLE 3. Correlation coefficient between	Aedes aegypti larval density	y and physicochemical	l parameter of breeding				
habitats							

Parameters	Correlation coefficient	<i>p</i> -value	
pH	0.80	0.20	
Total Dissolved Solids	-0.065	0.47	
Electrical conductivity	-0.065	0.47	
Total hardness	-0.505	0.33	
Calcium	0.952	0.99	
Chloride	-0.003	0.49	
Sulphate	-0.661	0.27	
Phosphate	0.22	0.42	

The maximum larval index was noticed in 2013 in the month of August and September (16-18) in all three breeding habitats. During this time, the average rainfall was high 26.9 and 22.9 and Dengue cases were also prevalent in Thanjavur. The larval index remained high till December (15.33) and it again decreased considerably in January. In March a decline in larval index (6.00) occurred and less rainfall (3.6) also was noticed. Due to extreme summer Aedes aegypti larval improper development and mortality was noticed. The density of container breeding mosquitoes (mean \pm SE mean) was the highest in Coconut shell (18.00 ± 1.00) followed by waste bucket (11.66 ± 1.52) and Tree holes (8.5±0.50). Analysis of the physicochemical characteristics of water in the mosquito breeding sites indicated that the pH ranged from 6.38 \pm 0.38 in waste bucket to 6.73 ± 0.67 in Coconut shell whereas the conductivity (mho/cm) ranged from 115.3 \pm 0.38 in waste bucket to 989.9 ± 0.73 in Tree holes. The Total dissolved solids (mg/l) was the lowest in waste bucket (79.83 \pm 1.75) and the highest in Tree holes (693.00 ± 0.5) whereas the total hardness (mg/l) was the highest in Tree holes (83.83 ± 0.76) and the lowest in waste bucket (22.06 \pm 0.60). The turbidity of breeding water ranged from 20.66 ± 2.08 in waste bucket to $29.66 \pm$ 1.52 in Coconut shell whereas the Calcium (mg/l) ranged from 55.93 ± 1.10 in coconut shell to 20.16 ± 1.25 in Tires (Table 1). One way analysis of variance (ANOVA) F (2, 21) = 3.49, gives p value 0.045, i.e., p.> 0.05. From this observation of the value p, it was clear that there is no significant difference between the larval densities among the three habitats. The correlation between the physicochemical characteristics and the larval density of container- breeding mosquitoes indicated that the pH,

calcium and the phosphate showed positive correlation with the larval density, the correlation coefficients being 0.2, 0.99 and 0.22 respectively. Total dissolved solids, electrical conductivity, total hardness, chloride, and sulphate were negatively correlated with larval abundance in the containers, with correlation coefficients of -0.65, -0.65, -0.65, -0.003 and -0.66 respectively. (Table 3).

DISCUSSION

The selection of breeding sites by mosquitoes is a critical factor for mosquito survival and population dynamics and has important implications for mosquito control. Site may be affected by chemical and physical factors: attractants and deterrents. A deterrent causes insects to avoid that area, inhibiting egg laying. Sharma et al (2008) demonstrated the repellent effect of some fatty acid esters on oviposition of mosquitoes. Oviposition site selection is important in the life cycle of mosquitoes. Stimuli for an ovipositional flight are linked with some environmental factors, such as rainfall, relative humidity, temperature and wind speed; the selection of a breeding site involves visual, olfactory, and tactile responses (O'Gower, 1963; Bentely and Jonathan, 1989). A. aegypti exhibits a great deal of specialization in breeding site selection and consequently the distribution of this species is limited by those sites. Female mosquitoes have adapted to laying eggs in artificial containers, especially discarded tires, which resist desiccation, the eggs then hatch after being transported in used tires from Asian countries to Western countries (Medlock et al, 2006; Straetemans, 2008). It is possible species that breed in containers, both natural and artificial, may rely on some sort of visual analysis for site location and ensuing ovipositon. Ae. albopictus females

may utilize smell and contact chemoreception for choosing breeding sites and prefer water with low illumination and dark color (Gubler, 1977). In a study in Delhi (Seghal and Pillar, 1970), A. aegypti and Ae. albopictus were found to favor identical breeding sites in regard to turbidity, pH, alkalinity, chloride and phosphate levels. However, they did not prefer the same water conditions. Ae. albopictus preferred less turbid waters with moderate alkalinity whereas *Culex* and *Anopheles* mosquitoes preferred more turbid water with high alkalinity and less oxygen content. The sulphate content was lower in the Culex and Anopheles breeding sites than the Aedes breeding sites, but the chloride content was higher in the *Culex* an *Anopheles* breeding sites and lower in the Aedes breeding sites. The phosphate content was low in A. aegypti breeding sites. The authors concluded mosquitoes exhibited species specific preferences in oviposition in regard to the chemical composition of water. Various physical and chemical factors of larval habitats contribute to mosquito breeding site selection, such as temperature, pH, ammonia, nitrate, sulphate, phosphate and dissolved solids (Oyewole et al, 2009; Oleymi et al, 2010). In our study we found a correlation between factors investigated and larval prevalence. Coconut shells were chosen more often by mosquitoes for breeding sites than other containers, with more A. aegypti eggs found in these habitats. Most of the coconut shells were discarded after use or had fallen from a tree and had been gnawed open by a rodent. Coconuts are rich in calcium, potassium, sodium, sulphur and magnesium forming an ideal breeding ground for A. aegypti. They have a small orifice and are dark inside, making it an ideal mosquito breeding site. The pH of water has an impact on mosquitoes, influencing osmoregulation and oxygen transportation (Umar and Donpedro, 2008). In this study of A. aegypti breeding, an increase in pH resulted in a significant decrease in larval density. The container sites sampled had a comparable pH levels. This demonstrates that A. aegypti mosquitoes favor a specific pH range for breeding. This could be an important factor in breeding site selection and larval survival. A pH variation outside the range of 7-8 could be used as a tool for management of this vector. Manipulation of the pH of preferred breeding sites, such as coconut shells, tires and plastic containers could be used to control this mosquito where breeding site reduction is not possible due to local factors. Further studies regarding this aspect need to be carried out. Spraying biopesticides, such as neem oil with a pH >8 could be useful for this purpose. Neem oil, a natural product, is environmentally safe and may be less harmful than synthetic insecticides. Salinity is another important factor that can have a repelling or attracting effect on oviposition. Navarro et al (2003) found increasing salinity in the laboratory decreased oviposition. However, our findings showed A. aegypti was tolerant to variations in salinity, possibly leading to further geographical expansion of this mosquito's breeding sites. Previous studies found Aedes prefers to breed in less turbid waters, similar to that found in plastic containers and tires; however, the high turbidity of the coconut shell breeding sites contradicts those findings. The rich organic content of coconut shells could be a reason for this. Carbonate was not detected in any of the samples we analyzed. Our study determined that larvae were present in the Coconut shells, Waste buckets and Tree holes. This indicates that larval

density was associated with physicochemical parameters of breeding sites. During field surveys of mosquito breeding sites in the Garhwal region in the state of Uttaranchal, India, several environmental factors that could be quantified were found to be promising as predictors of mosquito occurrence (Pemola and Jauhari 2005). The present study showed that the physicochemical parameters, Nitrate, Sulphate, Salinity, Iron, Calcium, pH and temperature was affect the *A. aegypti* larval density. Various chemical properties of the larval habitat in relation to vegetation, optimum pH and temperature, concentration of ammonia, nitrate and sulphate have been reported to affect larval development and survival (Pal, 1945; Mutero *et al.*, 2004; Thangamathi *et al.*, 2011).

In India Dengue infections generally occur during or after rain along with the rise in the vector Aedes aegypti population (Pandya, 1982). In Agra, Aedes aegypti larval density is high from July (17) to October (24.5) in 2009 and from July (20.7) to October (20) in 2010. In the neighboring city Delhi, Aedes aegypti larval density was high during July to October (Katyal et al., 2003), whereas in Gorakhpur it was during June to October (Rao, 1967). In the neighboring states, Rajasthan and Madhya Pradesh the larval density was also high during July to October (Pandya, 1982). In Vellore of the state of Tamilnadu Aedes aegypti larval density was high from September to December (Rao, 1967). In coastal plains of Orissa, Rohilkhand and Avadh plains and Haryana the larval density is high in wet season whereas in Assam valley in dry season the larval population is high (Kalra et al., 1997). The rise in dengue incidence during the monsoon and the post-monsoon season is due to the increased larval density and in the increased number of potential breeding sites due to water loggings. In India, all dengue/DHF outbreaks are associated with a higher container index of more than 20 Aedes Aegypti larvae (Sharma et al., 2005). Aedes aegypti larval density in Agra reach to a peak in the month of September in 2009 and in the months of July and October in 2010. In Gurgaon the maximum larval density was observed in the month of August in 1991 and in May in 1992 (Sharma, 1997). In Delhi the maximum larval density was noticed from August and September (Sharma et al., 2005), and July to September (Pramanik et al., 2007). In Gorakhpur (Rao, 1967) the maximum larval density was observed during August, in Vellore] during September and in Kolkata during August and November.

Because of the variation in the incidence of temperature and rainfall, fluctuation in larval density happens. In the months of October in 2009 and November in 2010 the larval density again decreases. Pandya in 1982 described that one of the possible reason for the seasonal aberration could be the fluctuating breeding habitat of Aedes aegypti in different types of containers e.g. dumped and moist tyres or earthen-wares used for storing water during summers (Ananya Bar and Andrew 2012). Maximum larval density above 20 was observed in the months of September and October in 2013. Prevalence of the maximum larval density of Aedes aegypti in Thanjavur city had a direct impact over the incidence of Dengue cases. In conclusion, rain fall and physicochemical characteristics exert a significant influence on mosquito breeding site selection among A. aegypti. Coconut shells were the preferred breeding site for this mosquito in our study, followed by plastic containers and tree holes. These preferences could be exploited to develop novel techniques to deter oviposition.

REFERENCES

American Public Health Association (APHA). (2006) Standard methods for the examination of water and waste water. Washington, DC: APHA.

Ananya Bar, and Andrew, J., (2012) Seasonal Prevalence of *Aedes aegypti* Larvae in Agra. Research in Zoology 2(3), 15-18.

Bentely, M.D., Day, J.F. (1989) Chemical ecology and behavioral aspects of mosquito oviposition. Annu Rev Entomol 34; 401-21.

Bhaskar, B., Rao, Harikumar, PS., Jayakrishnan, T., and George, B. (2011) Characteristics of *aedes (stegomyia) Albopictus* skuse (diptera : culicidae) Breeding sites. Southeast Asian J Trop Med Public Health. Vol 42 No. 5.

Delatte, H., Paupy, C., Dehecq, J.S., Thiria, J., Failloux, A.B., Fontenille, D. (2008) *Aedes albopictus*, vector of chikungunya and dengue viruses in Reunion Island: biology and control. Parasite 15: 3-13 (in French).

Gubler, D.J. (1977) Studies on the behavior of *Aedes* (*Stegomyia*) albopictus and *Aedes* (*Stegomyia*) polynesiensis Marks. J Med Entomol 8: 675-82.

Gubler, D.J., Rieler, P., Cbr, K.L., Yap, W., Nasci, R., Ratz, J.A. (2001) Climate variability and change in the US potential impact on vector and rodent borne diseases. Environ Perspect 109 (suppl 2): 223-33.

Hawley, W.A.I.(1988) Biology of *Aedes albopictus*. J Am Mosq Control Assoc Suppl 1: 1-39.

Household disposables as breeding habitats of dengue vectors: Linking wastes and public health.

Kalra, N.L., Kaul, S.M., Rastogi, R.M. (1997) Prevalence of *Aedes aegypti* and *Aedes* albopictus- Vectors of Dengue and Dengue haemorrhagic fever in North, North-East and Central India. Dengue Bulletin, vol. 21.

Katyal, R., Kumar, K., Singh Gill, K., and Sharma, R.S., (2003) Impact of Intervention Measures on DF/DHF Cases and *Aedes aegypti* Indices in Delhi, India: An Update, 2001". Dengue Bulletin, vol. 27, pp. 163-167.

Medlock, J.M., Avenell, D., Barrass, I., Leach, S. (2006) Analysis of the potential for survival and seasonal activity of *Ae.albopictus* (Diptera: Culicidae) in UK. J Vector Ecol 31: 292-304.

Mutero, C.M., Nga'ang'a, P.N., Wekoyela, P., Githure, J., Konradsen, F. (2004) Ammonium sulphate fertilizer increases larval populations of *Anopheles arabiensis* and culicine mosquitoes in rice fields. Acta Trop. 89: 187-192.

Navarro, D.M.A.F., De Oliveira, P.E.S., Potting, R.P.J., Brito, A.C., Fital, S.J.F., Goulart Sant'Ana, A.E. (2003) The potential attractant or repellent effects of different water types on oviposition in Ae. aegypti L (Dipt., Culicidae). J Appl Entomol 127: 46-50.

O'Gower, A.K. (1963) Environmental stimuli and oviposition behavior of *Ae. aegypti* var. *queenslandis* Theobald (Diptrera: Culicidae). Anim Behav 11: 189-97.

Oleymi, I.K., Omalu, I.C.J., Famotele, O.I., Shegna, S.P., Idris, B. (2010) Distribution of mosquito larvae in relation to physicochemical characteristics of breeding habitats in Minna, North Central Nigeria. Rev Infec 1: 49-53.

Oyewole, I.O., Momoh, O.O., Anyasor, G.N. (2009) Physico-chemical characteristics of *Anopheles* breeding sites: Impact on fecundity and progeny development. Afr J Envirol Sci Technol 3: 447-52.

Pal, R. (1945) On the bionomics of *Anopheles culicifacies* Giles, Pt II: the ecology of the immature stages. J. Malaria Insti. India 6: 53-74.

Pandya, G. (1982) "Prevalence of dengue infection in India", Defence Science Journal, vol. 32, pp.359-370.

Pemola Devi, N., Jauhari, R.K. (2005) Species diversity patterns among mosquitoes (Diptera: Culicidae) from certain parts in Garhwal Himalayas, India. *Journal of Applied Bioscience*. 31:105–113.

Pramanik, M.K., Aditya, G., and Raut, S.K. (2007) Seasonal prevalence of *Aedes aegypti* immature in Kolkata, India, South-East Asian Journal of Trop Med Public Health vol. 38, pp. 442-447.

R. S. Sharma, R.S., Kaul, S.M., Jotna Sokhay. (2005) Seasonal fluctuations of dengue fever vector, *Aedes aegypti* (Diptera: Culicidae) in Delhi, India, The Southeast Asian journal of tropical medicine and public health, vol. 36, no.1, pp.186-190.

Rao, B.B., Harikumar, P.S., Jayakrishnan, T., George, B. (2011) Characteristics of *Aedes (Stegomyia) albopictus* Skuse (Diptera : Culicidae) breeding sites. Southeast Asian J Trop Med Public Health 42: 1077–82.

Rao, T.R. (1967) Distribution, Density and Seasonal Prevalence of *Aedes aegypti* in the Indian Sub-continent and South-East Asia. Bulletin of World Health Organization vol. 36, pp. 547-551.

Reji Gopalakrishnan, Momi Das, Indra Baruah, Vijay Veer and Prafulla Dutta. (2013) Physicochemical characteristics of habitats in relation to the density of container-breeding mosquitoes in Asom, India. J Vector Borne Dis 50, pp. 215–219.

Seghal, S.S., Pillai, M.K.K. (1970) Preliminary studies on the chemical nature of mosquito breeding waters in Delhi. Bull WHO 42: 647-50.

Service, M.W. (1995) Mosquitoes (Culicidae). In: Lane RP, Crosskey RW editors. Medical Insects and Arachnids. London, UK: Chapman & Hall. p. 120–240.

Sharma, K.R., Sreenivasarajan, T., Rao, A.N. (2008) Oviposition response of *Aedes aegypti* and *Ae. albopictus* to certain fatty acid esters. Parasitol Res 103: 1065-73.

Sharma, R.S. (1997) Seasonal prevalence of *Anopheles culacifacies, Anopheles stephensi, Culex quinquefasciatus* and *Aedes aegypti* (Diptera: Culicidae) in urban Gurgaon, Proceedings of the second symposium on vectors and vector borne diseases 227-231.

Soumyajit Banerjee, Gautam Aditya, Goutam K Saha. (2012) Household disposables as breeding habitats of dengue vectors: Linking wastes and public health. Waste Management. 10/2012; DOI:10.1016/j.

Straetemans, M. (2008) Vector related risk mapping of the introduction and establishment of *Ae. albopictus* in Europe. Euro Surveill 13: 14.

Thangamathi, P., Ananth, S., and Jenifer, P. (2011) Physico-chemical characteristics of Malarial mosquito *Anopheles stephensi* habitats in relation to density at Kumbakonam, Tamil Nadu, India. Journal of Modern Science. 3(2) 37-40.

Umar, A., Don Pedro, K.N. (2008) The effects of pH on the larvae of *Ae. aegypti* and *Cx. quinquefasciatus*. Int J Pure Appl Sci 2: 58-62.

WHO. (2012) Global strategy for dengue prevention and control 2012-2020. World Health Organization, Geneva pp. 1-34.